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## Acknowledgments

We thank the women who participated in this study, research nurses, pediatricians, medical technologists, and the ZIKAction Project Management team (Francesca Viero and Giorgia Dalla Valle).

As Global Infectious Diseases Scholars, C.M. and Y.L. received mentored research training in the development of this manuscript. This training was supported in part by the University of Buffalo Clinical and Translational Science Institute award UL1TR001412 and the Global Infectious Diseases Research Training Program award D43TW010919.

This project was funded from the Horizon 2020: EU Programme for Research and Innovation under grant agreement no. 734857.

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### References

- Duncan J, Gordon-Johnson KA, Tulloch-Reid MK, Cunningham-Myrie C, Ernst K, McMorris N, et al. Chikungunya: important lessons from the Jamaican experience. Rev Panam Salud Publica. 2017;41:e60. https://doi.org/10.26633/RPSP.2017.60
- Webster-Kerr KR, Christie C, Grant A, Chin D, Burrowes H, Clarke K, et al. Emergence of Zika virus epidemic and the national response in Jamaica. West Indian Med J. 2016; 65:243–9.
- Gallian P, Leparc-Goffart I, Richard P, Maire F, Flusin O, Djoudi R, et al. Epidemiology of chikungunya virus outbreaks in Guadeloupe and Martinique, 2014: an observational study in volunteer blood donors. PLoS Negl Trop Dis. 2017;11:e0005254. https://doi.org/10.1371/ journal.pntd.0005254
- Poirier MJ, Moss DM, Feeser KR, Streit TG, Chang G-JJ, Whitney M, et al. Measuring Haitian children's exposure to chikungunya, dengue and malaria. Bull World Health Organ. 2016;94:817–825A.

- Simmons G, Brès V, Lu K, Liss NM, Brambilla DJ, Ryff KR, et al. High incidence of chikungunya virus and frequency of viremic blood donations during epidemic, Puerto Rico, USA, 2014. Emerg Infect Dis. 2016;22:1221–8. https://doi.org/ 10.3201/eid2207.160116
- Gay N, Rousset D, Huc P, Matheus S, Ledrans M, Rosine J, et al. Seroprevalence of Asian lineage chikungunyua virus infection on Saint Martin Island, 7 months after the 2013 emergence. Am J Trop Med Hyg. 2016;94:393–6. https://doi.org/10.4269/ajtmh.15-0308
- Langerak T, Brinkman T, Mumtaz N, Arron G, Hermelijn S, Baldewsingh G, et al. Zika virus seroprevalence in urban and rural areas of Suriname, 2017. J Infect Dis. 2019;220:28–31. https://doi.org/10.1093/infdis/jiz063
- Gallian P, Cabié A, Richard P, Paturel L, Charrel RN, Pastorino B, et al. Zika virus in asymptomatic blood donors in Martinique. Blood. 2017;129:263–6. https://doi.org/ 10.1182/blood-2016-09-737981
- 9. Ehmen C, Medialdea-Carrera R, Brown D, Bispo de Filippis AM, de Sequeira PC, Nogueira RMR, et al. Accurate detection of Zika virus IgG using a novel immune complex binding ELISA. Trop Med Int Health. 2020.
- Rodriguez-Barraquer I, Costa F, Nascimento EJM, Nery N, Castanha PMS, Sacramento GA, et al. Impact of preexisting dengue immunity on Zika virus emergence in a dengue endemic region. Science. 2019;363:607–10. https://doi.org/ 10.1126/science.aav6618

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# Group IV Getah Virus in *Culex* Mosquitoes, Malaysia

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DOI: https://doi.org/10.3201/eid2802.204887

A new Getah virus (GETV) strain, B254, was isolated from *Culex fuscocephalus* mosquitoes captured at Mount Ophir, Malaysia, in 2012. Phylogenetic analyses revealed that GETV B254 is distinct from the old Malaysia GETV MM2021 strain but closely related to group IV GETV from Russia (LEIV16275Mag), China (YN12031), and Thailand (GETV/SW/Thailand/2017).

Getah virus (GETV) is an emerging mosquitoborne alphavirus of the family *Togaviridae*. The virus was first reported in 1955 from *Culex gelidus* mosquitoes collected near Kuala Lumpur, Malaysia (1). Serologic evidence of GETV infection was found in various large domestic mammals (2) and humans (3). GETV infections in these populations, however, were mostly inapparent. In other regions, reproductive failures in pregnant sows, death in piglets, and hind limb edema in racehorses, as well as neurologic symptoms and death in blue foxes, have been reported (4–7).

We conducted mosquito surveillance in the forests of 4 different states in Peninsular Malaysia, Perak, Pahang, Selangor, and Johor, during 2011–2014. We captured a total of 4,160 mosquitoes from the study sites by BG-Sentinel CO<sub>2</sub> trap (Biogents AG, https:// eu.biogents.com) with the addition of UV light and sorted them into 208 pools according to their species, determined by morphologic keys, and collection sites. We homogenized the pooled mosquitoes, inoculated

them into C6/36 mosquito cells, and blind passaged for 5 passages. We obtained cytopathic effects from 1 of the pools on day 4 postinfection during the last passage. The pool comprised *Cx. fuscocephalus* mosquitoes collected from the forested area at Mount Ophir (Malay: Gunung Ledang) in Johor in 2012. We screened the culture supernatants by reverse transcription PCR (RT-PCR) using the in-house developed alphavirus generic primers targeting the nsP4 gene and flavivirus primers targeting the nonstructural 5-3' untranslated region junction. A similar sample pool that showed cytopathic effects was found positive for the presence of alphaviruses, as indicated by a 683 bp amplicon. Sanger sequencing of the amplicon revealed high sequence similarity to GETV. We obtained the complete coding sequence of the GETV isolate, designated as B254, using 10 pairs of RT-PCR amplification primers and genome sequencing. We deposited the genome in GenBank (accession no. LR990838).

We constructed a maximum clade credibility tree using the Bayesian Markov chain Monte Carlo analysis based on the complete coding regions of GETV B254 and other GETV isolates available in GenBank (Figure). The GETV phylogeny revealed 4 major groups of viruses: group I (GI), GII, GIII, and GIV (8). GI and GII consisted of early identified GETV isolates; GI contained the Malaysia MM2021 isolate (1955), and GII, Japan Sagiyama (1956). The GIII and GIV comprised the



**Figure.** Maximum-clade credibility tree of complete coding sequences of Getah virus (GETV) strain B254 from Malaysia (black dot) and reference strains. Horizontal branches are drawn to a scale of estimated year of divergence. Times to the most recent common ancestor with 95% highest posterior density values (ranges in parentheses) are shown at nodes. Posterior probability values  $\geq$ 0.6 of key nodes are shown. G, group.

most recent circulating virus strains. The newly isolated Malaysia GETV B254 is phylogenetically distinct from the ancestral Malaysia MM2021. It clustered within the GIV clade, which included the strains recovered in Russia (LEIV16275Mag, 2007), China (YN12031, 2012) and Thailand (GETV/SW/Thailand/2017). The B254 strain showed the highest nucleotide sequence homology with YN12031 (98.8%), isolated from the *Armigeres subalbatus* mosquito, and GETV/SW/Thailand/2017 (98.6%), isolated from pig serum.

Currently, only the GIII GETV lineage is associated with epidemic outbreaks. The GIV GETV, however, consisted of only a few virus strains. We calculated the mean evolutionary rate of GIV GETV at  $4.10 \times 10^{-4}$  substitutions/site/year and the rate for GIII at 2.98  $\times 10^{-4}$  substitutions/site/year. These results suggest that the GIV viruses may be under a different selection pressure, potentially involving a different host. It is also possible that GIV viruses are maintained in a yet-to-be-identified enzootic transmission cycle involving mosquitoes and asymptomatic hosts.

The *Culex* mosquitoes, particularly *Cx. gelidus* and *Cx. tritaeniorhynchus*, have been identified as the most predominant vector host for GETV in Malaysia (1,2). *Cx. fuscocephalus* mosquitoes, however, are not an uncommon vector; they have been known to carry the virus in China and Sri Lanka (9,10). *Cx. fuscocephalus* mosquitoes could be an emerging vector for GETV transmission in Malaysia along with the other *Culex* mosquitoes.

The high similarity of genome sequences of GIV GETV strains recovered thousands of kilometers apart raised the possibility that the virus may share a common dispersal route. One possible route linking the virus is the East Asian–Australasian flyway of migratory birds. GIV GETV could have been introduced to the regions through the migratory birds along the flyway, which included Malaysia as one of the stopover sites; the abundantly available *Culex* mosquitoes at the roosting sites could have maintained the transmission.

In summary, we identified a new GETV strain, B254, in Malaysia that is phylogenetically distinct from the old Malaysia MM2021 strain. The virus strain shares high similarities to the widely distributed GIV GETV. Although further surveillance studies are needed for confirmation, this finding suggests that GIV GETV strains could share a common dispersal origin, possibly through the bird transmigratory flyways.

### Acknowledgments

We thank World Refence Center for Emerging Viruses and Arboviruses, The University of Texas Medical Branch, Galveston, USA, for kindly providing us with the Malaysia GETV strain MM2021. This study was supported in parts by the Ministry of Education Malaysia for niche area research under the Higher Institution Centre of Excellence (HICoE) program (project MO002–2019), the Fundamental Research Grant Scheme (FRGS FP014-2019A), and the Long Term Research Grant Scheme (LRGS/2011A/UM/ Penyakit-Berjangkit).

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#### References

- Simpson DI, Way HJ, Platt GS, Bowen ET, Hill MN, Kamath S, et al. Arbovirus infections in Sarawak, October 1968–February 1970: GETAH virus isolations from mosquitoes. Trans R Soc Trop Med Hyg. 1975;69:35–8. https://doi.org/10.1016/0035-9203(75)90008-5
- Marchette NJ, Rudnick A, Garcia R, MacVean DW. Alphaviruses in peninusular Malaysia: I. virus isolations and animal serology. Southeast Asian J Trop Med Public Health. 1978;9:317–29.
- Marchette NJ, Rudnick A, Garcia R. Alphaviruses in peninsular Malaysia: II. serological evidence of human infection. Southeast Asian J Trop Med Public Health. 1980;11:14–23.
- Brown CM, Timoney PJ. Getah virus infection of Indian horses. Trop Anim Health Prod. 1998;30:241–52. https://doi.org/10.1023/A:1005079229232
- Nemoto M, Bannai H, Tsujimura K, Kobayashi M, Kikuchi T, Yamanaka T, et al. Getah virus infection among racehorses, Japan, 2014. Emerg Infect Dis. 2015;21:883–5. https://doi.org/ 10.3201/eid2105.141975
- Yang T, Li R, Hu Y, Yang L, Zhao D, Du L, et al. An outbreak of Getah virus infection among pigs in China, 2017. Transbound Emerg Dis. 2018;65:632–7. https://doi.org/ 10.1111/tbed.12867
- Shi N, Li LX, Lu RG, Yan XJ, Liu H. Highly pathogenic swine Getah virus in blue foxes, eastern China, 2017. Emerg Infect Dis. 2019;25:1252–4. https://doi.org/10.3201/eid2506.181983
- Li YY, Liu H, Fu SH, Li XL, Guo XF, Li MH, et al. From discovery to spread: the evolution and phylogeny of Getah virus. Infect Genet Evol. 2017;55:48–55. https://doi.org/10.1016/j.meegid.2017.08.016
- Peiris JS, Amerasinghe PH, Amerasinghe FP, Calisher CH, Perera LP, Arunagiri CK, et al. Viruses isolated from mosquitoes collected in Sri Lanka. Am J Trop Med Hyg. 1994;51:154–61. https://doi.org/10.4269/ajtmh.1994.51.154
- Zhang HL, Zhang YZ, Yang WH, Feng Y, Nasci RS, Yang J, et al. Mosquitoes of Western Yunnan Province, China: seasonal abundance, diversity, and arbovirus associations. PLoS One. 2013;8:e77017. https://doi.org/ 10.1371/journal.pone.0077017

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