rabies-endemic countries, physicians should emphasize the high likelihood of transmission of rabies virus after mucosal exposure and try to persuade persons at risk to receive postexposure prophylaxis.

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Endemic Severe Fever with Thrombocytopenia Syndrome, Vietnam

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Severe fever with thrombocytopenia syndrome (SFTS), a tickborne viral disease, has been identified in China, South Korea, and Japan since 2009. We found retrospective evidence of SFTS virus (SFTSV) infection in Vietnam, which suggests that SFTSV infections also occur in Vietnam, where the virus has not been known to be endemic.

Severe fever with thrombocytopenia syndrome virus (SFTSV) is a tickborne virus (genus Phlebovirus, family Phenuiviridae) that can cause a mild to severe febrile illness similar to hemorrhagic fever (J). Phleboviruses have been found in the Americas, Asia, Africa, and the Mediterranean region. For example, Heartland virus (HRTV), another tickborne phlebovirus, was identified in northwestern Missouri, USA, in 2009 (2). Malsoor virus, a new bat phlebovirus closely related to SFTSV and HRTV, was identified in western India, and a phlebovirus similar to SFTSV and HRTV was isolated from ticks in Australia (3,4).

Severe fever with thrombocytopenia syndrome (SFTS) illness was first confirmed in China in 2009. It was retrospectively identified in South Korea in 2010 and the western regions of Japan in 2013 (1,5,6). SFTS is characterized by acute high fever, thrombocytopenia, leukopenia, elevated serum hepatic enzymes, gastrointestinal symptoms, and multiorgan failure and has a death rate of 16.2%–30% (1,6,7). Atypical signs and symptoms and asymptomatic infections also have been identified (5,8). Most SFTSV infections occur through Haemaphysalis longicornis ticks, although SFTSV transmission can also occur through close contact with an infected patient (8).

To investigate evidence of SFTSV infections in Vietnam, we collected serum samples from 80 patients with acute febrile illnesses admitted to Hue University Hospital (Hue, Vietnam)
during October 1, 2017–March 31, 2018. The Institutional Review Board of Hue University Hospital approved the study.

For the molecular diagnosis of SFTSV, we extracted RNA from stored patient serum using a QIAamp Viral RNA Mini Kit (QIAGEN, https://www.qiagen.com) and performed real-time reverse transcription PCR (rRT-PCR) to amplify the partial small (S) segment of the viral RNA from the stored serum and confirm SFTSV infection (9). rRT-PCR showed 2 positive results, from the stored serum of 2 patients with thrombocytopenia who had been seen at Hue University Hospital during 2017 and who had no history of travel to SFTSV-endemic countries, such as China, South Korea, and Japan. We also detected IgM in the serum of 1 of these patients (Appendix Table, https://wwwnc.cdc.gov/EID/article/25/5/18-1463-App1.pdf) (8).

On October 29, 2017, a 29-year-old woman (Hue 06-Vietnam-10-2017) was hospitalized at Hue University Hospital because of headache, vomiting, and gum bleeding. She lived in Hue City and was unaware of having received an insect bite. Her temperature was 38°C, and blood tests showed leukopenia (leukocyte count 1,900 cells/µL [reference 4,000–10,000 cells/µL]), thrombocytopenia (platelet count 125 × 10^9/µL [reference 150–450 × 10^9/µL]), and a low hematocrit level (34.3% [reference 36%–44%]). The patient fully recovered without other complications after 5 days.

On November 2, 2017, a 27-year-old man (Hue 13-Vietnam-11-2017) was hospitalized at Hue University Hospital because of headache and fatigue. He had had dengue fever at 8 years of age. Blood tests showed thrombocytopenia (platelet count 14 × 10^9/µL), normal leukocyte count (7,410 cells/µL), mildly elevated aspartate aminotransferase (84 IU/L [reference 8–38 IU/L]), elevated alanine aminotransferase (98 IU/L [reference 4–44 IU/L]), and mildly elevated hematocrit (47.6% [reference 36%–44%]). He fully recovered without other complications after 7 days.

We sequenced rRT-PCR products from the stored serum samples using a BigDye Terminator Cycle Sequencing kit (Applied Biosystems, http://www.thermofisher.com). We performed phylogenetic analysis of the partial S segment sequences with MEGA6 (https://www.megasoftware.net) and constructed phylogenetic trees using the maximum-likelihood method, which confirmed SFTSV infection (Appendix Figure).

We confirmed 2 SFTSV infections in Hue in 2017 by amplifying the partial S segment of the viral RNA in stored serum from patients with thrombocytopenia; elevated levels of serum hepatic enzymes, including aspartate aminotransferase and alanine aminotransferase; and gastrointestinal symptoms, such as vomiting. The signs and symptoms were milder than the major signs and symptoms of SFTS, which has a high death rate.

**References**


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**About the Author**

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Tuberculosis in elephants is primarily caused by *Mycobacterium tuberculosis*. We identified mixed *M. tuberculosis* lineage infection in 2 captive elephants in Nepal by using spoligotyping and large sequence polymorphism. One elephant was infected with Indo-Oceanic and East African–Indian (CAS-Delhi) lineages; the other was infected with Indo-Oceanic and East Asian (Beijing) lineages.

*Mycobacterium tuberculosis* is a primary cause of tuberculosis (TB) in elephants (1). Culture of trunk wash samples is regarded as the standard method for the diagnosis of TB in elephants; however, this method has many limitations (2). We previously reported TB in 3 elephants in Nepal that was caused by *M. tuberculosis* of Indo-Oceanic lineage (3). Here, we report on mixed *M. tuberculosis* lineage infection in 2 captive elephants from Chitwan National Park (CNP) in Nepal.

Elephant A was a female elephant ≈65–70 years old. She had been in retirement for 3 years before she died in February 2013. We observed TB-like lesions in the lungs postmortem (Appendix Figure 1, https://wwwnc.cdc.gov/EID/article/25/5/18-1898-App1.pdf). Elephant B was a 32-year-old male. His body condition had substantially deteriorated for several months before he died. We found extensive TB-like lesions in the lungs at postmortem.

We performed the DPP VetTB Assay (Chembio Inc., http://chembio.com), a serologic test, on the postmortem wash samples is regarded as the standard method for the diagnosis of TB in elephants; however, this method has many limitations (2). We previously reported TB in 3 elephants in Nepal that was caused by *M. tuberculosis* of Indo-Oceanic lineage (3). Here, we report on mixed *M. tuberculosis* lineage infection in 2 captive elephants from Chitwan National Park (CNP) in Nepal.

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We performed the DPP VetTB Assay (Chembio Inc., http://chembio.com), a serologic test, on the postmortem wash samples. We confirmed the presence of antibodies to TB by using Löwenstein–Jensen media. We performed genetic analyses on the 2 *M. tuberculosis* isolates by using spoligotyping and large-sequence polymorphism (LSP) as described previously (5). We amplified the direct-repeat region with a primer pair and hybridized the PCR products to a set of 43 oligonucleotide probes corresponding to each spacer covalently bound to the membrane. We identified the spoligo-international type by comparing spoligotypes with the international spoligotyping database (SpolDB4) (6). We performed LSP on the isolates by using specific primers for respective lineages, as described previously (7).

We identified the elephant isolates as a mixture of 2 strains based on uneven spoligotyping color development (suggesting mixture) and LSP detection PCR results (2 bands were observed). The spoligotyping results showed that the elephant A isolate had a new spoligotype that was not found in the international spoligotyping database. The elephant B isolate belonged to the Indo-Oceanic lineage (East African–Indian 5 spoligointernational type 1365) (Table). The prevalence of the Indo-Oceanic lineage among human TB patients in Nepal is only 11.5% (8). The drug...