Increase in Enterovirus D68 Infections in Young Children, United Kingdom, 2006–2016

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We determined the change in seroprevalence of enterovirus D68 (EV-D68) in the United Kingdom in age-stratified cohorts from 2006 to 2016, the period during which EV-D68 emerged as a cause of severe respiratory disease occasionally leading to paralysis. Infections were acquired primarily in infants and young children, and incidence was markedly higher in 2016.

Enterovirus D68 (EV-D68) is a member of the Enterovirus D species (genus Enterovirus; family Picornaviridae). EV-D68 is distributed worldwide and is typically associated with upper respiratory tract infections. Although EV-D68 infections were infrequently reported in the United States and elsewhere before 2010 (1,2), multiple novel clades of EV-D68 have emerged worldwide (3) and have been associated with occasional outbreaks of more severe respiratory infections (4). However, it was not until 2014 that a series of large-scale EV-D68 outbreaks resulting in severe illness and death were reported from the United States and Canada and subsequently, in 2016, from Europe (Netherlands, Spain, Germany, Italy, Ireland, Austria, France, Luxembourg), Japan, China, and elsewhere in Asia (4–6).

While primarily regarded as a respiratory pathogen, EV-D68 has been occasionally associated with acute flaccid myelitis (AFM) (7). This apparent change in tropism for cells in the central nervous system (8) may be linked to the emergence of novel genetically distinct EV-D68 lineages (3). An alternative possibility is that the increased number of reports of severe, AFM-associated infections with EV-D68 reflects larger-scale changes in population immunity that have enabled outbreaks to occur in potentially vulnerable age groups. Severe infections typically target infants >6 months of age, when maternal antibody protection wanes (4). The World Health Organization recently identified EV-D68 as a potential major public health risk and recommends enhanced surveillance and more effective diagnostics (9). We investigated potential changes in exposure to EV-D68 in the general population of the United Kingdom over the period in which the worldwide outbreaks of EV-D68 occurred.

The Study

For this study, we obtained serum samples collected as an approximately representative age-stratified cross section of the UK population (10). These samples were collected in 2006, before the reports of increased number of EV-D68 cases (n = 516), and in 2016, after the 2014 EV-D68 outbreak (n = 566) (Figure 1, panel A). We used a standard microneutralization assay for serum samples (Appendix, https://wwwnc.cdc.gov/EID/article/25/6/18-1759-App1.pdf).

To determine the optimal strain to measure neutralizing antibody titer (Nab) assays, we compared titers of selected serum samples to the prototype Fermon (1962) strain and those of more recent EV-D68 isolates isolated in 2005 (FI_2005) and 2016 (FI_2016) (Appendix Figure 1). We identified genotypes of D for FI_2005 and B3 for FI_2016 by phylogenetic comparison of the viral protein 1 sequences. We selected samples for comparing titers to narrow down times of EV-D68 exposure: patients >40 years of age in 2006, representing serologic responses to infections acquired substantially before 2006; patients 6 months–5 years of age in 2006, representing responses to infections acquired during 2001–2006; and patients 6 months–5 years of age in 2016, representing responses to infections acquired during 2011–2016. Geometric mean titers (GMTs) of NABts to Fermon and FI_2016 were comparable between exposure groups (Appendix Figure 2), whereas samples collected from children infected during 2001–2006 showed some evidence for proportionately higher seroreactivity to the 2005 strain. Overall, differences in GMTs were minor, and we selected the FI_2016 strain for Nab screening.

We determined seroreactivity to EV-D68 by GMT calculations for each age and year category and proportions of samples with different neutralizing antibody titers (Figure 1, panel A). We determined seroprevalence and inferred frequencies of past infection using a conservative 1:16 titer threshold (Figure 1, panel B). Frequencies and titers of
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EV-D68 NAbss differed substantially between the 2 collection years in young children (in the categories 0.5–1 year, 1–5 years, and 6–10 years of age). The difference narrowed in older age groups, and seroprevalence approached 100% in those >40 years of age. Seroprevalence and titer distributions were elevated in the <0.5-year age group, likely reflecting the presence of maternal antibody in these infants.

The differences in seroprevalence in the young children between 2006 and 2016 demonstrate greater infection rates in 2016. To identify the age at which this greater exposure occurred, we divided sample sets into narrower age bands and determined seroprevalence (Figure 2, panel A). For both groups, infections were acquired at a very early age, with extremely high incidences in the 0.5–2-year and 3–4-year age ranges in both sample years but a marked reduction for children >5 years of age. Annualized incidence of EV-D68 infection in the 0.5–5-year age band increased from 36 infections/1,000 population during 2001–2005 to 53 infections/1,000 population during 2012–2016, an increase of 50% (Figure 2, panel B). The increased incidence in the <5-year age group in our study would equate to >35,000 additional EV-D68 infections/year, primarily in young children 0.5–2 years of age. Incidence in older age groups was comparable or reduced, as we expected with greater rates of EV-D68 exposure and seroconversion in the younger age ranges.

Conclusions

The seroprevalence of EV-D68 approached 100% in adult UK populations in this study, consistent with previous seroepidemiology studies conducted in Finland and China (11,12). Our findings provide further evidence for the wide circulation of EV-D68 infections before reported outbreaks in 2014 and 2016. We demonstrated high incidences of EV-D68 infection in young children; around one half are already infected by 2 years of age in both sampling periods. However, the seroprevalence of EV-D68 infection was consistently higher throughout childhood in 2016 samples than in 2006 samples (Figure 1).

Changes in population immunity, virus antigenicity, transmissibility, cellular tropism, and pathogenicity may contribute to the recent upsurge of severe EV-D68 infections worldwide. Concerning the first potential explanation, seroprevalences in age groups <40 years were consistently lower in 2006 than 2016 (Figure 1, panel B), although EV-D68 circulated extensively, and adults were almost always seropositive and immune before and after periods of greater disease severity in the United Kingdom and elsewhere (4,13). Changes in NAb susceptibility and escape from population immunity appear unlikely to be a cause of changes in incidence; we found no evidence for major differences in antigenicity between Fermon, genotype D, or B3 isolates (Appendix; Appendix Figure 1). Another possible cause is increased transmissibility of subgenotypes B2 and B3 in younger age groups (Figure 2, panel A), reflecting a possible change in tropism or persistence of virus shedding and longer periods of infectivity after changes in receptor use, cellular tropisms, or both. The Fermon prototype strain of EV-D68 isolated in 1962 has a stronger affinity to α2,6-sialic acid primarily in the airways than to α2,3-linked sialic acid present in the airways.
lower respiratory tract (14). Whether the newer EV-D68 strains causing an outbreak of severe respiratory infections in 2008–2010 were evolved to use primarily α2,3-linked sialic acid or have switched to a sialic acid-independent mechanism of virus entry (14), such as ICAM5 (15), needs further investigation. Of note, only contemporary EV-D68 strains were able to infect neuroblastoma-derived neuronal cell line SH-SY5Y and cause paralysis in a mouse model, indicating that additional changes in virus strains might have occurred around or after 2012 (8), although the relationship of this finding to increased transmissibility of EV-D68 has not been studied.

In summary, we document the greater circulation and force of EV-D68 infection in infants and young children in the United Kingdom over the period in which EV-D68 has emerged as an important respiratory pathogen and potential cause of paralytic disease, reflecting its changed transmissibility and pathogenic potential. Such rapid changes in virus behavior emphasize the importance of ongoing surveillance and appropriate diagnostics for emerging enteroviruses in the future.

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Appendix

Neutralization Assay

Aliquots of serum samples were inactivated at +56°C for 30 min before assaying for neutralizing activity. Replicate serial 2-fold dilutions of samples were mixed with an equal volume of virus (100 TCID$_{50}$) in a volume of 100 µL to produce a dilution range from 1:8 to 1:1024 and incubated at +37°C for 1 hr. Subsequently, 100 µL of medium containing RD (human epithelial lung) cells was added and incubated at +37°C for 5 days. Cells were then scored for cytopathic effect (CPE) indicating the presence of non-neutralized virus. Each run included the following controls:

a) Toxicity control. A 1/8 dilution of each test sample was incubated with cells and no virus to ensure than any observed CPE in the titration was virally induced.

b) Back-titration of virus infectivity, to demonstrate that $\approx$100 TCID$_{50}$ has been used in each well.

c) Neutralization susceptibility, titration of virus with the ATCC D68 (ATCC VR-1826) control antiserum, where titers of between 1/2048 and 1/4096 were observed.

Titers were recorded as the highest dilution preventing virus replication. In situations where 1 replicate was positive and 1 was negative, titers were recorded as an intermediate titer (half-log2) dilution. Samples with no CPE at either 1/8 dilution were scored as 1/4; those showing neutralization in both 1/1024 dilutions were scored as 1/2048. For analysis purposes, a conservative threshold of 1/16 was used to assign samples as seropositive and seronegative because samples lacked a clear differentiation into negative and positive titer ranges. The 1/16
titer threshold corresponds to the 1/8 titers reported in (12) because the latter did not account for the 1:2 dilution effect after adding virus to the antibody dilution.

**Sequencing**

Viral RNA was extracted from culture supernatants using QIAamp Viral RNA mini kit (cat. 1020953; QIAGEN, www.qiagen.com). A fragment of VP1 gene (750 bp) was amplified for species and type assignment. Extracted RNA was reverse transcribed using random primers and Superscript III (all, Invitrogen, www.thermofisher.com) and then amplified in PCR reactions using Taq 2X Master Mix (NEB, https://www.neb.com) with primers: 5’-GTNACMTGTTTYATGCARACMAACCT-3’ (Forward) and 5’-AATGCWAATGTMGGNTATGTNACMTG-3’ (Reverse). The resulting sense and antisense sequences were assembled to contigs using Sequencher v5.0 (Gene Codes, https://www.genecodes.com).

**Accession Numbers**

Assembled complete genome sequences of the FI_2005 and FI_2016 used in the study have been submitted to GenBank and have been assigned accession numbers MK216564 and MK216565.
Appendix Figure 1. Phylogenetic analysis of Fermon, FI_2005, and FI_2016 EV-D68 strains used in neutralization assays. Sequences were assembled from Illumina libraries as previously described in Sun et al. (1), aligned, and analyzed using SSE platform v1.3 (2). Sequences were compared in the VP1 region with published reference sequences; selection was based upon possession of complete VP1 sequences and divergence of <1% from other sequences in this region. Maximum likelihood phylogenetic trees using an optimal substitution model (Tamura 3 parameter model with gamma distribution and allowing for invariant sites as determined by model testing) were constructed using Molecular Evolutionary Genetics Analysis (MEGA) 6 software (3). Trees were bootstrap resampled 100 times to assess the robustness of the branches.
Appendix Figure 2. Geometric mean titers of selected samples from a representative sample of UK residents with 3 strains of enterovirus, representing early (pre-2006), intermediate (2001–2006), and late (2011–2016) exposure. Each group contained 16 samples.

References

