SFTSV RNA at $2.4 \times 10^5$ copies/mL in his semen that day. On day 44, we could no longer detect semen SFTSV RNA, and he was discharged on day 51 after onset (Figure 1).

In this study, SFTSV RNA was detected in semen, and SFTSV persisted longer in semen than in serum. It is well known that some viruses, such as Zika virus and Ebola virus, can be sexually transmitted; these viruses have been detected in semen for a prolonged period after symptom onset (6,7). Thus, we considered the potential risk for sexual transmission of SFTSV.

Compared with that of Zika and Ebola viruses, the clinical significance of potential sexual transmission of SFTSV is unknown. However, this possibility should be taken into consideration in sexually active patients with SFTSV. Our findings suggest the need for further studies of the genital fluid of SFTS patients, women as well as men, and counseling regarding sexual behavior for these patients.

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References

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Canine Distemper Virus in Asiatic Lions of Gujarat State, India


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DOI: https://doi.org/10.3201/eid2511.190120

In September 2018, an epizootic infection caused by canine distemper virus emerged in an Asiatic lion population in India. We detected the virus in samples from 68 lions and 6 leopards by reverse transcription PCR. Whole-genome sequencing analysis demonstrated the virus strain is similar to the proposed India-1/Asia-5 strain.

Canine distemper virus (CDV; genus *Morbillivirus*) causes highly contagious disease in a wide range of carnivores. Epizootic disease in lions in a wildlife sanctuary in California, USA, in 1992 and Serengeti National Park, Tanzania, in 1994 underlined the potential of CDV to cause fatality in wild felids (1,2). The disease often manifests as respiratory and gastrointestinal signs that progress to neurologic disease (2).

A single isolated population of Asiatic lions (*Panthera leo persica*) resides in the Gir forests of Gujarat State, India, the last natural habitat for this species. Conservation efforts brought this lion population back from the brink of extinction and increased their numbers (3).

During 2 weeks in September 2018, the unusual death of 28 lions of all age groups was reported from Gir Wildlife Sanctuary. A detailed investigation revealed 18 additional lions exhibited dullness, dehydration, lacrimation, cough, diarrhea, and seizures. Necropsy of 2 carcasses showed edema and purulent exudates in the lungs. Histopathology of lungs from both lions showed mononuclear cell infiltration with mild thickening of interalveolar septa.

The Indian Council of Medical Research, National Institute of Virology (Pune, India), received ocular,
nasal, and rectal swab specimens packed in a viral transport medium and blood samples from 229 wild and 87 captive lions and visceral organs, including lung, liver, heart, and kidney, from 3 dead lions for virologic investigation. Of the 229 wild lions, 20 showed clinical signs, including dullness, lacrimation, cough, diarrhea, and seizures; 2 of the 87 captive lions showed lacrimation and respiratory distress.

We extracted RNA by using Magmax Total RNA Isolation Kit (ThermoFisher Scientific, https://www.thermofisher.com) and processed samples for heminested PCR for CDV and nested PCR for *Paramyxovirus*, as described previously (4). We obtained 287 bp by heminested reverse transcription PCR (RT-PCR) and 500 bp by *Paramyxovirus* nested RT-PCR. We detected CDV from ≥1 sample from 68 (21.3%) lions, including 56 (24.5%) wild and 12 (13.8%) captive lions. All 22 of the lions with clinical signs were PCR-positive for CDV. Among the samples tested, 18/90 (20%) blood, 26/131 (21.4%) nasal, and 10/132 (7.5%) ocular specimens, as well as the visceral organs from the 3 dead lions, were CDV positive.

We performed RNA library preparation and quantification for next-generation sequencing by using methods described previously (5) and analyzed reads by using CLC Genomics Workbench version 11.0.1 (QIAGEN, https://www.qiagen.com). We retrieved the near-complete genome (15 kb) of CDV from 11 lion samples by using a combination of de novo assembly and reference mapping. We calculated percent nucleotide differences by using the p-distance method in MEGA version 7.0 (6). We performed similarity and divergence calculations for the hemagglutinin (H) gene and generated a neighbor-joining tree for the complete genome and H gene region. We performed bootstrap replication of 1,000 cycles to assess the robustness of the tree.

Phylogenetic analysis showed that sequences from this outbreak clustered with East-African CDV strains (Appendix Figure 1, http://wwwnc.cdc.gov/EID/article/25/11/19-0120-Techapp1.pdf). The complete genome of CDV among the positive samples displayed 100% similarity, except for the spleen and lung of 1 lion, which were 99.9% similar. We noted a 3.4% nucleotide difference between the complete genome of Gir CDV and the East-African strain (Table 1).

Phylogenetic analysis of the CDV sequences of Gir lions against available partial CDV sequences of H gene previously collected from tigers and a dog from India demonstrated a distinct cluster for sequences from India (Appendix Figure 2). Bhatt et al. (7) hypothesized a novel CDV strain, India-1/Asia-5, among domestic dogs in India. Our analysis of sequences from the CDV outbreak in Gir lions strengthened that hypothesis. We observed 95.8%–96.8% nucleotide similarity for the H gene region (Table 1) of CDV sequences from India (Appendix Figure 2) and previously collected from tigers and a dog from India demonstrated a distinct cluster for sequences from India (Appendix Figure 2).

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<th>GenBank accession no.</th>
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<th>Year</th>
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*Reference sample is GenBank accession no. MK037469. NA, not available. GenBank accession nos. MK037459–68 represent sequence data from this study.
Gir CDV outbreak strains and Asia 3 strains and maximum similarity with Rockborne-like strains. Whole-genome sequence analysis showed that Gir CDV strains had ≥8% nucleotide difference from the 2 known vaccine-derived strains of America 1 genotypes (Table 1).

After we confirmed CDV in Asiatic lions, samples were collected from other wild animals in Gir Wildlife Sanctuary. We detected CDV in 6/52 (12%) samples from Panthera pardus leopards but not in samples from Vivericula indica civets, Panthera tigris tigers, Canis lupus pallas wolves, or Caracal caracal cats.

Before 2019, few instances of CDV were reported in lions, tigers, red pandas, and leopards from zoos and forests in India. However, canine distemper is prevalent among dogs in India (8), and the free-ranging dog population often poses a threat of CDV transmission to wildlife (9). Other wildlife species also could play a role in maintenance and transmission of CDV (9). Vaccination is an option; attenuated Onderstepoort strain was used successfully in captive African lions in Maasai Mara National Reserve in Kenya, and a live attenuated canine vaccine was used in a vaccine trial in tigers (10). Reintegration of the existing lion population from the Gir region to different sanctuaries can ensure the protection and conservation of the species.

Acknowledgments
The authors thank Dimpal Nyayanit, Shilpi Jain, Pravin Kore, Anish Srivastava, Savita Patil, Triparna Majumdar, Swapnil Patil, Rajen Lakra, Prasad Sarkale, Deepak Suryawanshi, and Manoj Kadam for technical support.

Disclaimer: Indian Council of Medical Research (ICMR), New Delhi, India provided funding. ICMR has no role in study design, data collection, or interpretation.

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References

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Appendix

Figure 1. Phylogenetic analysis of the complete genome of canine distemper virus from samples collected from Asiatic lions, Gir National Forest, India, 2018. A maximum-likelihood method along with GTR + gamma + I model was used to generate the tree. Different colors represent different lineages of canine distemper virus. Red text denotes sequences from the reported outbreak. Pink text represents sequences reported from other outbreaks in India. Scale bar represents nucleotide substitutions per site.
Appendix Figure 2. Phylogenetic analysis of the H gene from canine distemper virus from samples collected from Asiatic lions, Gir National Forest, India, 2018. A maximum-likelihood method along with Tamura 3 parameter + gamma model was used to generate the 688 bp tree. Scale bar represents nucleotide substitutions per site.