Molecular Genotyping of Hepatitis A Virus, California, USA, 2017–2018

William S. Probert, Carlos Gonzalez, Alex Espinosa, Jill K. Hacker
Author affiliation: California Department of Public Health, Richmond, California, USA
DOI: https://doi.org/10.3201/eid2508.181489

We implemented subgenomic and whole-genome sequencing to support the investigation of a large hepatitis A virus outbreak among persons experiencing homelessness, users of illicit drugs, or both in California, USA, during 2017–2018. Genotyping data helped confirm case-patients, track chains of transmission, and monitor the effectiveness of public health control measures.

The United States has seen a resurgence of hepatitis A virus (HAV) infections; several states have reported outbreaks of HAV subgenotype IB, primarily among persons experiencing homelessness and users of illicit drugs (1). In California, during November 2016–May 2018, an outbreak of HAV IB infections resulted in 708 case-patients, 465 hospitalizations, and 21 deaths (1; https://www.cdph.ca.gov/Programs/CID/DCDC/CDPH%20Document%20Library/Immunization/2016-18CAOutbreakAssociatedDrugUse-Homelessness.pdf). To better respond to the surge in hepatitis A cases and facilitate vaccine acquisition and distribution, California declared a public health emergency in October 2017 (2). As part of this response, the California Department of Public Health implemented molecular genotyping of HAV to support epidemiologic investigation of suspect cases.

We requested serum samples for symptomatic, HAV IgM-positive case-patients from local public health jurisdictions for genotyping. We amplified a segment of the HAV viral protein 1–amino terminus of 2B (VP1–P2B) genomic region by using nested reverse-transcription PCR and performed sequencing on 160 specimens collected during August 2017–May 2018 (3; Appendix, https://wwwnc.cdc.gov/EID/article/25/8/18-1489-App1.pdf). HAV subgenotype classification by VP1–P2B sequence yielded 48 IA-positive, 109 IB-positive, and 3 II-IA-positive specimens (Figure, panel A). We identified 19 unique HAVIAVP1–P2B sequences with an overall average genetic distance of 0.043 nt substitutions per site. Eighteen (37.5%) HAV IA specimens yielded sequences that matched 2 sequences (VRD_521_2016 and RIVM-HAV16–090), previously associated with HAV outbreaks among men who have sex with men (MSM) (4,5). All 18 case-patients were male, and all but 1 identified as MSM.

We identified 11 unique VP1–P2B sequences with an overall average genetic distance of 0.014 nt substitutions per site among the HAV IB specimens. Phylogenetic analysis indicated 2 distinct HAV IB clusters: 1 cluster of 9 closely related sequences (CA IB cluster) identified from 107 California patients, and 1 cluster of 2 specimens matching sequences associated with a concomitant outbreak in Michigan (MI IB cluster) (Figure). Both case-patients with MI IB strains reported traveling to Michigan during the probable period of exposure. A search of GenBank and Hepatitis A Laboratory Network databases failed to reveal any exact matches to the CA and MI IB outbreak sequences (6; https://www.rivm.nl/en/Topics/H/HAVNET). However, Hepatitis A Laboratory Network sequence similarity analysis showed that the CA IB strains were most closely related to strains found in the Middle East and the MI IB strains to strains found in East Africa (data not shown). Three specimens with unique VP1–P2B sequences were classified as subgenotype IIIA, a genotype rarely reported in the United States (7). Two of these IIIA sequences (V17S07440 and V18S00013) shared 99.1% and 99.7% sequence identity, respectively, with strains from a 2018 outbreak in Denmark associated with dates imported from Iran (S. Midgley, Statens Serum Institut, Denmark, pers. comm., email, 2018 Aug 22). Neither of those case-patients had traveled internationally or had other known HAV risk factors within their exposure period. However, 1 case-patient had consumed dates from Iran, and the other reported eating dates from a local Middle Eastern grocery store.

We processed selected specimens representing the major IB VP1–P2B sequence variants for whole-genome sequencing (Appendix). Whole-genome sequencing of HAV can provide higher resolution strain typing than sequencing of short subgenomic regions (8). We deposited nearly complete genome sequences (7,306 nt) for strains representing the CA and MI outbreaks in GenBank (accession nos. MH577308–14). Phylogenetic comparison with other IB genome sequences in GenBank confirmed that the CA and MI IB outbreak sequences represented distinct clades (Figure, panel B). The CA IB outbreak strains shared 95.5%–95.6% nt and 99.7% aa sequence identity with the HAV IB reference strain, HM175. Similarly, the MI IB outbreak strains shared 95.8% nt and 99.6% aa sequence identity with HM175.

We rapidly implemented genotyping to help guide the public health response to a surge in reported HAV infection cases in California. Paired with epidemiologic data, genotyping information was used to confirm cases, distinguish outbreak-related cases from sporadic cases, track modes and chains of transmission, and identify populations at increased risk for infection. Our study revealed
several phylogenetic clusters of HAV. A large cluster of IB strains was confirmed as the primary cause of an outbreak that was chiefly transmitted person-to-person and was associated with risk factors of homelessness and illicit drug use (1). Genetically similar strains and risk factors have since been described for outbreaks in other states (1). Partly because of these outbreaks, hepatitis A vaccination was recently recommended for persons experiencing homelessness; recommendations for vaccination of users of injection and noninjection drugs were established in 1996 (9, 10). By April 2018, implementation of public health control measures, including educational awareness and targeted vaccination and environmental remediation, reduced the number of reported HAV infection cases to baseline levels in California.

Limitations of our investigation were the paucity of archival genotyping data from California for strain comparisons and the lack of genotyping capabilities during early stages of the IB outbreak. Sustained public health laboratory capacity for HAV genotyping, along with diligent epidemiologic surveillance, offer the opportunity to detect outbreaks earlier and monitor the effectiveness of prevention and control efforts in California.

Acknowledgments
We gratefully acknowledge Sumathi Ramachandran and Guo-liang Xia for initially identifying the IB clusters and for their technical support in implementing VP1–P2B genotyping. We are also grateful for the epidemiological and data support provided by Monique Foster and Jennifer Zipprich, Rosie Glenn-Finer, Cynthia Yen, and Maria Salas.

About the Author
Dr. Probert is a senior microbiologist at the Viral and Rickettsial Disease Laboratory, California Department of Public Health. His research focuses on the development of molecular diagnostic tools for the detection of infectious agents.
Crimean-Congo Hemorrhagic Fever, Herat Province, Afghanistan, 2017

Aziz-ur-Rahman Niazi, Mohammad Jawed Jawad, Ahmad Amirnajad, Peter A. Durr, David T. Williams

Author affiliations: Herat University, Herat, Afghanistan (A.-u.-R. Niazi, M.J. Jawad); Department of Public Health, Herat (A. Amirnajad); CSIRO, Australian Animal Health Laboratory, Geelong, Victoria, Australia (P.A. Durr, D.T. Williams)

DOI: https://doi.org/10.3201/eid2508.181491

We studied the clinical and epidemiologic features of an outbreak of Crimean-Congo hemorrhagic fever in Herat Province, Afghanistan. The study comprised 63 patients hospitalized in 2017. The overall case-fatality rate was 22.2%; fatal outcome was significantly associated with a negative IgM test result, longer prothrombin time, and nausea.

Crimean-Congo hemorrhagic fever (CCHF) is a geographically widespread tickborne disease caused by the CCHF virus (genus Ortho*nairovirus, family Nairoviridae). In humans, CCHF is associated with a case-fatality rate (CFR) of 5%–50% (1) and is considered a major public health threat (2).

CCHF cases were first reported in Afghanistan in 1998; no additional cases were reported until 2007 (3). During 2007–2016, the Afghanistan Ministry of Public Health documented 478 cases, of which Herat Province accounted for 263 (55.0%) (4). In 2017, an unusual increase in CCHF cases occurred in Afghanistan, mostly in Herat Province (Appendix Figures 1, 2, https://wwwnc.cdc.gov/EID/article/25/8/18-1491-App1.pdf). We analyzed the clinical and epidemiologic features of this outbreak.

A descriptive case series study at Herat Regional Hospital during January–December 2017 was undertaken. Clinical and epidemiologic features of all confirmed and probable CCHF cases were recorded. The Human Ethics Committee of Herat University approved the study protocol (approval #0317).

The first recorded case in this study occurred in a 90-year-old male farmer who visited Herat Regional Hospital on May 5. Later, more patients sought care for acute febrile syndrome matching the World Health Organization CCHF case definition (5). A total of 64 patients sought care for CCHF signs and symptoms over a 6-month period, of whom 1 did not consent to hospitalization and left the hospital without medical consultation.
Appendix

Nested RT-PCR Amplification and Sequencing of the VP1-P2B Region of Hepatitis A Virus

Total nucleic acids were extracted and purified from 250 µL of serum using the NucliSENS easyMag system (bioMérieux, Marcy-l'Étoile, France). Nested RT-PCR was performed using the external oligonucleotide primers 2870 and 3381, and the internal oligonucleotide primers 2897 and 3288 (1). Twenty microliters of total nucleic acids were reverse transcribed and amplified using the OneStep RT-PCR kit (QIAGEN Inc., Valencia, CA) and the oligonucleotide primers 2870 and 3381 at 300 nM in a total reaction volume of 50 µL. The thermal cycling conditions included a reverse transcription step for 40 min at 50°C, an initial PCR activation step of 15 min at 95°C, followed by a 3-step cycling program of 15 s at 95°C, 20 s at 50°C, and 40 s at 72°C for 45 cycles. A second round of amplification was then performed using the PerfeCTa SYBR Green Fast mix (QuantaBio, Beverly, MA) and included 5 µL of first round amplified product and oligonucleotide primers 2897 and 3288 at 300 nM in a total reaction volume of 50 µL. Thermal cycling parameters for the second round of amplification included an initial PCR activation step of 15 min at 95°C, followed by a 3-step cycling program of 30 s at 95°C, 30 s at 55°C, and 30 s at 72°C for 40 cycles. All amplification reactions were performed with an ABI 9700 thermal cycler (Thermo Fisher Scientific, Waltham, MA). Amplified products were purified using the QIAquick PCR purification kit (QIAGEN inc.) and the purified products subjected to bidirectional dye terminator cycle sequencing using a DNA sequencing service provided by Sequetech Corporation (Mountain View, CA). DNA sequences were assembled, edited, and trimmed to 315 bp with MEGA version 5.05. Phylogenetic analysis was performed in MEGA version 5.05 using ClustalW for sequence alignment and the Neighbor-Joining algorithm and Kimura 2-parameter evolutionary model for dendrogram generation.
Whole-Genome Sequencing of Hepatitis A Virus

Total nucleic acids were extracted and purified as described above and concentrated 5-fold using RNA Clean & Concentrator-5 spin columns (Zymo Research, Irvine, CA). DNA removal was performed using the DNA-free kit (Thermo Fisher Scientific) followed by ribosomal RNA depletion using the NEBNext rRNA depletion kit (New England Biolabs, Ipswich, MA). Libraries were constructed using the NEBNext Ultra II RNA Library Preparation kit (New England Biolabs). Pooled libraries were sequenced using MiSeq Reagent Kit v2 (500-cycles) chemistry on the MiSeq instrument (Illumina, San Diego, CA). Raw FASTQ files were cleaned and contigs generated using the Viral NGS Analysis Platform (CDC, OAMD portal). Final genome assembly and editing was performed with Geneious version 10.0.9 (Biomatters Ltd, Auckland, New Zealand). Genome sequences were trimmed to 7306 nt and phylogenetic analysis was performed in MEGA version 5.05 using ClustalW for sequence alignment and the Neighbor-Joining algorithm and Kimura 2-parameter evolutionary model for dendrogram generation.

Reference