

Genomic Analysis of Novel Poxvirus Brazilian Porcupinepox Virus, Brazil, 2019

Appendix

Summary of Viral Enrichment to Next-Generation Sequencing

For viral particle enrichment before next-generation sequencing, the eyelid lesion was cut into small pieces, added to 1mL of TE 1x (pH 8.0), and frozen and thawed twice. The sample was clarified at 16,000xg/3min and filtrate at 0.45um syringe filter. Nuclease treatment was performed in a final volume of 300uL with 50U of Benzonase Nuclease (Sigma-Aldrich, <https://www.sigmaaldrich.com>), 20U of Turbo DNase (ThermoFisher Scientific, <https://www.thermofisher.com>), 6U of RNase Cocktail Enzyme Mix, and 1x Turbo DNase Buffer. Reaction was incubated at 37°C for 1h and DNA extraction was performed immediately by using DNeasy Blood and Tissue kit (QIAGEN, <https://www.qiagen.com>).

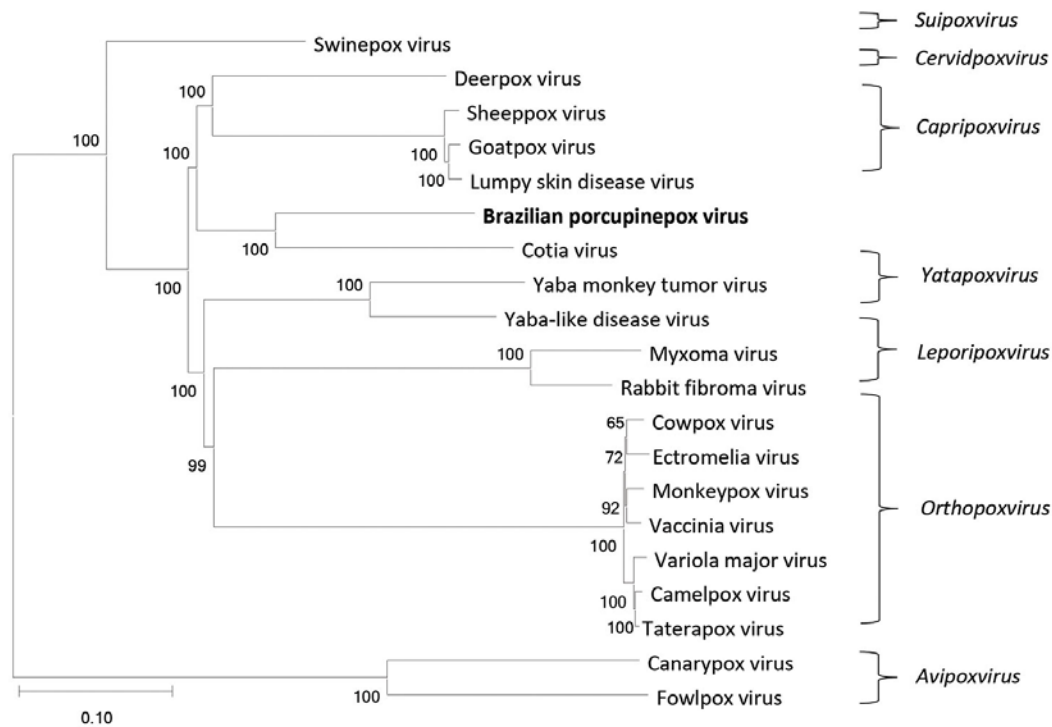
Summary of Genome Annotation

We annotated the genome by using a combination of 3 methods: Genome Annotation Transfer Utility (GATU) (1), GeneMarkS-2 (2), and Vgas (3). The open reading frames >50 aa with $\leq 25\%$ overlap with neighboring genes were initially selected. Two proteins were considered similar if BLAST search resulted an e-value $\leq 10^{-5}$ and maximal scoring pair alignment in the BLASTp searches covered $\geq 40\%$ of the longer protein.

References

- <jrn>1. Tcherepanov V, Ehlers A, Upton C. Genome Annotation Transfer Utility (GATU): rapid annotation of viral genomes using a closely related reference genome. BMC Genomics. 2006;7:150. [PubMed https://doi.org/10.1186/1471-2164-7-150](https://doi.org/10.1186/1471-2164-7-150)</jrn>
- <jrn>2. Lomsadze A, Gemayel K, Tang S, Borodovsky M. Modeling leaderless transcription and atypical genes results in more accurate gene prediction in prokaryotes. Genome Res. 2018;28:1079–89. [PubMed https://doi.org/10.1101/gr.230615.117](https://doi.org/10.1101/gr.230615.117)</jrn>

<jrn>3. Zhang KY, Gao YZ, Du MZ, Liu S, Dong C, Guo FB. Vgas: a viral genome annotation system. *Front Microbiol.* 2019;10:184. [PubMed](https://doi.org/10.3389/fmicb.2019.00184)
<https://doi.org/10.3389/fmicb.2019.00184></jrn>



Appendix Figure 1. Phylogenetic tree constructed in genomic analysis of novel poxvirus Brazilian porcupinepox virus, Brazil, 2019. Tree constructed by using the Neighbor-Joining method and Jukes-Cantor model for complete genomes of selected strains representing different genera of chordopoxviruses with low GC contents. The numbers next to each node represent the values of 1,000 bootstrap repetitions, and only those >50% are shown. Evolutionary analyses were conducted in MAFFT (<https://mafft.cbrc.jp>). GenBank accession numbers are as follows: Brazilian porcupinepox virus, MK944278.1; Camelpox virus, AY009089.1; Canarypox virus, NC005309.1; Cotia virus, KM595078.1; Cowpox virus, DQ437593.1; Deerpox virus, AY689437.1; Ectromelia virus, NC004105.1; Fowlpox virus, NC002188.1; Goatpox virus, MH381810.1; Lumpy skin disease virus, NC003027.1; Monkeypox virus, DQ011157.1; Myxoma virus, NC001132.2; Rabbit fibroma virus, NC001266.1; Sheeppox virus, NC004002.1; Swinepox virus, NC003389.1; Taterapox virus, NC008291.1; Vaccinia virus, M35027.1; Variola major virus, L22579.1; Yaba monkey tumor virus, NC005179.1; Yaba-like disease virus, NC002642.1.

