

# Tracing the Origin, Spread, and Molecular Evolution of Zika Virus, Puerto Rico, 2016–2017

## Appendix

### Methods

#### Sample Selection

The ZIKV genomes sequenced in this study were obtained from residual clinical serum samples collected through two surveillance systems and tested at the CDC Dengue Branch: the Sentinel Enhanced Dengue Surveillance System (SEDSS) operated by Ponce Health Sciences University (PHSU) and the island-wide arboviral disease surveillance system coordinated by PRDH. All residual samples were handled in accordance with the institutional review boards of CDC protocol # 6731. Serum samples meeting all of the following criteria were selected randomly and independently from each of the eight health regions of Puerto Rico (Figure, panel D): collection within 5 days of symptom onset, ZIKV RNA detected by RT-PCR (*I*) with a cycle threshold (CT) value lower than 32, recorded municipality of residence of the case-patient, and sample collection dates during March 2016 to January 2017. An average of 10 samples were selected for sequencing from each health region.

#### Next-generation Sequencing

Complete ZIKV genomes were generated directly from clinical serum samples using a modification of PrimalSeq, a targeted method using overlapping PCR amplicon sequencing (2,3). Viral RNA was extracted from selected archived serum samples using the automated MagNA Pure 96 system (Roche) with the MP96 DNA and Viral RNA Small or Large volume protocol suggested by the manufacturer. All RNA samples were re-tested by RT-PCR (*I*) to confirm eligibility by relative RNA concentration using CT value. First-strand cDNA was synthesized with random hexamers using SuperScript IV reverse transcription (ThermoFisher) and overlapping 400 bp PCR amplicons were generated using 2 separate primer pools (2) and Q5®

high-fidelity DNA polymerase (New England Biolabs). Only samples presenting clearly visible bands of target size in DNA gel electrophoresis for both primer pools progressed to DNA library preparation. Qualifying PCR products were purified with magnetic beads (AMPure XP, Beckman Coulter) and concentration reassessed using Qubit 4.0 (ThermoFisher). DNA libraries were prepared using NEBNext Ultra II DNA Library Prep Kit for Illumina (New England Biolabs) reducing all reagents volumes to 25% of the suggested manufacturer protocol and the resulting products were screened for size and quality with the 2100 Bioanalyzer instrument (Agilent Technologies) and for quantity using the Qubit 4 fluorometer (ThermoFisher). Libraries that passed QC were pooled and run in MiSeq (Illumina) using the MiSeq Reagent Kit v3 in 600-cycle protocol.

### **Sequence Analysis and Generation of Consensus Genomes**

The resulting sequence reads were assembled into complete ZIKV genomes using the iVar (3) computational package designed for assemblies of amplicon-based sequencing. Briefly, reference-based amplicon alignments were performed using bwa-mem. Primers and low-quality reads were trimmed in the pipeline. Samtools was used for sorting, indexing and variant calling (3). Consensus genomes were called by iVar for positions with at least 10x depth and a minimum of 75% frequency of the consensus base. All consensus sequences were inspected manually using MEGA X (<https://www.megasoftware.net/>) or Geneious Prime (<https://www.geneious.com/prime/>). A total of 91 samples were sequenced but only 83 genomes assembled with more than 75% coding sequence coverage at 10X depth. Only these 83 complete genomes were considered for the rest of this study. All sequence data obtained for this study was submitted to GenBank: accession numbers MW122373-MW122455.

### **Phylogenetic Analyses**

Our final dataset for phylogenetic and evolutionary dynamics analyses comprised 316 complete genomes including the 83 PR genomes obtained for this study and an additional 233 published genomes, mostly from the Americas. A complete list of all the genome sequences used in this study including GenBank accession numbers can be found in the Supplemental Material (Appendix Table). All multiple sequence alignments were generated with MAFFT (4) with subsequent manual editing. All multiple sequence alignments were screened for recombination breakpoints using Recombination Detection Program (RDP4) (5) and no evidence of recombination was found. Phylogenetic trees for large datasets were reconstructed with

maximum likelihood (ML) methods including model finding functions or GTR+G pre-selected model options and over 1,000 bootstrap replicates using IQ-TREE v1.6.12 software (6). The temporal signal of the ML trees and compatibility with molecular clock phylogenies were assessed with TempEst v1.5.3 (7). Evolutionary hypothesis testing and further Bayesian phylogenetic trees to study the PR lineages were reconstructed with maximum clade credibility (MCC) methods using BEAST v1.10.4 (8). We estimated nucleotide substitution rates (evolutionary rate), effective population size and MCC trees using Bayesian Markov Chain Monte Carlo (MCMC) approach. For all BEAST analyses, the date of sample collection in the format of yyyy-mm-dd was used to time-calibrate tips and the model was parameterized using the SRD or Yang model of nucleotide substitution, strict or relaxed lognormal molecular clocks, and Bayesian constant coalescent or Skygrid model, depending on dataset. All MCMC were run for sufficient length to ensure stationary parameters with statistical errors reflected in 95% highest probability density ranges including 10% burn-in, with ESS values higher than 200 for each tree prior. The resulting trees were visualized in Figtree v1.4.4 software (<http://tree.bio.ed.ac.uk/software/figtree>). Additional information and real time tracking of ZIKV molecular evolution globally can be found here: <https://nextstrain.org/zika>.

## References

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<https://doi.org/10.1093/ve/vew007>
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<https://doi.org/10.1093/ve/vey016>

**Appendix Table.** Genomic sequences included in phylogenetic analyses.

GenBank accession no.	Country	Sample collection date
MW122373	Puerto Rico	7/20/2016
MW122374	Puerto Rico	4/27/2016
MW122375	Puerto Rico	7/13/2016
MW122376	Puerto Rico	6/28/2016
MW122377	Puerto Rico	7/30/2016
MW122378	Puerto Rico	7/1/2016
MW122379	Puerto Rico	8/15/2016
MW122380	Puerto Rico	8/26/2016
MW122381	Puerto Rico	10/17/2016
MW122382	Puerto Rico	6/23/2016
MW122383	Puerto Rico	6/25/2016
MW122384	Puerto Rico	9/19/2016
MW122385	Puerto Rico	6/23/2016
MW122386	Puerto Rico	7/17/2016
MW122387	Puerto Rico	7/14/2016
MW122388	Puerto Rico	10/31/2016
MW122389	Puerto Rico	9/7/2016
MW122390	Puerto Rico	10/11/2016
MW122391	Puerto Rico	8/6/2016
MW122392	Puerto Rico	11/7/2016
MW122393	Puerto Rico	6/24/2016
MW122394	Puerto Rico	7/14/2016
MW122395	Puerto Rico	7/23/2016
MW122396	Puerto Rico	8/11/2016
MW122397	Puerto Rico	9/27/2016
MW122398	Puerto Rico	9/19/2016
MW122399	Puerto Rico	11/4/2016
MW122452	Puerto Rico	1/19/2017
MW122400	Puerto Rico	11/10/2016
MW122401	Puerto Rico	9/15/2016
MW122453	Puerto Rico	7/27/2016
MW122402	Puerto Rico	7/1/2016
MW122403	Puerto Rico	6/30/2016
MW122404	Puerto Rico	8/15/2016
MW122405	Puerto Rico	8/19/2016
MW122406	Puerto Rico	8/18/2016
MW122407	Puerto Rico	8/31/2016
MW122408	Puerto Rico	9/27/2016
MW122409	Puerto Rico	10/20/2016

GenBank accession no.	Country	Sample collection date
MW122410	Puerto Rico	10/25/2016
MW122411	Puerto Rico	10/16/2016
MW122412	Puerto Rico	11/3/2016
MW122413	Puerto Rico	12/11/2016
MW122414	Puerto Rico	8/15/2016
MW122415	Puerto Rico	1/20/2017
MW122416	Puerto Rico	6/4/2016
MW122417	Puerto Rico	6/13/2016
MW122418	Puerto Rico	5/26/2016
MW122419	Puerto Rico	7/9/2016
MW122420	Puerto Rico	8/9/2016
MW122421	Puerto Rico	8/17/2016
MW122422	Puerto Rico	9/7/2016
MW122423	Puerto Rico	10/6/2016
MW122424	Puerto Rico	12/2/2016
MW122425	Puerto Rico	6/29/2016
MW122426	Puerto Rico	7/3/2016
MW122427	Puerto Rico	8/23/2016
MW122428	Puerto Rico	9/15/2016
MW122429	Puerto Rico	10/2/2016
MW122430	Puerto Rico	11/3/2016
MW122454	Puerto Rico	6/27/2016
MW122431	Puerto Rico	11/3/2016
MW122432	Puerto Rico	6/28/2016
MW122433	Puerto Rico	7/27/2016
MW122434	Puerto Rico	7/19/2016
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KU729217	Brazil	2015
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KU758877	French Guiana	12/2015
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KU991811	Brazil	3/6/2016
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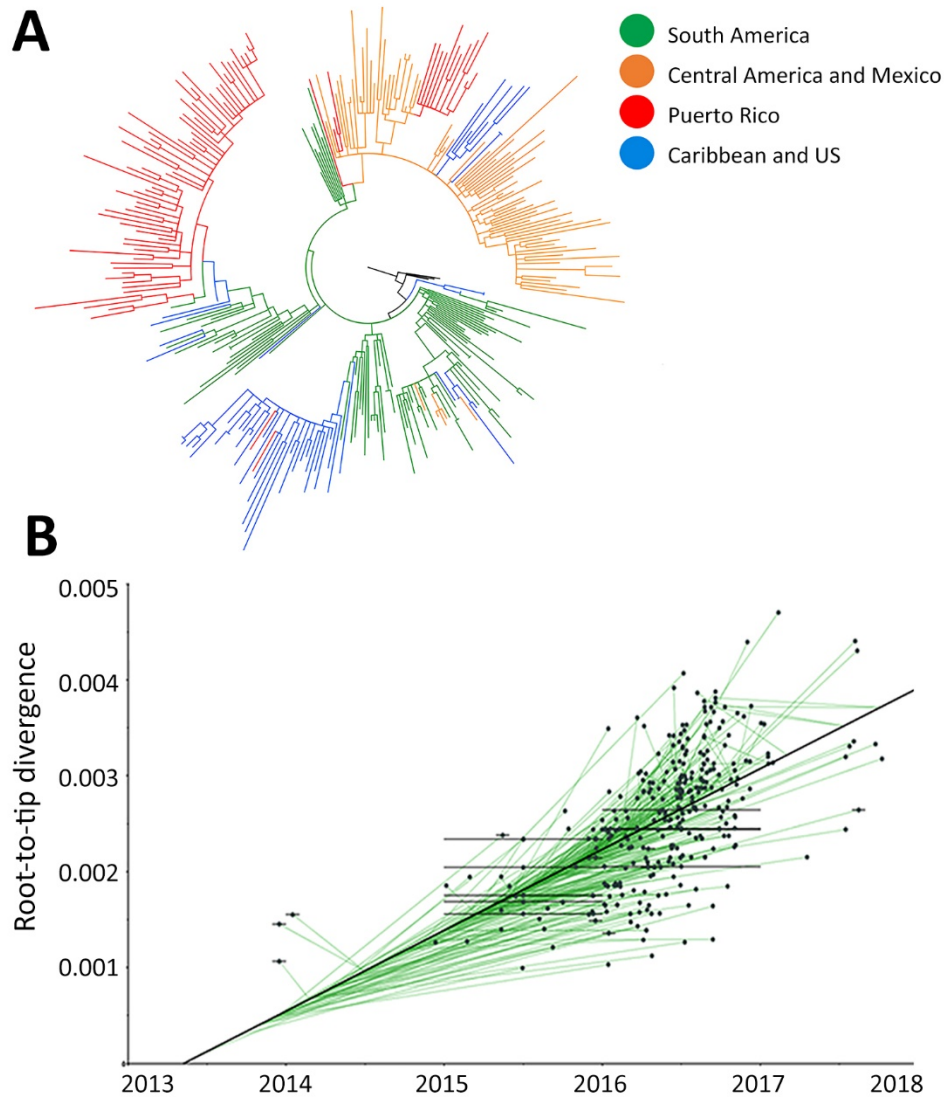
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KX702400	Venezuela	3/25/2016
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KX811222	Brazil	6/14/2016
KX832731	United States	8/24/2016
KX879603	Ecuador	04/2016
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KY325479	United States	9/28/2016
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KY558994	Brazil	1/18/2016
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KY559003	Brazil	8/28/2015
KY559005	Brazil	4/18/2016
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KY559020	Brazil	3/7/2016
KY559022	Brazil	3/13/2016
KY559025	Brazil	1/15/2016
KY559026	Brazil	1/15/2016
KY559027	Brazil	2/16/2016
KY559032	Brazil	01/2016
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KY606273	Mexico	6/30/2016
KY606274	Mexico	7/7/2016
KY631492	Brazil	1/8/2016
KY631494	Mexico	10/15/2015
KY648934	Mexico	2016
KY693676	Honduras	8/26/2016
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KY693678	Peru	6/28/2016
KY693679	Peru	7/11/2016
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MF073359	Brazil	3/1/2015
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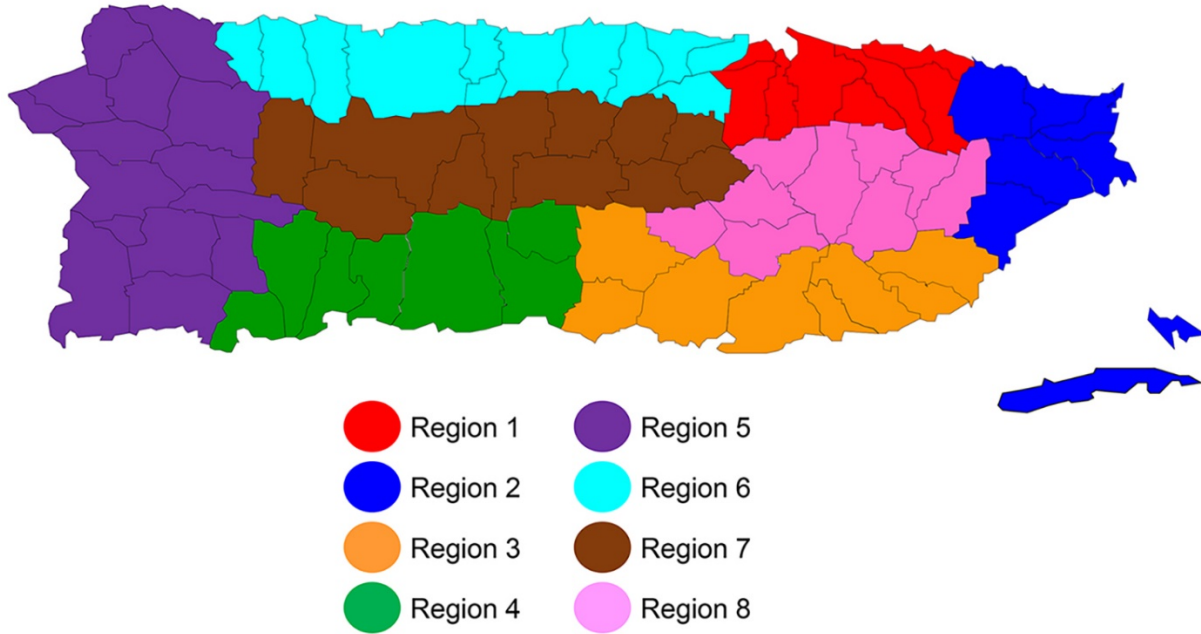
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MG494697	Mexico	11/24/2016
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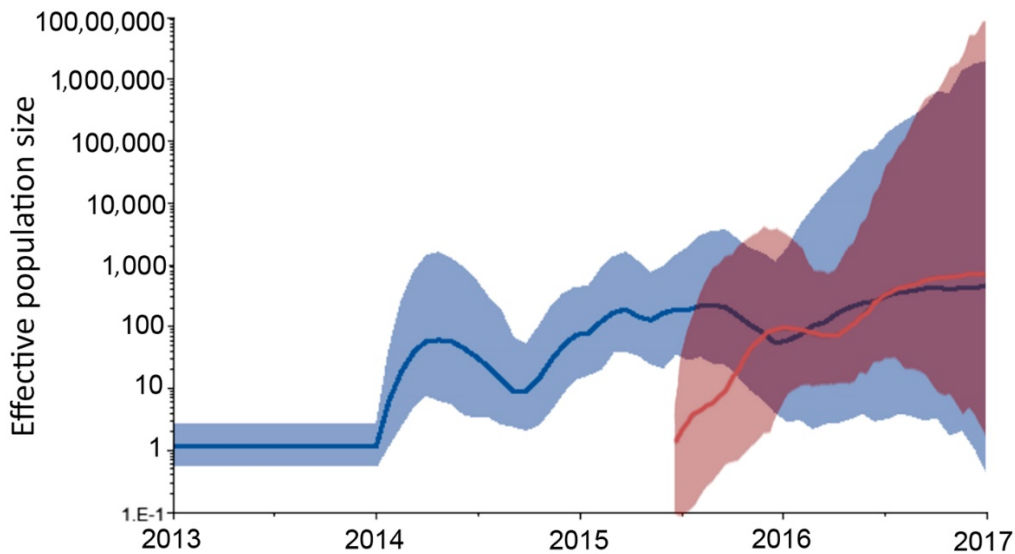
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**Appendix Figure 1.** Emergence and evolution of Zika virus in the Americas and Puerto Rico. (A) Maximum likelihood (ML) phylogenetic inference of 316 complete ZIKV genomes, including 83 genomes from Puerto Rico and 233 genomes from Asia and the Americas obtained from GenBank. The ML tree was rooted to 3 genomes from French Polynesia 2013–2014. Tree is color-coded by geographic region of genome collection; red branches represent PR genomes. (B) Root-to-tip regression analysis showing genome divergence versus sampling time, demonstrating that our dataset follows a molecular clock.



**Appendix Figure 2.** Map of Puerto Rico health regions. Health regions defined by the Puerto Rico Department of Health.



**Appendix Figure 3.** Genetic diversity of ZIKV in the Americas and in Puerto Rico. Nonparametric estimation of effective population size over time as an inference of past population dynamics from genomic sequence data using Bayesian Skygrid plots. Genetic diversity of ZIKV in the Americas estimated over 150 genomes excluding Puerto Rico (blue plot), compared to the genetic diversity upon emergence in Puerto Rico including 66 genomes from PR clade 1 (red plot).