

3. Newman KL, Rogers JH, McCulloch D, Wilcox N, Englund JA, Boeckh M, et al.; Seattle Flu Study Investigators. Point-of-care molecular testing and antiviral treatment of influenza in residents of homeless shelters in Seattle, WA: study protocol for a stepped-wedge cluster-randomized controlled trial. *Trials*. 2020;21:956. <https://doi.org/10.1186/s13063-020-04871-5>
4. Rogers JH, Link AC, McCulloch D, Brandstetter E, Newman KL, Jackson ML, et al.; Seattle Flu Study Investigators. Characteristics of COVID-19 in homeless shelters: a community-based surveillance study. *Ann Intern Med*. 2021;174:42–9. <https://doi.org/10.7326/M20-3799>
5. Chow EJ, Casto AM, Roychoudhury P, Han PD, Xie H, Pfau B, et al. The clinical and genomic epidemiology of rhinovirus in homeless shelters, King County, Washington. *J Infect Dis*. 2022;jiac239. <https://doi.org/10.1093/infdis/jiac239>
6. Chow EJ, Casto AM, Rogers JH, Roychoudhury P, Han PD, Xie H, et al. The clinical and genomic epidemiology of seasonal human coronaviruses in congregate homeless shelter settings: A repeated cross-sectional study. *Lancet Reg Health Am*. 2022;15:100348. <https://doi.org/10.1016/j.lana.2022.100348>
7. Olsen SJ, Winn AK, Budd AP, Prill MM, Steel J, Midgley CM, et al. Changes in influenza and other respiratory virus activity during the COVID-19 pandemic-United States, 2020–2021. *Am J Transplant*. 2021;21:3481–6. <https://doi.org/10.1111/ajt.16049>
8. Henrickson KJ. Parainfluenza viruses. *Clin Microbiol Rev*. 2003; 16:242–64. <https://doi.org/10.1128/CMR.16.2.242-264.2003>
9. Brady MT, Evans J, Cuartas J. Survival and disinfection of parainfluenza viruses on environmental surfaces. *Am J Infect Control*. 1990;18:18–23. [https://doi.org/10.1016/0196-6553\(90\)90206-8](https://doi.org/10.1016/0196-6553(90)90206-8)
10. Hall CB, Geiman JM, Breese BB, Douglas RG Jr. Parainfluenza viral infections in children: correlation of shedding with clinical manifestations. *J Pediatr*. 1977;91:194–8. [https://doi.org/10.1016/S0022-3476\(77\)80811-1](https://doi.org/10.1016/S0022-3476(77)80811-1)

Address for correspondence: Eric J. Chow, Division of Allergy and Infectious Diseases, University of Washington, 1959 NE Pacific St, Box 356423, Seattle, WA 98195, USA; email: ejchow@uw.edu

Presence of *Spirometra mansonii*, Causative Agent of Sparganosis, in South America

Jan Brabec,¹ Manuel Uribe,¹
Jenny J. Chaparro-Gutiérrez, Carlos Hermosilla

Author affiliations: Biology Centre of the Czech Academy of Sciences, České Budějovice, Czech Republic (J. Brabec); CIBAV Research Group, Universidad de Antioquia, Medellín, Colombia (M. Uribe, J.J. Chaparro-Gutiérrez); Justus Liebig University Giessen, Giessen, Germany (M. Uribe, C. Hermosilla)

DOI: <http://doi.org/10.3201/eid2811.220529>

We report molecular identification of an adult *Spirometra mansonii* tapeworm retrieved from a crab-eating fox (*Cerdocyon thous*) in Colombia, confirming presence of this parasite in South America. This tapeworm is the causative agent of human sparganosis, commonly reported from Southeast Asia, and represents the second congeneric species with known zoonotic potential in the Americas.

Sparganosis is a neglected human zoonosis caused by migrating larval stages of the broad tapeworm genus *Spirometra* (Diphyllobothriidea), whose natural definitive hosts include wild and domestic canids and felids. The life cycle of this tapeworm involves 2 intermediate hosts: a freshwater copepod crustacean as the first and various vertebrates, mostly amphibians, as the second. Human infections are commonly reported from Southeast Asia and propagate most often in the form of subcutaneous sparganosis; however, the larvae can enter other organs or parts of central nervous system and cause damage.

Taxonomy of *Spirometra* remains highly complicated. Numerous species of *Spirometra* have been described, often poorly (1), and representatives of just 6 species-level lineages have been characterized molecularly so far, a key prerequisite to achieve a convincing tapeworm identification when only strobila fragments or larval stages are available. Limitations of morphologic characters of *Spirometra* are numerous and include characters' great intraspecific and even intra-individual variability (overview of problematic traits in 2). Molecular sequence data thus represent the only unequivocal method of species identification.

Previous phylogenetic analysis of *Spirometra* has shown that the geographic distribution of the 6 lineages respects continental borders (2). North

¹These authors contributed equally to this article.

and South America were shown to share 2 lineages found exclusively on those continents (3), provisionally termed *Spirometra decipiens* complex 1 and 2 because of the lack of essential morphologic data precluding conclusive species determination (2). *S. decipiens* complex 1 was shown to house, among parasites of canids and felids, causative agents of cutaneous and proliferative sparganosis. Representatives of *S. decipiens* complex 2, on the other hand, have not yet been shown to cause the zoonosis. The frequently reported human cases of sparganosis from Southeast Asia, as well as numerous

specimens from wildlife from the region, corresponded to *S. mansoni* (2).

We report molecular identification of a tapeworm specimen retrieved from a dead crab-eating fox (*Cerdocyon thous*) from the vicinity of Ciudad Bolívar, Antioquia, Colombia. We characterized the specimen through Sanger-sequencing of 3 genetic loci (Appendix, <https://wwwnc.cdc.gov/EID/article/28/11/22-0529-App1.pdf>), including the complete mitochondrial cytochrome c oxidase subunit I gene (*cox1*) as the most densely sampled and phylogenetically informative gene of broad tapeworms.

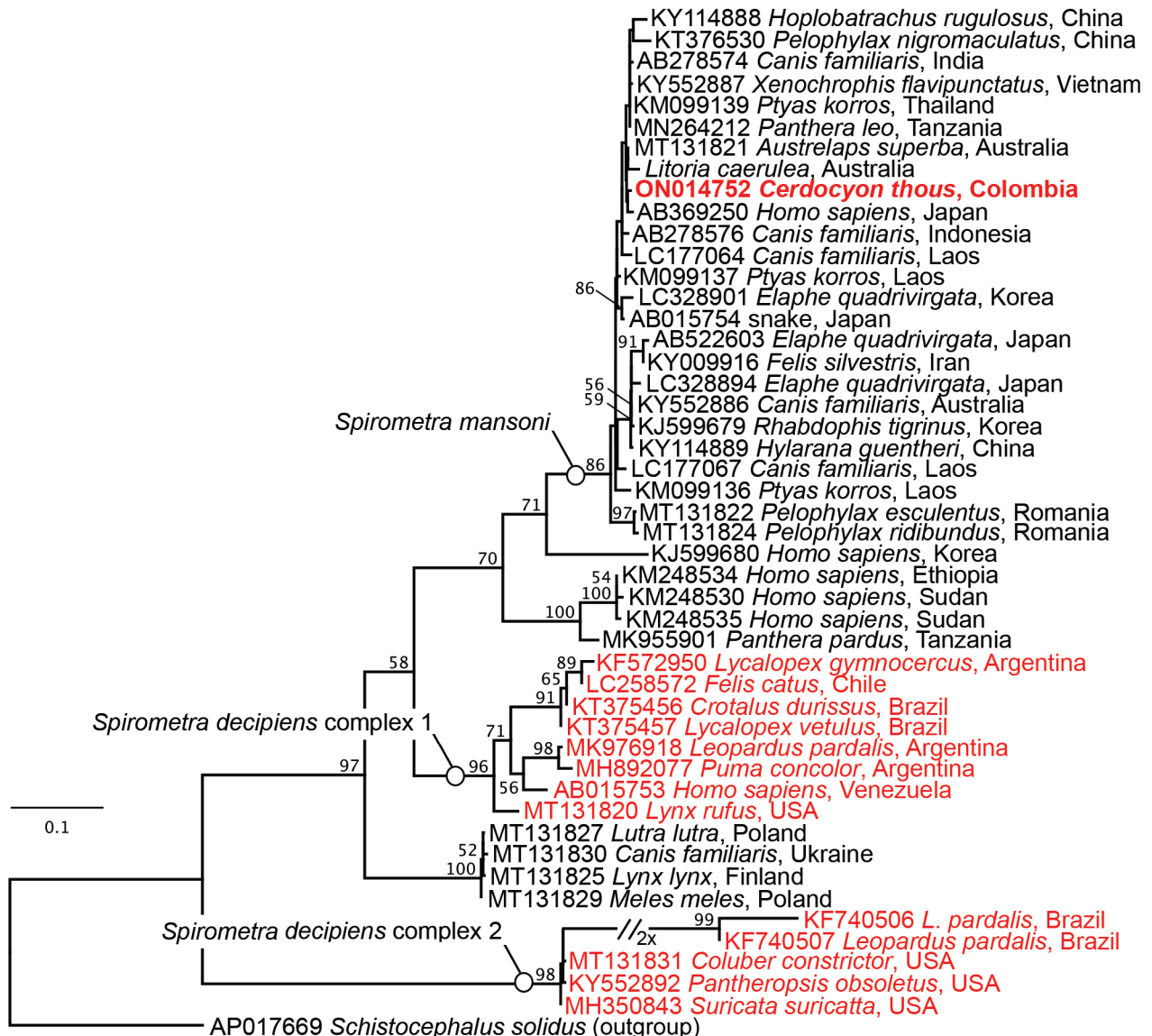


Figure. Maximum-likelihood estimate of the phylogenetic position of a *Spirometra mansoni* tapeworm collected from a crab-eating fox (*Cerdocyon thous*) in Colombia. Red indicates specimens from South America; bold indicates newly characterized *S. mansoni* from this report. Names of the 3 species-level lineages of *Spirometra* in South America are indicated; GenBank numbers are provided. Nodal support values show standard bootstrap supports >50. Scale bar indicates number of substitutions per site.

Phylogenetic analysis under maximum-likelihood criterion resolved the position of the tapeworm nested deep within the clade of *S. mansoni* (Figure), proving the presence of this causative agent of human sparganosis on the American continents.

S. mansoni represents by far the most frequently reported causative agent of sparganosis, previously misidentified as *S. erinaceieuropaei* (2). This species is responsible for virtually all human cases in Asia but has been also shown to infect wildlife in Africa, Australia, and Eastern Europe (2). Our finding of *S. mansoni* in Colombia in a crab-eating fox, a definitive host endemic and widely distributed across South America, from Panama to the Entre Ríos province of Argentina (4), expands the known distribution of *S. mansoni* into broader range than previously thought. This finding contrasts with the distribution of the remaining 5 lineages of *Spirometra*, which seem limited to continental regions (2). *S. mansoni* has been sporadically reported from the Americas in the past; however, morphology-altering fixation techniques and lack of critical molecular evidence did not support species identification. Reported hosts mostly included domestic cats (Appendix) and a single report from a crab-eating fox in Brazil (5).

The crab-eating fox inhabits savannah and woodland areas of various Neotropical habitats from coastal plains to montane forests and is considered omnivorous, opportunistically feeding on fruits, insects, and small vertebrates including amphibians and reptiles, with seasonal shifts to its diet (6,7). A broad range of Neotropical amphibians and reptiles has been found to serve as intermediate hosts of *Spirometra*; however, the record remains skewed toward herpetofauna of the more intensively surveyed coastal regions (8), and species identification of the parasite larvae has been, thanks to the lack of accompanying molecular data, either absent or ungrounded. As a result, the real range and the relevance of different intermediate hosts for the transmission of the sympatric South America species of *Spirometra* remain unknown. The situation in North America is even more obscure because of the virtually missing intermediate host record (1,9). Given the wide spectrum of suitable intermediate hosts of *S. mansoni*, which includes omnivores such as wild boar in Europe (10), the natural pools and the importance of different host species in the etiology of the zoonosis remain dubious. The concurrent presence of the second congeneric species with zoonotic potential urges deeper investigations into the parasite's life cycles and the epizootiology of a disease that could affect public health in the Americas.

Acknowledgments

The CIBAV research group thanks the Strategy of consolidation of Research Groups CODI 2018–2019, University of Antioquia, Medellín, Colombia.

This work was supported by the Czech Science Foundation project no. 19-28399X.

About the Author

Dr. Brabec is a research associate at the Institute of Parasitology, Biology Centre of the Czech Academy of Sciences. His primary research interests include molecular taxonomy, phylogenetics, phylogenomics, and evolution of parasitism in flatworms and protists.

References

- Scholz T, Kuchta R, Brabec J. Broad tapeworms (Diphyllobothriidae), parasites of wildlife and humans: recent progress and future challenges. *Int J Parasitol Parasites Wildl.* 2019;9:359–69. <https://doi.org/10.1016/j.ijppaw.2019.02.001>
- Kuchta R, Kołodziej-Sobocińska M, Brabec J, Młocicki D, Salamatin R, Scholz T. Sparganosis (*Spirometra*) in Europe in the molecular era. *Clin Infect Dis.* 2021;72:882–90. <https://doi.org/10.1093/cid/ciaa1036>
- Almeida GG, Coscarelli D, Melo MN, Melo AL, Pinto HA. Molecular identification of *Spirometra* spp. (Cestoda: Diphyllobothriidae) in some wild animals from Brazil. *Parasitol Int.* 2016;65(5 Pt A):428–31. <https://doi.org/10.1016/j.parint.2016.05.014>
- Lucherini M. *Cerdocoyon thous*. The IUCN Red List of Threatened Species. 2015:e.T4248A81266293 [cited 2022 Jul 18]. <https://doi.org/10.2305/IUCN.UK.2015-4.RLTS.T4248A81266293.en>
- Santos KR, Catenacci LS, Pestelli MM, Takahira RK, Silva RJ. First report of *Diphyllobothrium mansoni* (Cestoda, Diphyllobothridae) infecting *Cerdocoyon thous* (Mammalia, Canidae) in Brazil. *Arq Bras Med Vet Zootec.* 2004;56:796–8. <https://doi.org/10.1590/S0102-09352004000600016>
- Berta A. *Cerdocoyon thous*. *Mamm Species.* 1982;186:1–4. <https://doi.org/10.2307/3503974>
- Bianchi RC, Campos RC, Xavier-Filho NL, Olifiers N, Gompper ME, Mourão G. Intraspecific, interspecific, and seasonal differences in the diet of three mid-sized carnivores in a large neotropical wetland. *Acta Theriol (Warsz).* 2014;59:13–23. <https://doi.org/10.1007/s13364-013-0137-x>
- Oda FH, Borteiro C, da Graça RJ, Tavares LER, Crampet A, Guerra V, et al. Parasitism by larval tapeworms genus *Spirometra* in South American amphibians and reptiles: new records from Brazil and Uruguay, and a review of current knowledge in the region. *Acta Trop.* 2016;164:150–64. <https://doi.org/10.1016/j.actatropica.2016.09.005>
- McHale B, Callahan RT, Paras KL, Weber M, Kimbrell L, Velázquez-Jiménez Y, et al. Sparganosis due to *Spirometra* sp. (cestoda; Diphyllobothriidae) in captive meerkats (*Suricata suricatta*). *Int J Parasitol Parasites Wildl.* 2020;13:186–90. <https://doi.org/10.1016/j.ijppaw.2020.10.005>
- Kołodziej-Sobocińska M, Miniuk M, Ruczyńska I, Tokarska M. Sparganosis in wild boar (*Sus scrofa*) – implications for veterinarians, hunters, and consumers.

Vet Parasitol. 2016;227:115–7. <https://doi.org/10.1016/j.vetpar.2016.08.001>

Address for correspondence: Jan Brabec, Institute of Parasitology, Biology Centre of the Czech Academy of Sciences, Branišovská 31, České Budějovice, 37005, Czech Republic; email: brabcak@paru.cas.cz

TIGIT Monoallelic Nonsense Variant in Patient with Severe COVID-19 Infection, Thailand

Pimpayao Sodsai,¹ Chupong Ittiwut,¹ Vichaya Ruenjaiman, Rungnapa Ittiwut, Watsamon Jantarabenjakul, Kanya Suphapeetiporn, Vorasuk Shotelersuk,² Nattiya Hirankarn²

Author affiliations: Chulalongkorn University, Bangkok, Thailand (P. Sodsai, C. Ittiwut, V. Ruenjaiman, R. Ittiwut, W. Jantarabenjakul, K. Suphapeetiporn, V. Shotelersuk, N. Hirankarn); King Chulalongkorn Memorial Hospital, The Thai Red Cross Society, Bangkok (W. Jantarabenjakul, R. Ittiwut)

DOI: <https://doi.org/10.3201/eid2811.220914>

A heterozygous nonsense variant in the *TIGIT* gene was identified in a patient in Thailand who had severe COVID-19, resulting in lower *TIGIT* expression in T cells. The patient's T cells produced higher levels of cytokines upon stimulation. This mutation causes less-controlled immune responses, which might contribute to COVID-19 severity.

To investigate SARS-CoV-2 genomic variants, we recruited 46 COVID-19 patients from King Chulalongkorn Memorial Hospital in Bangkok, Thailand, in January 2020. Recruited patients were 16–79 years of age and had moderate to severe COVID-19 symptoms according to World Health Organization interim guidelines (<https://apps.who.int/iris/bitstream/handle/10665/331446/WHO-2019-nCoV-clinical-2020.4-eng.pdf>). We performed whole-exome sequencing on peripheral blood samples as described

¹These first authors contributed equally to this article.

²These authors were co-principal investigators.

(1). The institutional review board of the Faculty of Medicine, Chulalongkorn University, Bangkok, approved this study (COA no. 738/2020).

We filtered variants by using the following criteria. Variants had to pass the quality standards, have read depth >10, and be from the coding regions or canonical splice sites of 1,810 immune-related genes, including immune checkpoint genes (2). Variants also had to have <1% allele frequency in the Genome Aggregation Database (gnomAD, <https://gnomad.broadinstitute.org>), Exome Variant Server (University of Washington, <https://evs.gs.washington.edu/EVS>), 1000 Genomes Project Consortium (<https://www.genome.gov>), dbSNPs (<https://www.ncbi.nlm.nih.gov/projects/SNP>), and Thai Reference Exome (T-Rex) database (3). We called candidate variants novel pathogenic variants when they were not previously identified in patients in the literature.

In our patient cohort, exome sequencing identified no variants in type I interferon genes, which previously have been commonly observed in patients with severe COVID-19 (4). Of note, we identified a heterozygous nonsense variant (rs1386709957) in the T-cell immunoglobulin and ITIM domain (*TIGIT*) gene in 1 patient (Appendix Figure 1, <https://wwwnc.cdc.gov/EID/article/29/11/22-0914-App1.pdf>). We did not identify this nonsense variant among 3,742 persons in the T-Rex database but did observe it in 1 of 31,390 alleles in the gnomAD database, in an allele from a female patient from East Asia. This variant truncates the 245-amino acid residue proteins at residue 56 and is classified as a pathogenic variant American College of Medical Genetics guidelines (<https://www.acmg.net>).

We investigated *TIGIT* gene expression in T cells of the patient from our study (Co45), a 43-year-old man, and compared it with 2 other sex- and age-matched patients who had severe COVID-19 (Co6 and Co84) (Appendix). We collected peripheral blood mononuclear cells (PBMCs) from each of the patients 1 month after they recovered. We used RNA extracted from PBMCs for real-time reverse transcription PCR and found patient Co45 had the lowest *TIGIT* mRNA level (Figure, panel A). Because *TIGIT* is mainly expressed in T cells, we used flow cytometry to measure the mean fluorescence intensity of *TIGIT* expressed in the cytoplasmic domain (CD) T cells. Patient Co45 had lower *TIGIT* gene expression in all CD3+, CD4+, and CD8+ T cells than the other 2 patients, most remarkably in the CD8+ T cells (Figure, panels B–D). The percentages of CD3+, CD4+, and CD8+ T cells in patient Co45 were comparable those in the other 2 patients (Appendix Fig-