The durability of antibody responses to severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), the virus responsible for coronavirus disease (COVID-19), is of scientific and strategic interest for public health systems worldwide. After SARS-CoV-2 infection, antibodies are produced against multiple viral epitopes, including the nucleocapsid (N) protein, which is highly immunogenic and abundantly expressed (1). A key concern is the potential for rapid waning of antibodies and seroreversion (loss of detectable antibodies), as seen with other novel betacoronaviruses (2), which might represent declining immunity and could compromise serosurveillance.

Frontline healthcare workers are a vital population for serosurveillance because they are at greater risk than the general population. We describe findings from a serosurveillance study conducted in London, UK, by Public Health England (PHE).

The Study
We conducted prospective serosurveillance of healthcare professionals in secondary care settings across London beginning March 30, 2020. Healthcare workers were recruited by hospital research teams and provided written informed consent. Demographic, occupational, and clinical data were collected at baseline, including self-reported previous laboratory-confirmed COVID-19. Participants provided blood samples and completed symptom surveys at baseline and 2-weekly intervals until July 21, 2020, reporting any new illness or COVID-19 diagnosis. Blood samples were centrifuged and frozen locally; PHE then tested serum samples by using the Elecsys Anti-SARS-CoV-2 total antibody assay (Roche, https://www.roche.com), according to the manufacturer’s instructions. This test is an electrochemiluminescence immunoassay for antibodies targeting the N protein (IgG, IgM, or IgA) and produces a numeric cutoff index derived from comparison of the sample and calibrator signals (3). The surveillance protocol was approved by the PHE Research Ethics Governance Group (R&D REGG Ref: NR0192, March 31, 2020).

We compared differences in seropositivity between groups by using χ² tests and multivariable logistic regression to provide adjusted odds ratios (aORs). We estimated biweekly seroconversion and seroreversion rates and binomial 95% CIs. We analyzed trends in individual-level antibody responses beginning 4 weeks after the first positive antibody test, which allowed time for responses to stabilize. We used mixed effects regression to analyze trends in log antibody titers and assessed fixed effects for differences in antibody response through likelihood ratio tests.

Surveillance involved 1,069 participants from 4 hospitals: Charing Cross (n = 192), Northwick Park (n = 217), Royal Free (n = 126), and St. George’s (n = 534).
Of these, 850 participants had ≥4 sampling visits and 395 ≥6 sampling visits (over 10–12 weeks of follow-up). Overall, 312 (29%) participants had ≥1 positive antibody test (95% CI 26%–32%); of those, 181 (58%) had ≥8 weeks and 42 (13%) 12 weeks of follow-up after the first positive test (Appendix Table 1, https://wwwnc.cdc.gov/EID/article/27/4/20-4554-App1.pdf). Sero-positivity varied between hospitals (p = 0.042), from 25% to 35%. In total, 109 (10.2%) participants self-reported laboratory-confirmed COVID-19, 407 (32%) reported respiratory illness, 5 (0.47%) reported hospitalization, and 794 (61%) did not report illness.

We observed no difference in seropositivity by sex, profession, performance of aerosol-generating procedures, employment in the emergency department, or immunocompromised status (Appendix Figure).
Table 2). Participants 25–34 years of age had higher odds of seropositivity than those 35–44 years of age (aOR 1.57, 95% CI 1.09–2.26), but little difference was seen among older age groups. Those working in intensive care units had lower odds of seropositivity than participants from other hospital departments (aOR 0.58, 95% CI 0.38–0.91).

Most seropositive participants tested positive at baseline (279/312, 89%). Only 33 participants seroconverted during follow-up, corresponding to a biweekly rate of 1.2% (95% CI 0.8%–1.7%). We observed 4 seroreversions, corresponding to a biweekly rate of 0.4% (95% CI 0.1%–0.9%).

Log antibody titers remained stable over time in seropositive participants, and little within-individual variability was observed (Figure). The general trend across all subgroups was a slight increase over time, although data are sparse for some groups.

We modeled trends beginning 4 weeks after the first positive antibody test. The mean weekly change was a 3.9% increase (95% CI 3.2%–4.6%). The model enables individual variability and thus estimates a distribution in trends, which ranged from a 0.5% decrease to an 8.5% increase per week, at 1 SD below/above the mean.

Baseline response or subsequent trend did not differ by work setting, clinical symptoms, or laboratory-confirmed COVID-19; minimum likelihood ratio p value was 0.46. Participants ≥40 years of age had 30% higher antibody titers at baseline (p = 0.08) but less increase over time; weekly increase was 2.9% (95% CI 1.8%–4.0%) compared with 4.5% (95% CI 3.6%–5.4%) in those <40 years of age (p = 0.028). We observed similar baseline titers between women and men (p = 0.61) but different trends; women demonstrated a weekly increase of 3.4% (95% CI 2.6%–4.2%) compared with 5.2% (95% CI 3.8%–6.6%) in men (p = 0.035).

Conclusions
In this study, N-antibody seropositivity was 29% among healthcare workers, and a small, sustained rise in antibody titers occurred over 12 weeks. The increase could be explained by the natural boosting of antibodies through repeated SARS-CoV-2 exposure; however, we saw no evidence of sporadic, sharp increases in antibodies in seropositive participants, and we observed little deviation from an overall linear trend. High initial seroprevalence and low subsequent seroconversion rates (Appendix Figures 1, 2) indicate that most exposures occurred before surveillance began. The low seroconversion after April might be attributable to changes in hospital infection control practices and national lockdown.

These findings demonstrate the short-term stability of N-antibody titers in healthcare staff, regardless of demographic or clinical differences. Seropositive participants not reporting any COVID-19 diagnosis or previous illness (even mild or atypical symptoms) demonstrated the same antibody trends as those who reported symptoms or laboratory-confirmed COVID-19, thereby supporting N-antibody testing as a reliable surveillance indicator. Although seroreversion was uncommon, such rates, if sustained, might be concerning in the long term.

Although cross-reactivity against the N protein has been observed and appears more prevalent than cross-reactivity against the spike (S) protein (E.M. Anderson, unpub. data, https://doi.org/10.1101/2020.1.06.20227215; C.F. Houlihan, unpub. data, https://doi.org/10.1101/2020.06.08.20120584), the risk for false positives because of preexisting human coronavirus antibodies seems low on the basis of available data. The Elecsys assay demonstrated >99.5% specificity in 2 independent evaluations using large numbers of prepandemic control samples (3,4) and demonstrated high positive predictive value at an estimated 10% seroprevalence. Nonetheless, this study is limited by use of a single immunoassay, by self-reported data on COVID-19 diagnosis, and by limited testing early in the pandemic.

Several studies have demonstrated substantial declines in antibody titers over 3–5 months by using anti-S or anti–receptor-binding domain immunoassays (5–9). Although findings are not consistent across all reports (6,10), disparities could be explained by shorter follow-up periods that missed later decline. In contrast, the few studies conducting serial testing for ≥3 months by using N-antibody assays, particularly the Elecsys assay, report that titers remained steady (9) or increased (11; F. Muecksch, unpub. data, https://doi.org/10.1101/2020.08.05.20169128). These studies were limited by small sample sizes, single-site recruitment, and few time points with long sampling intervals. Our study replicates these findings in a large, multicenter cohort with frequent sampling and focuses on healthcare workers with mostly asymptomatic or mild disease, with robust statistical analysis to demonstrate consistent findings across all groups. These data can usefully inform serosurveillance strategies during the second wave.

For unknown reasons, N-antibodies appear highly stable in the short term, despite demonstrating no functional role; whether this stability would persist over longer follow-up periods remains to be answered. Although less useful as correlates of
immunity, N-antibodies could serve a critical role in serosurveillance as S-based vaccines are deployed, helping to distinguish infection-induced seroconversion from vaccine-induced seroconversion.

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About the Author
Dr. Shrotri is a specialty registrar in public health medicine based in London and holds an honorary contract with Public Health England. Her research interests include epidemiology of communicable diseases and vaccinology.

References

Address for correspondence: Madhumita Shrotri, Public Health England, Wellington House, 133-155 Waterloo Rd, London, SE1 8UG, UK; email: Madhumita.shrotri@phe.gov.uk