

- models. *Hum Vaccin Immunother*. 2016;12:1989–96. <http://dx.doi.org/10.1080/21645515.2016.1199309>
6. Fagerlin A, Valley TS, Scherer AM, Knaus M, Das E, Zikmund-Fisher BJ. Communicating infectious disease prevalence through graphics: results from an international survey. *Vaccine*. 2017;35:4041–7. <http://dx.doi.org/10.1016/j.vaccine.2017.05.048>
  7. Han PKJ, Zikmund-Fisher BJ, Duarte CW, Knaus M, Black A, Scherer AM, et al. Communication of scientific uncertainty about a novel pandemic health threat: ambiguity aversion and its mechanisms. *J Health Commun*. 2018;23:435–44. <http://dx.doi.org/10.1080/10810730.2018.1461961>
  8. Scherer AM, Knaus M, Zikmund-Fisher BJ, Das E, Fagerlin A. Effects of influenza strain label on worry and behavioral intentions. *Emerg Infect Dis*. 2017;23:1425–6. <http://dx.doi.org/10.3201/eid2308.170364>
  9. Zikmund-Fisher BJ, Scherer AM, Knaus M, Das E, Fagerlin A. Discussion of average versus extreme case severity in pandemic risk communications. *Emerg Infect Dis*. 2017;23:706–8. <http://dx.doi.org/10.3201/eid2304.161600>
  10. Champion V, Skinner C. The health belief model. In: Glanz K, Rimer BK, Viswanath K, editors. *Health behavior and health education: theory, research, and practice*, 4th ed. San Francisco: Jossey-Bass; 2008. p. 45–67.
  11. Noar SM, Benac CN, Harris MS. Does tailoring matter? Meta-analytic review of tailored print health behavior change interventions. *Psychol Bull*. 2007;133:673–93. <http://dx.doi.org/10.1037/0033-2909.133.4.673>
  12. Cappella JN. Integrating message effects and behavior change theories: organizing comments and unanswered questions. *Journal of Communication*. 2006;56(suppl1):S265–79.

Address for correspondence: Thomas S. Valley, University of Michigan, Division of Pulmonary and Critical Care Medicine, 2800 Plymouth Rd, Bldg 16-G028W, Ann Arbor, MI 48109, USA; email: valleyt@umich.edu

## Peripheral Plasma and Semen Cytokine Response to Zika Virus in Humans

Jean-Michel Mansuy,<sup>1</sup> Hicham El Costa,<sup>1</sup> Jordi Gouilly, Catherine Mengelle, Christophe Pasquier, Guillaume Martin-Blondel, Jacques Izopet, Nabila Jabrane-Ferrat

Author affiliations: Centre Hospitalier Universitaire de Toulouse, Toulouse, France (J.-M. Mansuy, H. El Costa, C. Mengelle, C. Pasquier, G. Martin-Blondel, J. Izopet); INSERM U1043-CNRS UMR5282 Université Toulouse III, Toulouse (H. El Costa, J. Gouilly, G. Martin-Blondel, J. Izopet, N. Jabrane-Ferrat)

DOI: <https://doi.org/10.3201/eid2504.171886>

<sup>1</sup>These authors contributed equally to this article.

We assessed Zika virus RNA and select cytokine levels in semen, blood, and plasma samples from an infected patient in South America. Viral RNA was detected in semen >2 months after viremia clearance; cytokine profiles differed in semen and plasma. After viremia, Zika virus appears to become compartmentalized in the male reproductive tract.

Before the 2015–2016 outbreak, Zika virus infection had been associated with only mild symptoms. However, the outbreak revealed infection could cause severe clinical manifestations, particularly for fetuses and newborns (1). Furthermore, detection of replicative virus in semen and sexual transmission of the infection resulted in a paradigm shift in Zika virus virology (2,3). Several animal models were developed to study these phenomena, and studies revealed that Zika virus persistence within the male reproductive tract (MRT) results in diminished testosterone and oligospermia (4). However, because of complex ethics considerations, the consequences of infection on the MRT remain poorly understood (5).

To characterize infection in the MRT further, we conducted a longitudinal 6-month study examining Zika virus load and immunologic profile in blood, plasma, and semen in 1 man. The study patient was a 32-year-old immunocompetent white man with an asymptomatic Zika virus infection acquired in South America in January 2016; the control was a healthy 40-year-old white man without risk factors for acute or chronic infection who lived in the same area. We evaluated the concentrations of a select panel of cytokines, including innate immune mediators (interferon [IFN]- $\gamma$ , interleukin [IL]-15, IFN- $\beta$ ); inflammatory factors (IL-6, IL-18, soluble intercellular adhesion molecule 1 [sICAM-1]); chemokines (CC-motif chemokine ligand [CCL] 3, CCL-4, CXC-motif chemokine ligand [CXCL] 1, CXCL-8, CXCL-10); hematopoietic factors (granulocyte colony-stimulating factor [G-CSF], granulocyte-macrophage colony-stimulating factor); the angiogenic factor vascular endothelial growth factor A (VEGF-A); and proteases (matrix metalloproteinase [MMP]-2, MMP-9). We quantified cytokines using ProcartaPlex Multiplex Assay (ebioscience, <https://www.thermofisher.com>).

At admission, the patient had moderate fever, maculopapular rash, myalgia, and arthralgia and recovered within a few days. He was HIV negative; dengue and chikungunya virus infections were ruled out using ELISA Diapro (Diagnostic Bioprobes Srl, <https://www.diapro.it>) and RealStar Dengue and Chikungunya RT-PCR Kit 2.0 (Altona Diagnostics, <https://www.altona-diagnostics.com>). The patient did not experience other genital or urinary tract infections during the study.

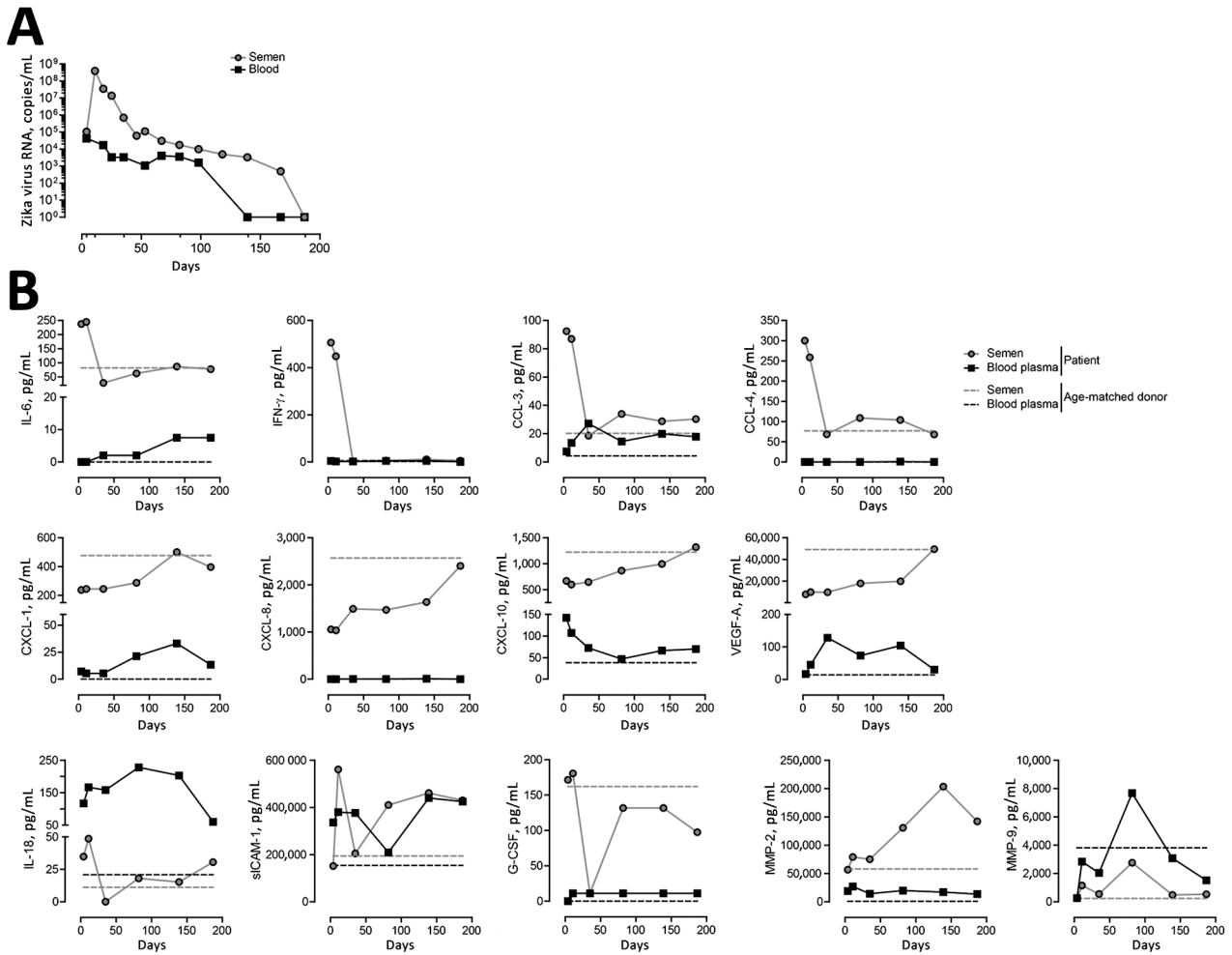
Two days after symptom onset, viral RNA was higher in semen ( $1.04 \times 10^5$  copies/mL) than in blood ( $9.4 \times 10^3$  copies/mL); RNA was detectable for up to 100 days in blood

and 168 days in semen (Figure). Soluble factors secreted in response to infection could be stratified into 3 profiles. The first profile was a surge of proinflammatory cytokines in semen (IL-6, IFN- $\gamma$ , CCL-3, CCL-4) and plasma (CCL-3, CXCL-10, VEGF-A) at early stages of infection. These cytokines returned to basal levels over time. The second profile (mainly observed in semen) involved decreased CXCL-1, CXCL-8, CXCL-10, and VEGF-A production during the first 50 days after infection, followed by a progressive return to reference ranges coinciding with viral clearance. The third profile involved sustained altered levels of factors in semen (IL-18, sICAM-1, G-CSF, MMP-2) and plasma (CCL-3, CXCL-1, sICAM-1, MMP-2, MMP-9), even after complete viral RNA clearance in plasma. Some factors were undetectable in plasma and semen samples (IL-15,

IFN- $\beta$ , granulocyte-macrophage colony-stimulating factor), and others were absent or barely detectable in plasma (IL-6, IFN- $\gamma$ , CCL-4, CXCL-8, G-CSF).

Our data revealed higher amounts of and more prolonged viral RNA shedding in semen than in blood. Zika virus infection altered cytokine secretion and induced distinct profiles in the plasma and semen; changes in the semen were more prominent. The discrepancies in kinetics and concentrations of cytokines and virus titers between semen and plasma suggest virus compartmentalization.

Cytokines produced in the MRT reflect local immune status, organ function, and tissue damage. The high levels of proinflammatory factors in semen samples from this patient illustrate an exacerbated local immune response established to control Zika virus infection. This response



**Figure.** Kinetics of Zika virus replication and cytokine secretion in semen, blood, and plasma samples from a Zika virus–infected patient, France, 2016. A) Kinetics of Zika virus RNA detection in whole blood and semen quantified by real-time reverse transcription PCR. B) Quantification of soluble factors in the blood plasma and semen at 4, 11, 35, 82, 139, and 187 days after symptom onset. Dashed lines represent baseline levels of soluble factors quantified in blood plasma and semen obtained from an age-matched healthy donor. CCL, CC-motif chemokine ligand; CXCL, CXC-motif chemokine ligand; G-CSF, granulocyte colony-stimulating factor; IFN- $\gamma$ , interferon  $\gamma$ ; IL, interleukin; MMP, matrix metalloproteinase; sICAM-1, soluble intercellular adhesion molecule 1; VEGF-A, vascular endothelial growth factor A.

might lead to recruitment of more host and inflammatory cells that further amplify viral replication and organ injury (6). Downregulation of several factors highlights the damage. For instance, the VEGF-A levels mirror the impairment of spermatogonia, primary spermatocytes, and Sertoli cells upon Zika virus infection (4). However, the decrease in CXCL-1, CXCL-8, and CXCL-10 levels in semen during infection could indicate a local immunosuppressive state induced by infection, limiting immune cell infiltration in the MRT and potentially virus dissemination throughout the body.

The different kinetics of virus replication and cytokine secretion in semen samples raises questions about the semen secretome in cases of couples aiming for conception and the necessity to extend the convalescence period beyond Zika disease recovery. In fact, at high concentration, most of these factors might alter the integrity of the mucosal barriers within the female reproductive tract and increase a woman's susceptibility to infection (7). They might also promote peroxidation and affect sperm function, potentially resulting in infertility (8).

The limitations of our study include small sample size. Further investigations with a larger cohort of patients and controls are warranted.

In summary, a profound disruption in the cytokine network is evident in plasma and semen starting at the earliest stage of Zika virus infection and is maintained over time even after viral clearance. Studies to characterize the mechanism involved in the establishment of compartmentalization and develop efficient antiviral therapies that interfere with virus replication in the MRT are needed.

#### Acknowledgment

We thank Isabelle Da Silva for technical assistance.

#### About the Author

Dr. Mansuy is a medical virologist at the University Hospital of Toulouse in Toulouse, France. His research interests include emerging viral diseases and respiratory and enteric diseases.

#### References

1. Brasil P, Pereira JP Jr, Moreira ME, Ribeiro Nogueira RM, Damasceno L, Wakimoto M, et al. Zika virus infection in pregnant women in Rio de Janeiro. *N Engl J Med*. 2016;375:2321–34. <http://dx.doi.org/10.1056/NEJMoa1602412>
2. Baud D, Musso D, Vouga M, Alves MP, Vulliamoz N. Zika virus: a new threat to human reproduction. *Am J Reprod Immunol*. 2017;77:e12614. <http://dx.doi.org/10.1111/aji.12614>
3. Mansuy JM, Suberbielle E, Chapuy-Regaud S, Mengelle C, Bujan L, Marchou B, et al. Zika virus in semen and spermatozoa. *Lancet Infect Dis*. 2016;16:1106–7. [http://dx.doi.org/10.1016/S1473-3099\(16\)30336-X](http://dx.doi.org/10.1016/S1473-3099(16)30336-X)
4. Govero J, Esakky P, Scheffer SM, Fernandez E, Drury A, Platt DJ, et al. Zika virus infection damages the testes in mice. *Nature*. 2016;540:438–42. <http://dx.doi.org/10.1038/nature20556>
5. Joguet G, Mansuy JM, Matusali G, Hamdi S, Walschaerts M, Pavili L, et al. Effect of acute Zika virus infection on sperm and virus clearance in body fluids: a prospective observational study. *Lancet Infect Dis*. 2017;17:1200–8. [http://dx.doi.org/10.1016/S1473-3099\(17\)30444-9](http://dx.doi.org/10.1016/S1473-3099(17)30444-9)
6. Shi C, Pamer EG. Monocyte recruitment during infection and inflammation. *Nat Rev Immunol*. 2011;11:762–74. <http://dx.doi.org/10.1038/nri3070>
7. Rametse CL, Olivier AJ, Masson L, Barnabas S, McKinnon LR, Ngcapu S, et al. Role of semen in altering the balance between inflammation and tolerance in the female genital tract: does it contribute to HIV risk? *Viral Immunol*. 2014;27:200–6. <http://dx.doi.org/10.1089/vim.2013.0136>
8. Fraczek M, Sanocka D, Kamieniczna M, Kurpisz M. Proinflammatory cytokines as an intermediate factor enhancing lipid sperm membrane peroxidation in vitro conditions. *J Androl*. 2008;29:85–92. <http://dx.doi.org/10.2164/jandrol.107.003319>

Address for correspondence: Jean-Michel Mansuy, Institut Fédératif de Biologie, Laboratoire de Virologie, 330 Avenue de Grande Bretagne, 31059 Toulouse CEDEX, France; email: mansuy.jm@chu-toulouse.fr

## Detection of Epizootic Hemorrhagic Disease Virus Serotype 1, Israel

Natalia Golender,<sup>1</sup> Velizar Y. Bumbarov<sup>1</sup>

Author affiliation: Kimron Veterinary Institute, Beit Dagan, Israel

DOI: <https://doi.org/10.3201/eid2504.180149>

During September 2016–February 2017, we detected epizootic hemorrhagic disease virus (EHDV) in ruminants in Israel. BLAST and phylogenetic analyses of segment 2 in 6 EHDVs isolated from field samples indicated a close relationship to the EHDV serotype 1 strain in Nigeria. Affected cattle had mostly mild or asymptomatic disease.

Epizootic hemorrhagic disease is an infectious, non-contagious viral disease of ruminants, transmitted by insects of the genus *Culicoides*; it mostly affects white-tailed deer and cattle. Epizootic hemorrhagic disease virus (EHDV) belongs to the genus *Orbivirus* within the family *Reoviridae* and is closely related to bluetongue virus (BTV) and African horse-sickness virus. At least

<sup>1</sup>Both authors share first authorship.