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Neurotropic Highly Pathogenic Avian Influenza A(H5N1) Virus in Red Foxes, Northern Germany

Appendix

Materials and Methods

RNA extraction, PCR investigations and Next Generation Sequencing (NGS) analyses

Viral RNA was extracted from the brain samples with the NucleoMag VET Kit (Macherey-Nagel) on a KingFisher 96-Flex machine, and real-time PCR targeting the viral M segment was performed with a commercial kit (Virotype Influenza A RT-PCR Kit, Indical Bioscience) according to the manufacturer's instructions. A second real-time PCR was used to confirm the H5 subtype in the samples with the following primers and probe (AIV-H5LH1: 5'-ACA TAT GAC TAC CCA CAR TAT TCA G-3', AIV-H5RH1: 5'-AGA CCA GCT AYC ATG ATT GC-3', 5'-FAM-TCA ACA GTG GCG AGT TCC CTA GCA-TAMRA-3') under these conditions: 50°C, 30min; 94°C, 2min; 40 × (94°C, 30sec; 56°C, 30sec; 68°C, 30sec) by using the SuperScript III One-Step RT-RCR-Kit (Invitrogen).

To recover full-length viral genome sequences, viral RNA segments were reverse transcribed and amplified with the Invitrogen Superscript III One-Step RT-PCR Kit (Thermo Fisher Scientific) with a universal Primer Set targeting conserved regions of all eight segments (1). Primers were adjusted to a stock concentration of 10pmol/μl. The RT-PCR reaction consisted of 12.5μl 2x reaction buffer, 1μl of each primer, 1μl of Superscript III RT / Platinum Taq enzyme mix, 3.5μl nuclease free water and 5μl RNA sample adding up to a total volume of 25μl. The RT-PCR cycler program consisted of an initial 45-minute period at 45°C followed by a 2-minute denaturation step at 94°C, then 5 cycles of 94°C for 30s, 45°C for 30s and 68°C for 3 minutes followed by 30 cycles of 94°C for 30s, 57°C for 30s and 68°C for 3 minutes. The final Elongation step was 68°C for 5 minutes.

Following this, Illumina compatible DNA libraries were prepared (ILMN DNA LP (M) Tagmentation Kit) and sequenced on an Illumina NextSeq 1000 benchtop sequencer. The quality of the reads was checked with FastQC (2) and fastp (3). The final AIV genome assembly was performed with IRMA (4).

Histopathological and immunohistochemical examination

Samples were collected from different organs and tissues including rhinencephalon, the cerebellum and other parts of the brain, fixed in 10% neutral buffered formalin, processed by routine methods, embedded in paraffin wax, sectioned (4 μ m), and stained with hematoxylin and eosin (H&E).

Immunohistochemistry was performed on paraffin-embedded tissue sections by using a murine monoclonal antibody specific for the nucleoprotein (5). Briefly, 3 μ m tissue sections were deparaffinized and rehydrated using a descending series of alcohol. Afterwards, slides were incubated in 85% ethanol with additional 0.5% H₂O₂ for 30 min. at room temperature (RT) for blocking endogenous peroxidase activities. Enzymatic pre-treatment with proteinase K was performed for 10min. at RT and unspecific background staining was blocked with inactivated goat serum diluted 1:5 in phosphate-buffered saline (PBS). The primary anti-influenza A antibody (CLONEGENE LLC, HB65) was diluted 1:200 in PBS with 1% bovine serum albumin and slides were incubated overnight at 4°C. For negative control, the primary antibody was substituted by ascites fluid from Balb/c mice. Biotinylated goat-anti-mouse IgG was used as secondary antibody and added for 45min at RT. For visualization of immunostaining, the avidin-biotin-peroxidase complex (VECTASTAIN Elite ABC Kit; Vector Laboratories, Burlingame, CA) was applied by using 3,3'-diaminobenzidine-tetrahydrochloride (Sigma Aldrich) as chromogen. Counterstaining was performed with Mayer's hemalaun.

References

1. Hoffmann E, Stech J, Guan Y, Webster RG, Perez DR. Universal primer set for the full-length amplification of all influenza A viruses. *Arch Virol*. 2001;146:2275–89 . [PubMed](#) <https://doi.org/10.1007/s007050170002>
2. Andrews S. 2010. FASTQC. A quality control tool for high throughput sequence data [cited 2023 Oct 4].<http://www.bioinformatics.babraham.ac.uk/projects/fastqc>
3. Chen S, Zhou Y, Chen Y, Gu J. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics*. 2018;34:i884–90. [PubMed](#) <https://doi.org/10.1093/bioinformatics/bty560>

4. Shepard SS, Meno S, Bahl J, Wilson MM, Barnes J, Neuhaus E. Viral deep sequencing needs an adaptive approach: IRMA, the iterative refinement meta-assembler. BMC Genomics. 2016;17:708. [PubMed](https://doi.org/10.1186/s12864-016-3030-6) <https://doi.org/10.1186/s12864-016-3030-6>

5. de Boer GF, Back W, Osterhaus AD. An ELISA for detection of antibodies against influenza A nucleoprotein in humans and various animal species. Arch Virol. 1990;115:47–61. [PubMed](https://doi.org/10.1007/BF01310622) <https://doi.org/10.1007/BF01310622>

Appendix Table 1. Overview of wild terrestrial carnivores examined from February 2022 to April 2023. The months in which animals tested positive are in bold type*

Year/month	Number of animals tested/number of AIV positive animals					
	Red fox	Raccoon	Badger	Marten	Raccoon dog	Total
2022/Feb	11/0	2/0	3/0	0/0	1/0	17/0
2022/Mar	17/0	2/0	0/0	1/0	0/0	20/0
2022/Apr	2/0	0/0	0/0	1/0	0/0	3/0
2022/May	7/0	0/0	1/0	0/0	0/0	8/0
2022/Jun	6/0	0/0	0/0	1/0	0/0	7/0
2022/Jul	4/0	3/0	2/0	1/0	0/0	10/0
2022/Aug	3/0	1/0	2/0	0/0	1/0	7/0
2022/Sep	4/0	3/0	0/0	0/0	0/0	7/0
2022/Oct	8/0	9/0	2/0	0/0	1/0	20/0
2022/Nov	5/0	1/0	3/0	0/0	2/0	11/0
2022/Dec	10/0	5/0	0/0	1/0	1/0	17/0
2023/Jan	9/3	0/0	2/0	0/0	0/0	11/3
2023/Feb	9/1	0/0	0/0	1/0	2/0	12/1
2023/Mar	12/1	1/0	0/0	2/0	0/0	15/1
2023/Apr	3/0	1/0	0/0	1/0	0/0	5/0
Total	110/5	28/0	15/0	9/0	8/0	170/5

*Months in which animals tested positive are in bold type.

Appendix Table 2. Isolates used in this study*

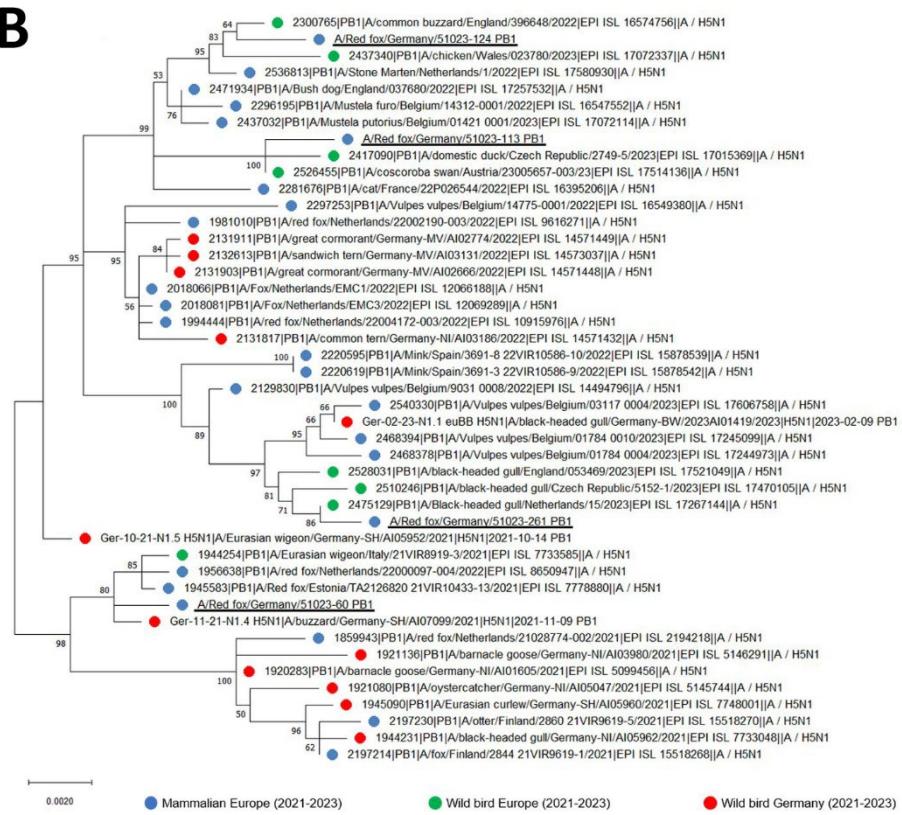
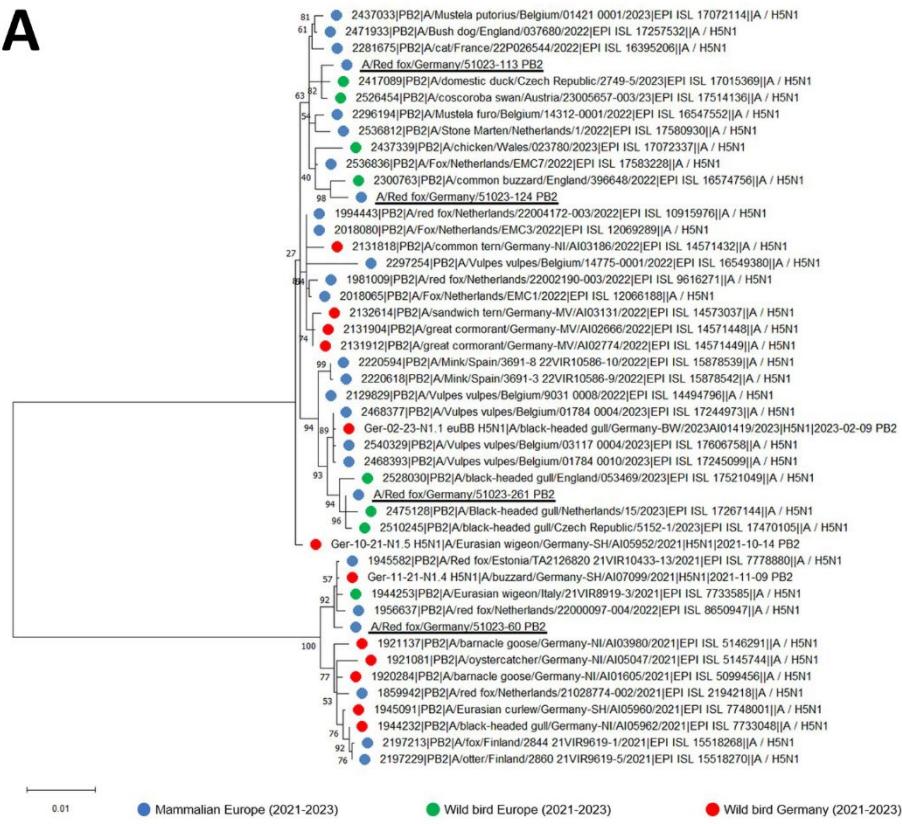
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EPI_ISL_17470105	Czech Republic	2023-Apr-04	A/black-headed_gull/Czech_Republic/5152-1/2023	State Veterinary Institute Prague	State Veterinary Institute Prague	Alexander,Nagy;Lenka,Cernikova;Martina,Stara
EPI_ISL_17267144	Netherlands	2023-Mar-01	A/Black-headed gull/Netherlands/15/2023	Erasmus Medical Center	Erasmus Medical Center	
EPI_ISL_17606758	Belgium	2023-Mar-29	A/Vulpes_vulpes/Belgium/03117_0004/2023	Sciensano - Animal Infectious Diseases	Sciensano, Department of Animal Infectious Diseases	Van Borm, Steven; Roupie, Virginie; Hostyn, Pierre; Mathijis, Elisabeth; Lambrecht, Benedicte; Steensels, Mieke
EPI_ISL_17245099	Belgium	2023-Feb-27	A/Vulpes_vulpes/Belgium/01784_0010/2023	Sciensano - Animal Infectious Diseases	Sciensano, Department of Animal Infectious Diseases	Van Borm, Steven; Roupie, Virginie; Hostyn, Pierre; Mathijis, Elisabeth; Lambrecht, Benedicte; Steensels, Mieke
EPI_ISL_17244973	Belgium	2023-Feb-27	A/Vulpes_vulpes/Belgium/01784_0004/2023	Sciensano - Animal Infectious Diseases	Sciensano, Department of Animal Infectious Diseases	Van Borm, Steven; Roupie, Virginie; Hostyn, Pierre; Mathijis, Elisabeth; Lambrecht, Benedicte; Steensels, Mieke
EPI_ISL_14494796	Belgium	2022-Jun-16	A/Vulpes_vulpes/Belgium/9031_0008/2022	Sciensano - Animal Infectious Diseases	Sciensano, Department of Animal Infectious Diseases	Van Borm, Steven; Mathijis, Elisabeth; Roupie, Virginie; Vervaeke, Muriel; Lambrecht, Benedicte; Steensels, Mieke
EPI_ISL_15878539	Spain	2022-Oct-18	A/Mink/Spain/3691-8_22VIR10586-10/2022	Laboratorio Central de Veterinaria	Istituto Zooprofilattico Sperimentale delle Venezie	Ruano, M.J.; Rocha, A.; Sanchez, A.; Aguero, M; Barbierato, G.; Zecchin, B.; Fusaro, A.; Schivo, A.; Salviato, A.; Palumbo, E.; Giussani, E.; Pastori, A.; Monne, I.; Terregino, C.
EPI_ISL_15878542	Spain	2022-Oct-18	A/Mink/Spain/3691-3_22VIR10586-9/2022	Laboratorio Central de Veterinaria	Istituto Zooprofilattico Sperimentale delle Venezie	Ruano, M.J.; Rocha, A.; Sanchez, A.; Aguero, M; Barbierato, G.; Zecchin, B.; Fusaro, A.; Schivo, A.; Salviato, A.; Palumbo, E.; Giussani, E.; Pastori, A.; Monne, I.; Terregino, C.
EPI_ISL_12069289	Netherlands	2022-Jan-22	A/Fox/Netherlands/EMC3/2022	Erasmus Medical Center	Erasmus Medical Center	R.A.M., Fouchier; S., Thewessen; O., Vuong; I., Chestakova; R., Sikkema; P., Lexmond; M., Pronk
EPI_ISL_14571448	Germany	2022-May-03	A/great cormorant/Germany-MV/AI02666/2022	Landesamt für Landwirtschaft, Lebensmittelsicherheit und Fischerei (LALLF)	Friedrich-Loeffler-Institut	
EPI_ISL_14573037	Germany	2022-May-31	A/sandwich tern/Germany-MV/AI03131/2022	Landesamt für Landwirtschaft, Lebensmittelsicherheit und Fischerei (LALLF)	Friedrich-Loeffler-Institut	

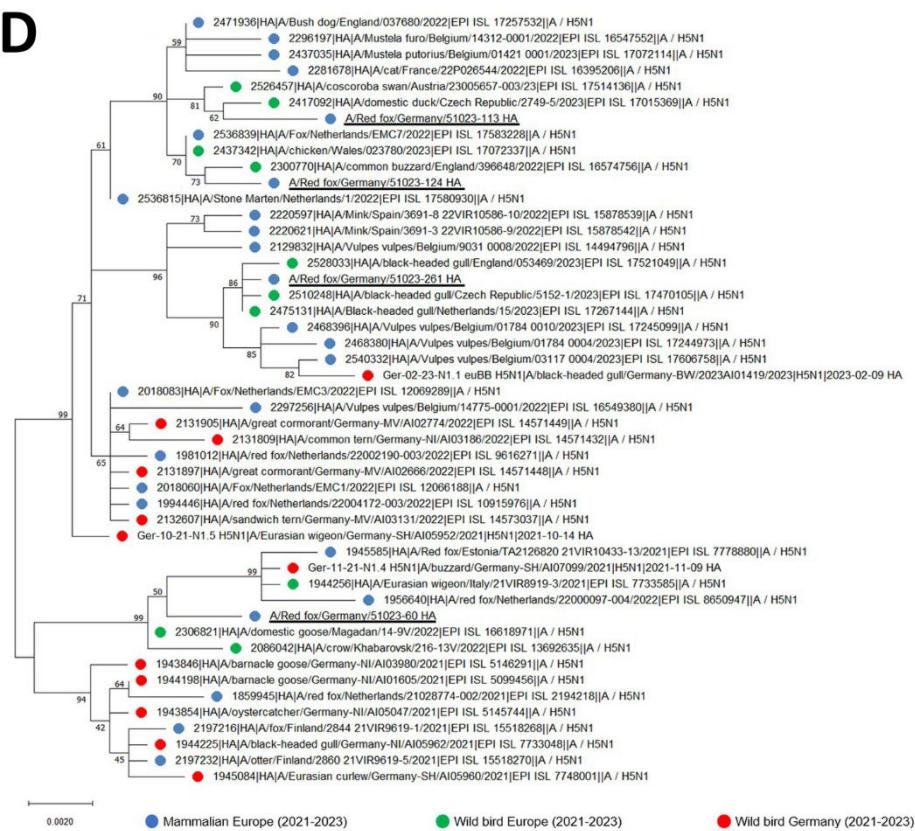
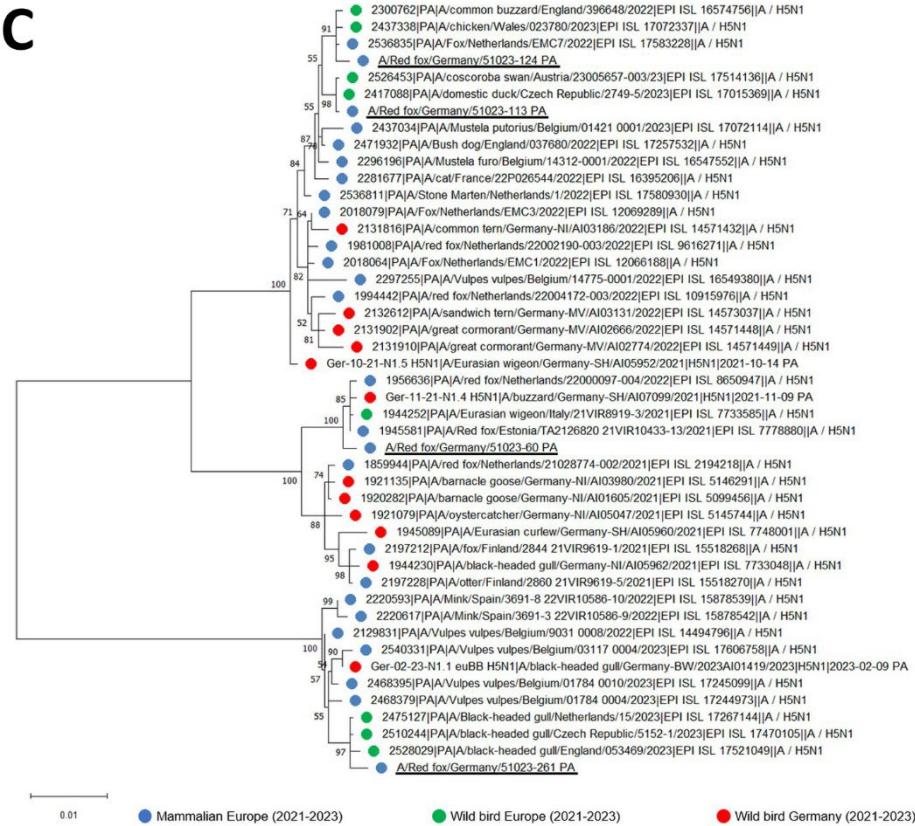
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EPI_ISL_12066188	Netherlands	2022-Jan-26	A/Fox/Netherlands/EMC1/2022	Erasmus Medical Center	Erasmus Medical Center	
EPI_ISL_9616271	Netherlands	2022-Jan-31	A/red fox/Netherlands/22002190–003/2022	Wageningen Bioveterinary Research	Wageningen Bioveterinary Research	Beerens, Nancy; Harders, Frank; Pritz-Verschuren, Sylvia; Roose, Marit; Venema, Sandra; Germeraad, Evelien; Engelsma, Marc; Heutink, Rene
EPI_ISL_10915976	Netherlands	2022-Mar-01	A/red fox/Netherlands/22004172–003/2022	Wageningen Bioveterinary Research	Wageningen Bioveterinary Research	Beerens, Nancy; Harders, Frank; Pritz-Verschuren, Sylvia; Roose, Marit; Venema, Sandra; Germeraad, Evelien; Engelsma, Marc; Heutink, Rene
EPI_ISL_14571449	Germany	2022-May-06	A/great cormorant/Germany-MV/AI02774/2022	Landesamt für Landwirtschaft, Lebensmittelsicherheit und Fischerei (LALLF)	Friedrich-Loeffler-Institut	
EPI_ISL_14571432	Germany	2022-Jun-07	A/common tern/Germany-NI/AI03186/2022	Lebensmittel- und Veterinärinstitut Oldenburg - Standort Veterinärinstitut	Friedrich-Loeffler-Institut	
EPI_ISL_17580930	Netherlands	2022-Jan-16	A/Stone Marten/Netherlands/1/2022	Erasmus Medical Center	Erasmus Medical Center	Vuong, O; Thewessen, S; Bellido-Martin, A.B; Chestakova, I; Sikkema, R; Fouchier, R.A.M.
EPI_ISL_17583228	Netherlands	2022-Sep-13	A/Fox/Netherlands/EMC7/2022	Erasmus Medical Center	Erasmus Medical Center	Vuong, O; Thewessen, S; Bellido-Martin, A.B; Chestakova, I; Sikkema, R; Fouchier, R.A.M.
EPI_ISL_17072337	United Kingdom	2023-Feb-11	A/chicken/Wales/023780/2023	Animal and Plant Health Agency (APHA)	Animal and Plant Health Agency (APHA)	
EPI_ISL_16574756	United Kingdom	2022-Dec-20	A/common_buzzard/England/396648/2022	Animal and Plant Health Agency (APHA)	Animal and Plant Health Agency (APHA)	
EPI_ISL_17015369	Czech Republic	2023-Feb-16	A/domestic_duck/Czech_Republic/2749–5/2023	State Veterinary Institute Prague	State Veterinary Institute Prague	Alexander, Nagy; Lenka, Cernikova; Martina, Stara
EPI_ISL_17514136	Austria	2023-Jan-18	A/coscoroba swan/Austria/23005657–003/23	Institute for Veterinary Disease Control Moedling, Austrian Agency for Health and Food Safety	Austrian Agency for Health and Food Safety (AGES)	

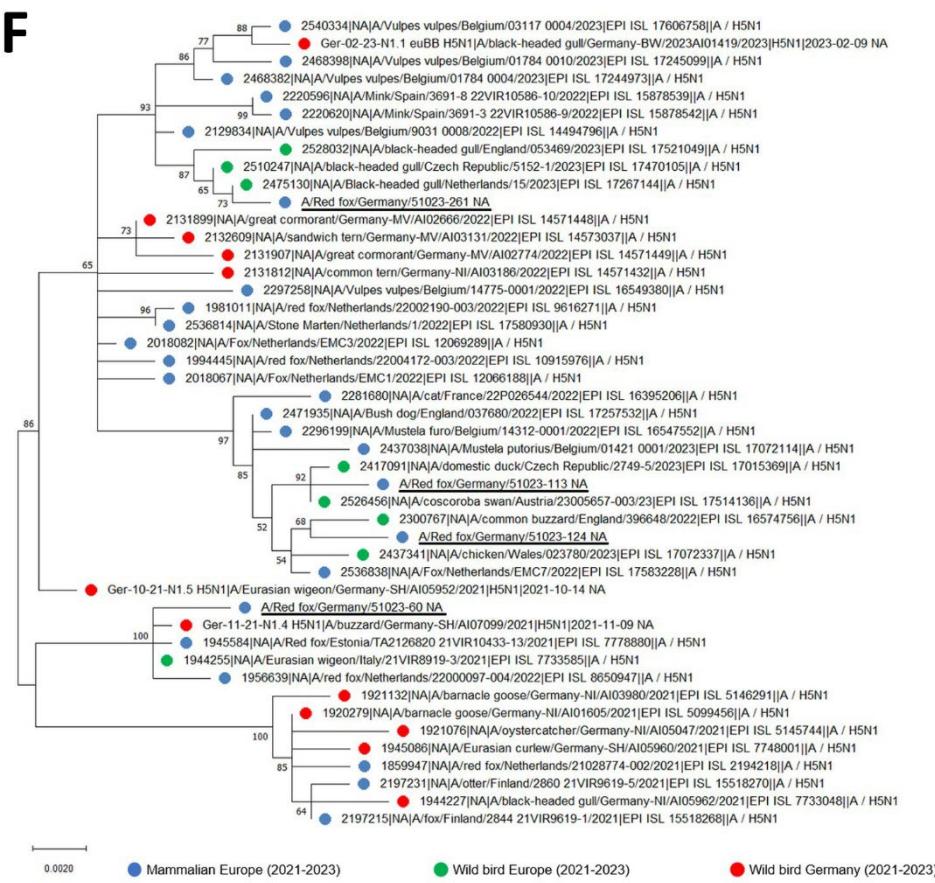
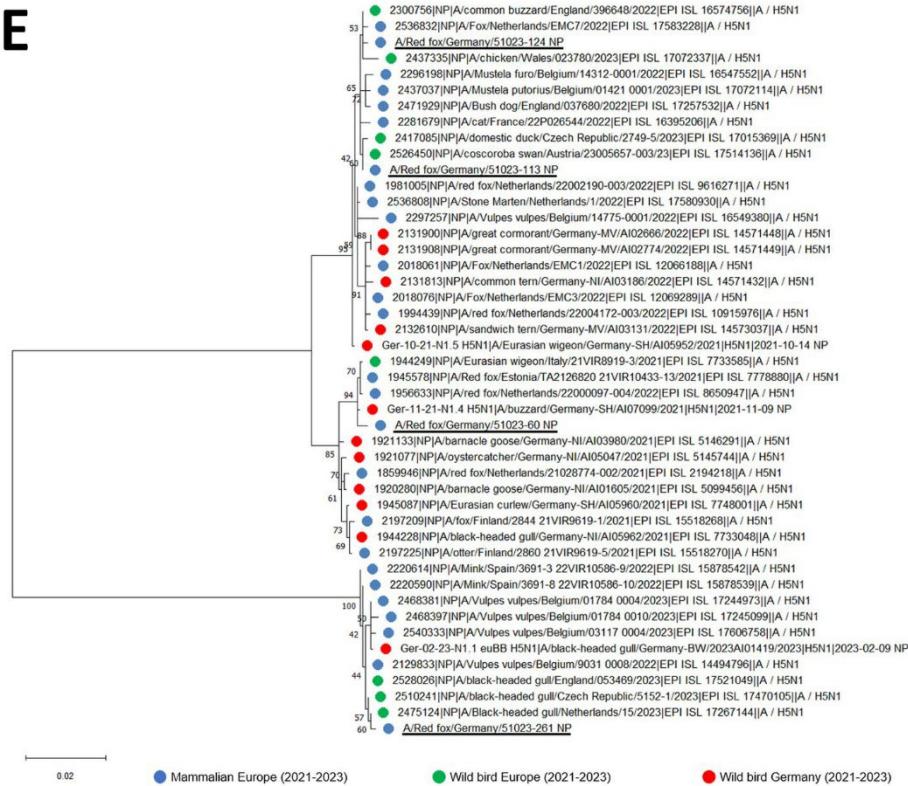
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EPI_ISL_17257532	United Kingdom	2022-Nov-18	A/Bush_dog/England/037680/2022	Animal and Plant Health Agency (APHA)	Animal and Plant Health Agency (APHA)	
EPI_ISL_16547552	Belgium	2022-Dec-08	A/Mustela_furo/Belgium/14312-0001/2022	Sciensano - Animal Infectious Diseases	Sciensano, Department of Animal Infectious Diseases	Van Borm, Steven; Roupie, Virginie; Hostyn, Pierre; Lambrecht, Benedicte; Steensels, Mieke
EPI_ISL_17072114	Belgium	2023-Feb-09	A/Mustela_putorius/Belgium/01421_0001/2023	Sciensano - Animal Infectious Diseases	Sciensano, Department of Animal Infectious Diseases	Van Borm, Steven; Roupie, Virginie; Hostyn, Pierre; Mathijs, Elisabeth; Lambrecht, Benedicte; Steensels, Mieke
EPI_ISL_7778880	Estonia	2021-Nov-08	A/Red_fox/Estonia/TA2126820_21VIR10433-13/2021	Estonian Veterinary and Food Laboratory	Istituto Zooprofilattico Sperimentale Delle Venezie	Nurmoja, I.; Vilem, A.; Juurik, T.; Zecchin, B.; Fusaro, A.; Schivo, A.; Salviato, A.; Palumbo, E.; Milani, A.; Giussani, E.; Pastori, A.; Monne, I.; Terregino, C.
EPI_ISL_11725994	Germany	2021-Nov-09	A/buzzard/Germany-SH/AI07099/2021	Landeslabor Schleswig-Holstein Wageningen Bioveterinary Research	Friedrich-Loeffler-Institut	
EPI_ISL_8650947	Netherlands	2022-Jan-03	A/red fox/Netherlands/22000097-004/2022		Wageningen Bioveterinary Research	Beerens, Nancy; Harders, Frank; Pritz-Verschuren, Sylvia; Roose, Marit; Venema, Sandra; Germeraad, Evelien; Engelsma, Marc; Heutink, Rene
EPI_ISL_16618971	Russian Federation	2022-Oct-12	A/domestic goose/Magadan/14-9V/2022	State Research Center of Virology and Biotechnology (VECTOR)	State Research Center of Virology and Biotechnology (VECTOR)	
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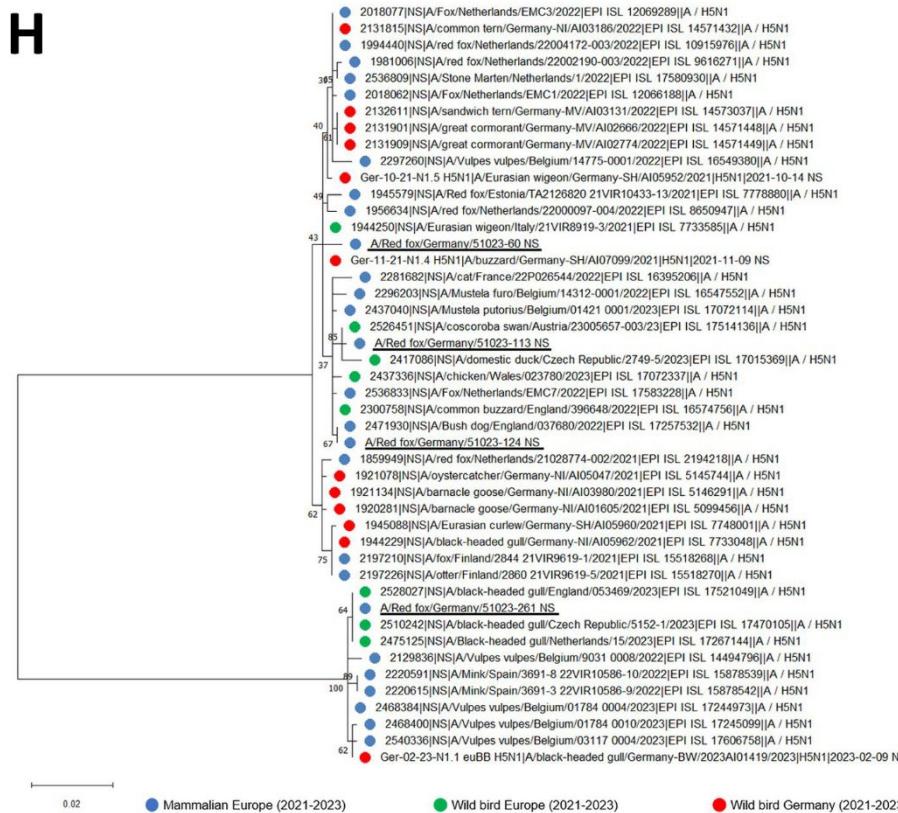
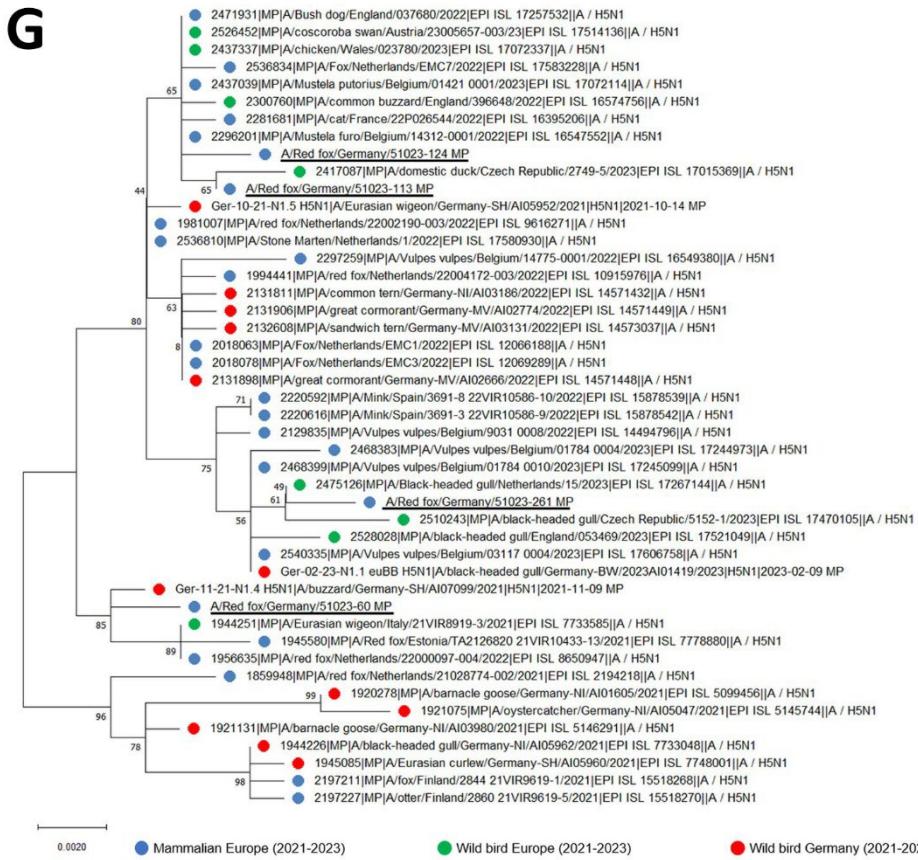
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EPI_ISL_15518270	Finland	2021-Sep-01	A/otter/Finland/2860_21VIR9619-5/2021	Finnish Food Authority	Istituto Zooprofilattico Sperimentale Delle Venezie	Tammiranta, N.;Kantala, T.;Laamanen, I.;Gadd, T.;Zecchin, B.;Fusaro, A.;Schivo, A.;Salviato, A.;Palumbo, E.;Milani, A.;Giussani, E.;Pastori, A.;Monne, I.;Terregino, C.
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EPI_ISL_7733585	Italy	2021-Oct-28	A/Eurasian_wigeon/Italy/21VIR8919-3/2021	Istituto Zooprofilattico Speriment. Lombardia, Emilia Ro. IZSLER	Istituto Zooprofilattico Sperimentale Delle Venezie	Zecchin, B.; Fusaro, A.; Schivo, A.; Salviato, A.; Palumbo, E.; Milani, A.; Giussani, E.; Pastori, A.; Monne, I.; Terregino, C.
EPI_ISL_17880904	Germany	2023-Jan-09	A/Red fox/Germany/51023-60/2023	Lower Saxony State Office for Consumer Protection and Food Safety	Lower Saxony State Office for Consumer Protection and Food Safety	This study
EPI_ISL_17880908	Germany	2023-Jan-31	A/Red fox/Germany/51023-113/2023	Lower Saxony State Office for Consumer Protection and Food Safety	Lower Saxony State Office for Consumer Protection and Food Safety	This study
EPI_ISL_17880911	Germany	2023-Feb-09	A/Red fox/Germany/51023-124/2023	Lower Saxony State Office for Consumer Protection and Food Safety	Lower Saxony State Office for Consumer Protection and Food Safety	This study
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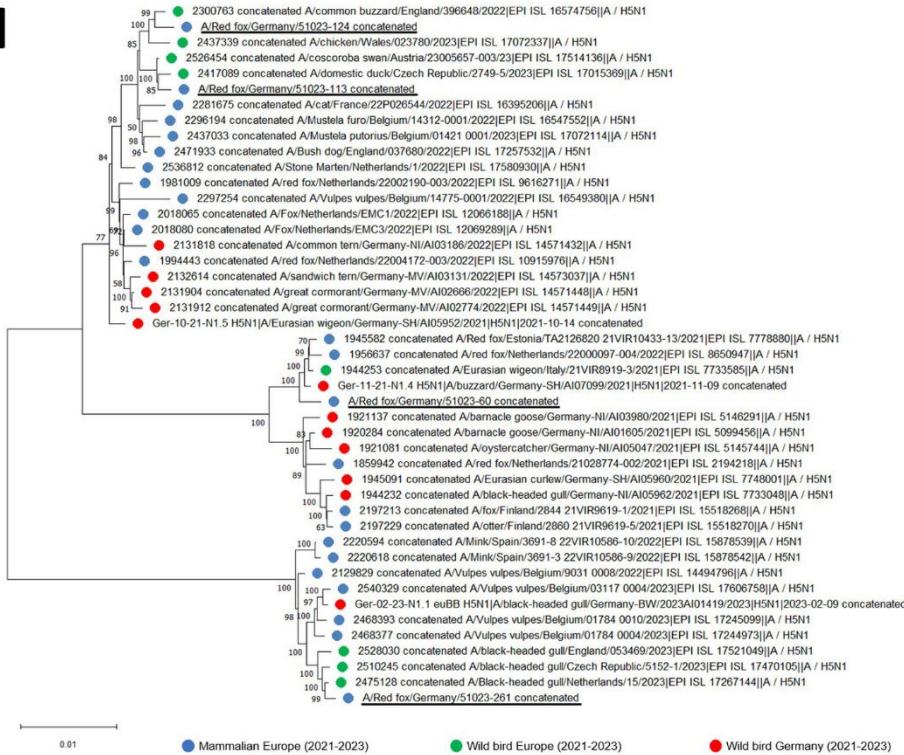
*We gratefully acknowledge the authors, originating and submitting laboratories of the sequences from GISAID's EpiFlu Database on which this research is based. All submitters of data may be contacted directly via www.gisaid.org. Sequences obtained in this study are listed at the end of this table.











Appendix Figure. Phylogenetic maximum-likelihood analyses of all 8 HPAIV H5 segmental sequences and concatenated genomes of mammals and wild birds with 500 bootstrap iterations. Virus variants found in this study are underlined. Numbers along branches indicate percentage bootstrap values. Scale bars indicate nucleotide substitutions per site. A) polymerase basic 2; B) polymerase basic 1; C) polymerase acidic; D) hemagglutinin; E) nucleoprotein; F) neuraminidase; G) matrix; H, nonstructural protein; I) concatenated.