

Laboratory Misidentifications Resulting from Taxonomic Changes to *Bacillus cereus* Group Species, 2018–2022

Appendix

Phylogeny Construction for Figure 1

Genomic data acquisition and taxonomic assignment. A total of 2,887 *Bacillus cereus* group genomes were downloaded and pre-processed as described previously (1). Briefly, after assessing the quality of each genome, Prokka v1.14.6 (2) was used to annotate each genome. Each genome was additionally assigned to a species within the Genome Taxonomy Database (GTDB) using the classify_wf workflow in GTDB-Tk v1.3.0 and GTDB Release 05-RS95 (GTDB R95) (3–5).

Identification of the clonal *B. anthracis* lineage most commonly associated with anthrax toxin production. FastANI v1.31 (6) was used to calculate average nucleotide identity (ANI) values between each of the 2,887 *B. cereus* group genomes and the genome of *B. anthracis* str. Ames (NCBI RefSeq accession no. GCF_000007845.1); *B. cereus* group genomes that shared ≥ 99.9 ANI with the *B. anthracis* str. Ames genome were considered to belong to the classic clonal *B. anthracis* lineage most commonly associated with anthrax toxin production, as described previously (6,7). Within a recently proposed standardized nomenclatural framework for the *B. cereus* group (7) (referred to as the 2020 Genomospecies/Subspecies/Biovar or 2020 GSB taxonomy in a recent review of *B. cereus* group taxonomy) (8), this lineage is referred to as *B. mosaicus* subspecies *anthracis* (full notation) or *B. anthracis* (using shortened subspecies notation) (7–9).

***In silico* virulence factor detection, multilocus sequence typing, and *panC* phylogenetic group assignment.** BTyper3 v3.1.1 (9) was used to query each genome for (i) anthrax toxin encoding genes *cya*, *lef*, and *pagA* and (ii) cereulide (emetic toxin) synthetase-encoding genes *cesABCD* using BLAST v2.9.0 (10) and default settings. BTyper3 was

additionally used to perform (i) *in silico* seven-gene multilocus sequence typing (MLST) using the PubMLST *B. cereus* database (accessed 25 October 2020); and (ii) *panC* phylogenetic group assignment, using an adjusted eight-group (Group I–VIII) framework (9).

Phylogeny construction. GFF files produced by Prokka associated with all genomes assigned to GTDB’s (i) *B. anthracis*, (ii) *B. paranthracis*, and (iii) *B. tropicus* species (n = 325, 226, and 53 genomes, respectively; see section “Genomic data acquisition and taxonomic assignment” above), plus (iv) *panC* Group II *B. cereus* group strain FSL W8–0169 (used as an outgroup; NCBI RefSeq Assembly Accession GCF_001583695.1) (11), were supplied as input to Panaroo v1.2.8 (12). Panaroo was used to identify “core” genes among all 605 genomes, using the following parameters (all other parameters were set to their default values): (i) “strict” mode (–clean-mode strict), (ii) core genome alignment using MAFFT (–a core–aligner mafft), (iii) a core genome sample threshold of 95% (–core_threshold 0.95). The resulting nucleotide alignment was queried using snp-sites v2.5.1 (13), which was used to identify (i) core single nucleotide polymorphisms (SNPs) and (ii) constant sites among all 605 genomes. The resulting core SNP alignment was supplied as input to IQ-TREE v1.5.4 (14), which was used to construct a maximum likelihood (ML) phylogeny using the General Time-Reversible (GTR) nucleotide substitution model (15), one thousand replicates of the ultrafast bootstrap approximation (16), and an ascertainment bias correction obtained using constant sites output by snp-sites. The resulting phylogeny was annotated and displayed using the bactaxR package (7) in R v4.1.2 (17).

Data availability. Metadata and quality information for all genomes displayed in Figure 1 of this study are available in Supplemental Tables S1–S4 of Carroll, et al., 2022 (1).

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