

investigation reveals highly conserved, mosaic, recombination events associated with capsular switching among invasive *Neisseria meningitidis* serogroup W sequence type (ST)-11 strains. *Genome Biol Evol.* 2016;8:2065–75. <https://doi.org/10.1093/gbe/evw122>

10. Clemence MEA, Maiden MCJ, Harrison OB. Characterization of capsule genes in non-pathogenic *Neisseria* species. *Microb Genom.* 2018;4. <https://doi.org/10.1099/mgen.0.000208>

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## Hepatitis A Virus Genotype IB Outbreak among Internally Displaced Persons, Syria

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DOI: <https://doi.org/10.3201/eid2602.190652>

In 2018, a hepatitis A virus outbreak was identified among internally displaced persons in Syria. Sequence analysis based on the viral protein 1/2A junction revealed that the causative virus belonged to genotype IB. A high displacement rate, deteriorated sanitary and health conditions, and poor water quality likely contributed to this outbreak.

**H**epatitis A virus (HAV) is the leading cause of acute hepatitis infections worldwide, infecting ≈1.5 million persons annually (1). Symptoms, which are usually mild, include nausea, vomiting, abdominal pain, restlessness, body weakness, myalgia, loss of appetite, and fever. However, HAV may progress into fulminant liver failure, necessitating liver transplant. Generally, HAV is self-limiting (2). HAV (genus *Hepatovirus*, family *Picornaviridae*) is a nonenveloped virus with a single-stranded, positive-sense

RNA linear genome (7.5 kb). The viral protein (VP) 1/2A junction (168 nt) is used to classify HAV into 6 genotypes: I–III (subgenotypes A and B) of human origin and IV–VI of simian origin (3). Genotype IA is the most commonly reported worldwide, whereas genotype IB is predominant in the Middle East (4–6).

On September 9, 2018, the governorate of Aleppo, Syria, informed the World Health Organization office in Syria that internally displaced persons (IDPs; displaced since early 2018) and local host community members in Tal Refaat, Fafin, and surrounding areas in the northwestern and western parts of Aleppo were experiencing a suspected hepatitis outbreak. The affected area included 17 locations in Azaz and Jabal Sem'an districts in western Aleppo (Appendix, <https://wwwnc.cdc.gov/EID/article/26/2/19-0652-App1.pdf>). Outbreak field investigation found sporadic cases of the disease among IDPs starting July 21, 2018; as of November 8, a total of 638 cases of suspected acute hepatitis infection had been reported. Most patients (98.59%) were <15 years of age and the rest 16–54 years of age. A total of 105 patients (16.5%) were admitted into the Fafin hospital; no fatalities were reported. No field investigations were performed in the first half of 2018 because of the crisis that led to weakness in the routine surveillance system.

A total of 48 unidentified serum and plasma samples were collected from 24 IDP children with suspected hepatitis and sent to the laboratory on October 29. The specimens originated from 3 locations in Syria: 13 from Fafin camp in Aleppo, 6 from eastern rural Daraa, and 5 samples from rural Quneitra. Even though the main outbreak was in the Aleppo governorate, Daraa and Quneitra were also experiencing a notable upsurge in reported cases of suspected acute hepatitis infection. For this reason, additional samples were collected from these governorates.

We analyzed the serum specimens by serology (total HAV antibodies and HAV IgM) using the enzyme-linked fluorescent assay VIDAS (bioMérieux Diagnostics, <https://www.biomerieux-diagnostics.com>) and the plasma specimens by real-time reverse transcription PCR (RT-PCR) for the detection of HAV (using the HAVNET protocol) and hepatitis E virus (HEV) (7). Seven samples had insufficient volume to perform both total HAV antibody and HAV IgM tests; thus, only the IgM test was performed. Overall, 19 plasma specimens were positive for HAV and none for HEV by PCR (Table). Eighteen serum specimens had detectable HAV IgM. All the specimens with sufficient volume (n = 17) were positive for total HAV antibodies. Of these, 5 were from past infections, as indicated by the negative HAV PCR and HAV IgM

**Table.** Serologic and PCR analysis of serum and plasma specimens from the suspected hepatitis outbreak\*

Lab ID	District	Serology		Molecular analysis		Genotype
		HAV IgM	HAV total	HAV vRNA	HEV vRNA	
1	Fafin	+	+	+	–	ND
2	Fafin	+	+	+	–	IB
3	Fafin	+	+	+	–	IB
4	Fafin	+	+	+	–	ND
5	Fafin	+	+	+	–	ND
6	Fafin	+	+	+	–	IB
7	Fafin	+	+	+	–	IB
8	Fafin	+	+	+	–	ND
9	Fafin	–	+	–	–	–
10	Fafin	+	+	+	–	ND
11	Fafin	+	NS	+	–	ND
12	Fafin	–	+	–	–	–
13	Fafin	–	+	–	–	–
14	Quneitra	+	+	+	–	ND
15	Quneitra	–	+	+	–	ND
16	Quneitra	–	+	–	–	–
17	Quneitra	+	+	+	–	ND
18	Quneitra	+	+	+	–	IB
19	Daraa	+	NS	+	–	ND
20	Daraa	+	NS	+	–	ND
21	Daraa	+	NS	+	–	IB
22	Daraa	+	NS	+	–	ND
23	Daraa	+	NS	+	–	ND
24	Daraa	–	NS	–	–	–

\*HAV, hepatitis A virus; HEV, hepatitis E virus; ND, not determined; NS, no sufficient volume; +, positive; –, negative.

results. One patient had detectable HAV vRNA but negative HAV IgM, which indicates the early start of acute infection (8). This patient's serum was positive for total HAV antibodies, indicating a previous exposure with a current breakthrough infection.

We successfully sequenced the VP1/2A region for 6 specimens (Table) and used ClustalW in BioEdit 7.0 to align the sequences (9). Sequence-based genotypes were inferred by comparing the obtained sequences with genotype reference and contemporary strains obtained from GenBank. The phylogenetic analysis indicated that all Syria specimens belonged to genotype IB.

The Office for the Coordination of Humanitarian Affairs reported that, as of September 3, 2018, a total of 23,279 families (107,083 persons) were displaced from Afrin to Tel Refaat, Fafin, and surrounding villages. These IDPs were in addition to 4,766 families (38,843 persons) in the host community. Tents, destroyed or empty dwellings, schools, mosques, and warehouses were used as collective shelters but are relatively distant from active frontlines. IDPs had restricted freedom of movement and no access to proper sanitation facilities as a result of infrastructure damage; 70% of the population rely on water trucking services, and 30% live on less than 20 L of water per day. Moreover, 88% of the respondents reported accumulation of solid waste in their areas (World Health Association, unpub. data). A health assessment in the Afrin district found that 153 of 180 (85%) assessed communities have no access to health services. Vaccination

campaigns have been largely suspended since November 2016. These poor living and health conditions render the IDPs highly prone to vaccine-preventable diseases, including hepatitis A, measles, polio, and cholera (10). Although water testing for HAV was not possible in the affected areas, deterioration in water quality was reported down the supply chain and may have contributed to this outbreak (World Health Association, unpub. data).

In summary, we report a large outbreak of hepatitis among IDPs in Syria. Laboratory testing confirmed current HAV IB infection among most screened patients. The high displacement rate, deteriorated sanitary and health conditions, and poor water quality may have all contributed to the increased HAV reports among this population.

#### Acknowledgments

We thank George Araj and the Clinical Microbiology Laboratory at the American University Medical Center for their help with serologic analysis.

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## References

1. Franco E, Meleleo C, Serino L, Sorbara D, Zaratti L. Hepatitis A: epidemiology and prevention in developing countries. *World J Hepatol.* 2012;4:68–73. <https://doi.org/10.4254/wjh.v4.i3.68>
2. Jeong S-H, Lee H-S. Hepatitis A: clinical manifestations and management. *Intervirology.* 2010;53:15–9. <https://doi.org/10.1159/000252779>
3. Lemon SM, Ott JJ, Van Damme P, Shouval D. Type A viral hepatitis: a summary and update on the molecular virology, epidemiology, pathogenesis and prevention. *J Hepatol.* 2017;68:167–84. <https://doi.org/10.1016/j.jhep.2017.08.034>
4. Vaughan G, Goncalves Rossi LM, Forbi JC, de Paula VS, Purdy MA, Xia G, et al. Hepatitis A virus: host interactions, molecular epidemiology and evolution. *Infect Genet Evol.* 2014;21:227–43. <https://doi.org/10.1016/j.meegid.2013.10.023>
5. Hamza H, Abd-Elshafy DN, Fayed SA, Bahgat MM, El-Esnawy NA, Abdel-Mobdy E. Detection and characterization of hepatitis A virus circulating in Egypt. *Arch Virol.* 2017;162:1921–31. <https://doi.org/10.1007/s00705-017-3294-4>
6. Yilmaz H, Karakullukcu A, Turan N, Cizmecigil UY, Yilmaz A, Ozkul AA, et al. Genotypes of hepatitis a virus in Turkey: first report and clinical profile of children infected with sub-genotypes IA and IIIA. *BMC Infect Dis.* 2017; 17:561. <https://doi.org/10.1186/s12879-017-2667-3>
7. Jothikumar N, Cromeans TL, Robertson BH, Meng XJ, Hill VR. A broadly reactive one-step real-time RT-PCR assay for rapid and sensitive detection of hepatitis E virus. *J Virol Methods.* 2006;131:65–71. <https://doi.org/10.1016/j.jviromet.2005.07.004>
8. Bower WA, Nainan OV, Han X, Margolis HS. Duration of viremia in hepatitis A virus infection. *J Infect Dis.* 2000;182:12–7. <https://doi.org/10.1086/315701>
9. Hall TA. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. 1999 [cited 2019 Apr 17]. <http://brownlab.mbio.ncsu.edu/JWB/papers/1999Hall1.pdf>
10. Connolly MA, Gayer M, Ryan MJ, Salama P, Spiegel P, Heymann DL. Communicable diseases in complex emergencies: impact and challenges. *Lancet.* 2004;364:1974–83. [https://doi.org/10.1016/S0140-6736\(04\)17481-3](https://doi.org/10.1016/S0140-6736(04)17481-3)

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## *Rickettsia parkeri* and *Candidatus Rickettsia andeanae* in *Amblyomma maculatum* Group Ticks

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DOI: <https://doi.org/10.3201/eid2602.190664>

We determined prevalence of *Rickettsia* spp. in 172 ticks of the *Amblyomma maculatum* group collected from 16 urban sites in Oklahoma City, Oklahoma, USA, during 2017 and 2018. Most ticks (59.3%) were collected from 1 site; 4 (2.3%) were infected with *Rickettsia parkeri* and 118 (68.6%) with *Candidatus Rickettsia andeanae*.

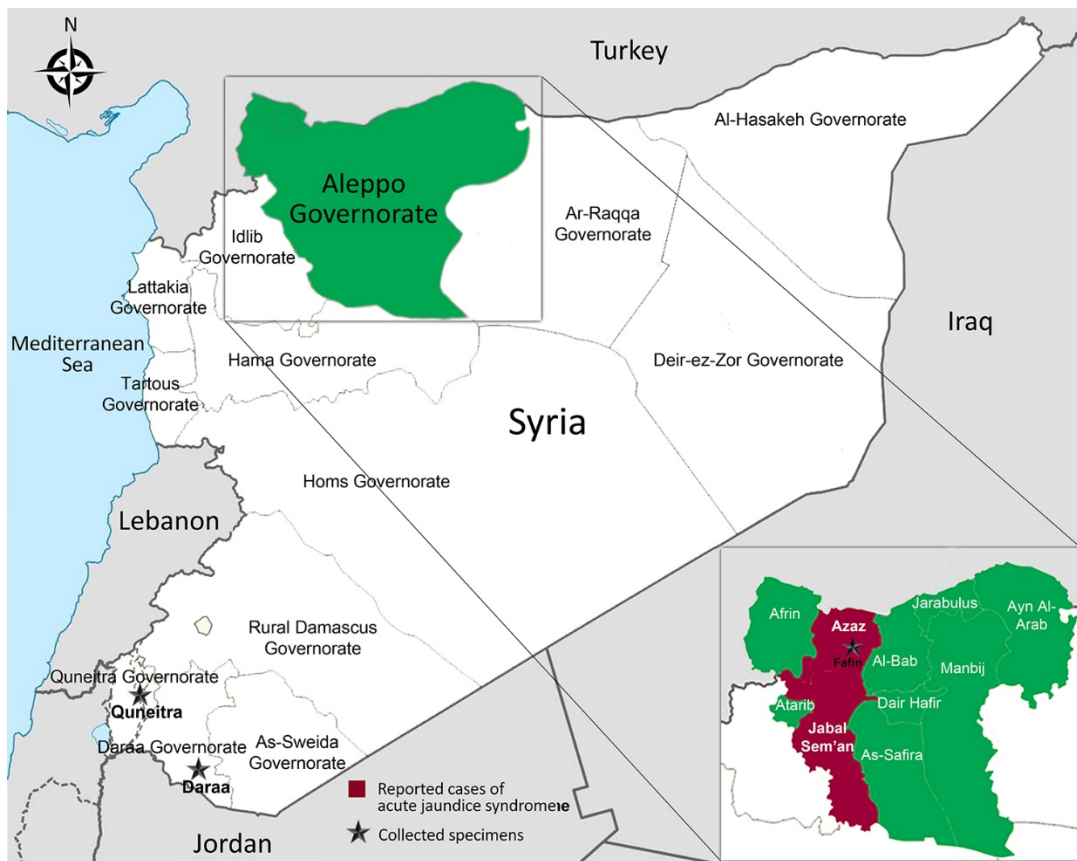
*Rickettsia parkeri*, part of the spotted fever group *Rickettsia* (SFGR), affects humans throughout much of the southern United States (1). Although *R. parkeri* in an engorged nymph was reported once in Oklahoma, *R. parkeri* has not been reported in adult *A. maculatum* ticks in Oklahoma or Kansas. To date, all test-positive adult ticks in Kansas and Oklahoma have been infected with *Candidatus Rickettsia andeanae* (2). The absence of *R. parkeri* in Oklahoma is surprising because it was detected in *A. maculatum* group ticks recovered from dogs in Arkansas counties bordering eastern Oklahoma (3) and in adult *A. maculatum* ticks in Texas (4), and *A. maculatum* ticks have been present in Oklahoma since the 1940s (4). We collected *A. maculatum* ticks in the Oklahoma City metropolitan area during May–August 2017 and 2018 and tested them for *Rickettsia* spp.

We selected 16 sites as part of a larger study of tickborne disease epidemiology (Figure). We performed collections during May–August by flagging vegetation or using CO<sub>2</sub> traps (5). We completed identification by using established keys (6).

We tested field-collected ticks for rickettsial DNA by using established PCR protocols (7,8). To limit DNA contamination, we conducted DNA extractions by using site-specific reagents in a separate laboratory. After soaking adult ticks in deionized water for 30 minutes and surface-sterilizing with 70% ethanol, we longitudinally bisected ticks; we used one half for DNA extraction and stored the other half at –80°C. DNA extraction followed established protocols (5). In 2017, we screened all ticks by using assays targeting the *gltA* and *ompA* (8) genes and retested positive samples by using an assay targeting the *ompB* gene (primer pair 120–2788/120–3599) (7). In 2018, we

# Hepatitis A Virus Genotype 1B Outbreak among Internally Displaced Persons, Syria

## Appendix



**Appendix Figure.** Map of Syria showing the locations of the hepatitis A outbreak.