# Two Autochthonous Cases of Anaplasmosis, Washington, USA, 2022–2023

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We describe 2 cases of autochthonous human anaplasmosis in Washington, USA, where anaplasmosis has been rarely reported. Clinicians should consider anaplasmosis in the differential diagnosis for patients with compatible clinical symptoms after tick bite or time spent outdoors in an area where *Ixodes pacificus* ticks are present.

naplasmosis is a tickborne disease caused by Anaplasma phagocytophilum, a bacterium spread by some *Ixodes* spp. ticks (1). *I. pacificus*, the western black-legged tick, is the primary vector for A. phagocytophilum on the West Coast of the United States (2). Most patients with anaplasmosis experience moderate illness, including fever, malaise, headache, myalgia, nausea, vomiting, or diarrhea (3–6). The disease can progress to severe illness, with 31% of reported case-patients hospitalized, and rarely to death; the case-fatality rate is 0.3% in the United States (7). The risk for severe illness increases with advanced patient age, immunosuppression, and delayed diagnosis and treatment (6,7). Laboratory testing often shows transaminitis and cytopenias, including anemia, thrombocytopenia, and leukopenia (3-6). Peripheral blood smear tests may show morulae in granulocytes during acute illness, but that test is not a sensitive method

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for diagnosis (4,8). Definitive diagnosis relies on molecular testing, immunohistochemistry, or culture. Serologic testing is less specific (4).

The seasonality of human anaplasmosis cases in the United States coincides with vector activity. Nymphal *Ixodes* spp. ticks are active during March through early July; most anaplasmosis cases in the United States occur during May–August. A smaller peak in cases takes place during October–November, when adult *Ixodes* ticks are active (9).

In Washington, USA, the range of *I. pacificus* ticks encompasses western Washington and the eastern slopes of the Cascade mountains (10). Tick surveillance conducted by the Washington State Department of Health during 2011–2017 identified *A. phagocyto-philum* in *I. pacificus* and *I. spinipalpis* ticks (2). Despite detection of *A. phagocytophilum* in ticks in the state, reports of autochthonous cases are rare but have been documented in canines with no recent travel outside of western Washington (11). We describe 2 reported human cases of autochthonous anaplasmosis in Washington.

#### The Case-Patients

In July 2022, an 81-year-old man (case-patient 1) visited an urgent care center with symptoms of fever (starting that day), shortness of breath, and dizziness. His medical history was notable for paroxysmal atrial fibrillation, congestive heart failure with dyspnea on exertion, pulmonary embolism, hypertension, dyslipidemia, and stroke. Bloodwork revealed thrombocytopenia (78,000 platelets/ $\mu L$ ), reference range 140,000–400,000 platelets/ $\mu L$ ), elevated aspartate aminotransferase (58 U/L; reference 10–35 U/L), and acute kidney injury with elevated blood urea nitrogen (32 mg/dL; reference 7–25 mg/dL) and creatinine

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(1.51 mg/dL; reference 0.70–1.22 mg/dL). Treating physicians discharged the patient with instructions to return for care if his symptoms worsened.

The patient sought treatment at an emergency department 2 days later for worsening shortness of breath, fatigue, weakness, and fever reaching 103°F. His platelet count had worsened to 44,000 platelets/µL, and his aspartate aminotransferase level was 86 U/L. A computed tomography angiogram identified mild cardiomegaly and trace bilateral pleural effusion but no pulmonary embolism. Computed tomography of the abdomen and pelvis revealed unremarkable results, and results of a respiratory virus panel (Biofire Respiratory 2.1 Panel, https://www.biofiredx.com) were negative. The patient received a dose of ceftriaxone and was admitted to the hospital. Consultation with infectious disease specialists prompted blood sample collection for tickborne disease testing, and physicians initiated empiric doxycycline 3 days after admission. The patient showed improvement in platelet count and liver function on that day and was discharged. Five days after discharge, the patient's

blood sample results returned positive for *A. phago-cytophilum* by qualitative real-time PCR conducted at a commercial laboratory, and physicians prescribed a continued 10-day course of doxycycline. His symptoms resolved ≈1 month after discharge.

The commercial laboratory reported the patient's PCR results to the local health jurisdiction, who forwarded the sample to the Centers for Disease Control and Prevention (Atlanta, GA, USA), where real-time PCR and sequence analysis confirmed A. phagocytophilum. Upon interview, the patient reported no travel outside of Washington state and no tick detections or tick bites during the exposure period (5-21 days before symptom onset). Three weeks before symptom onset, the patient did visit Mason County, Washington, for several days and performed yard work in an area where a neighbor recently reported a tick bite. Public health officials presumed that location to be the patient's likely exposure location. Environmental investigation did not begin until April 2023, because nymphal ticks are not likely to be active in August; however, limited drag sampling in April revealed no ticks for collection.



**Figure.** Geographic data from a study of 2 autochthonous cases of anaplasmosis, Washington, USA, 2022–2023. Blue indicates counties where health investigators have reported *I. pacificus* ticks (*12*); dark blue indicates counties of likely exposure for the 2 cases reported.

In June 2023, a woman (case-patient 2) residing in Washington began experiencing fever, back pain, neck stiffness, and headache. The woman had a history of psoriasis and hypothyroidism, managed with levothyroxine and a topical corticosteroid. Four days after symptom onset, she visited an urgent care center, where peripheral blood smear testing showed evidence of circulating, unidentified atypical cells and neutropenia, suggesting an infectious disease process or leukemia. Attending physicians discharged the patient with instructions to seek care if symptoms worsened. The woman visited an emergency department 2 days later, reporting fever, severe headache, back pain, and neck stiffness. Physicians admitted her to the hospital, where bloodwork revealed leukopenia (leukocytes 2,540/μL; reference range 4,000–12,000/μL), thrombocytopenia (platelets 96,000/µL reference 150,000–450,000/µL), anemia (erythrocytes 3.5 million/µL; reference 4-5.5 million/µL), and transaminitis (alanine transaminase 691 IU/L, reference 6-60 IU/L; aspartate aminotransferase 786 IU/L, reference 5-40 IU/L; alkaline phosphatase 435 IU/L, reference 28-126 IU/L). Computed tomography showed hepatosplenomegaly but no other abnormalities, and additional serologic testing showed negative results for cytomegalovirus, hepatitis (A, B, and C), HIV, and *Toxoplasma*. A respiratory virus panel (VERIGENE Respiratory Pathogens Flex Test, https://us.diasorin.com) and 2 sets of blood cultures revealed negative results. The patient tested positive for group A Streptococcus on throat culture and had low-level viremia with Epstein-Barr virus, although antibody testing was negative. An infectious disease specialist consulted on the day of admission recommended an extensive work-up for viral and tickborne diseases, and attending physicians initiated a course of cefepime and doxycycline the next day. A blood sample collected the day after admission tested positive for A. phagocytophilum via real-time PCR on day 6 of hospitalization, and physicians prescribed a continuing regimen of doxycycline for 10 days. Thrombocytopenia and leukopenia resolved 4 days after starting doxycycline, and physicians discharged the patient 10 days after admission, noting gradual improvement in symptoms.

The hospital laboratory reported the real-time PCR results for this patient to officials at the local health jurisdiction, who interviewed the patient. The patient reported hiking in multiple parks in Pierce County, but no travel outside of Washington during the exposure period. No environmental investigation took place because the patient did not report a tick bite and researchers could identify no single exposure location.

#### Conclusion

Washington is a low incidence region for tickborne diseases (2), posing a challenge for public health surveillance, prevention communication, and provider education. We charted the counties in Washington with documented *I. pacificus* tick presence and the suspected counties of exposure for the 2 human cases of anaplasmosis (Figure).

Clinicians should be aware of the local presence of A. phagocytophilum and the distribution of I. pacificus ticks in Washington. I. pacificus ticks can also transmit Borrelia burgdorferi sensu stricto, the causative agent of Lyme disease, and Borrelia miyamotoi, the causative agent of hard tick relapsing fever (2). Health officials have increased tick surveillance in Washington, noting increased I. pacificus ticks activity in spring and most ticks collected during March-May (2,13). To support prompt diagnosis and treatment of suspected tickborne disease among Washington residents, clinicians should consider travel history, exposure to tick habitats, seasonality in tick activity, and the patient's clinical manifestations. More robust tick surveillance in the state could help better define vector distribution, abundance, and infection risk to inform public health prevention messaging and provider education.

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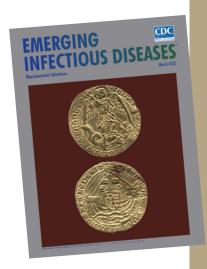
#### References

- Chen SM, Dumler JS, Bakken JS, Walker DH. Identification of a granulocytotropic *Ehrlichia* species as the etiologic agent of human disease. J Clin Microbiol. 1994;32:589–95. https://doi.org/10.1128/jcm.32.3.589-595.1994
- 2. Dykstra EA, Oltean HN, Kangiser D, Marsden-Haug N, Rich SM, Xu G, et al. Ecology and epidemiology of tickborne

- pathogens, Washington, USA, 2011–2016. Emerg Infect Dis. 2020;26:648–57. https://doi.org/10.3201/eid2604.191382
- Bakken JS, Krueth J, Wilson-Nordskog C, Tilden RL, Asanovich K, Dumler JS. Clinical and laboratory characteristics of human granulocytic ehrlichiosis. JAMA. 1996;275:199–205. https://doi.org/10.1001/jama.1996. 03530270039029
- Biggs HM, Behravesh CB, Bradley KK, Dahlgren FS, Drexler NA, Dumler JS, et al. Diagnosis and management of tickborne rickettsial diseases: Rocky Mountain spotted fever and other spotted fever group rickettsioses, ehrlichioses, and anaplasmosis – United States. MMWR Recomm Rep. 2016;65:1–44. https://doi.org/10.15585/mmwr.rr6502a1
- Bakken JS, Krueth J, Wilson-Nordskog C, Tilden RL, Asanovich K, Dumler JS. Clinical and laboratory characteristics of human granulocytic ehrlichiosis. JAMA. 1996;275:199–205. https://doi.org/10.1001/ jama.1996.03530270039029
- Bakken JS, Dumler S. Human granulocytic anaplasmosis. Infect Dis Clin North Am. 2008;22:433–48, viii. https://doi.org/10.1016/j.idc.2008.03.011
- Dahlgren FS, Heitman KN, Drexler NA, Massung RF, Behravesh CB. Human granulocytic anaplasmosis in the United States from 2008 to 2012: a summary of national surveillance data. Am J Trop Med Hyg. 2015;93:66–72. https://doi.org/10.4269/ajtmh.15-0122
- Schotthoefer AM, Meece JK, Ivacic LC, Bertz PD, Zhang K, Weiler T, et al. Comparison of a real-time PCR method with serology and blood smear analysis for diagnosis of

- human anaplasmosis: importance of infection time course for optimal test utilization. J Clin Microbiol. 2013;51:2147–53. https://doi.org/10.1128/JCM.00347-13
- Centers for Disease Control and Prevention.
   Anaplasmosis epidemiology and statistics [cited 2023 Oct 10]. https://www.cdc.gov/anaplasmosis/hcp/statistics/index.html
- Eisen RJ, Eisen L, Beard CB. County-scale distribution of *Ixodes scapularis* and *Ixodes pacificus* (Acari: Ixodidae) in the continental United States. J Med Entomol. 2016;53:349–86. https://doi.org/10.1093/jme/tjv237
- Poitout FM, Shinozaki JK, Stockwell PJ, Holland CJ, Shukla SK. Genetic variants of *Anaplasma phagocytophilum* infecting dogs in western Washington State. J Clin Microbiol. 2005;43:796–801. https://doi.org/10.1128/ JCM.43.2.796-801.2005
- Centers for Disease Control and Prevention. CDC tick surveillance data sets. December 2023 [cited 2024 Jan 4]. https://www.cdc.gov/ticks/surveillance/ TickSurveillanceData.html
- 13. Washington State Department of Health. Washington tracking network—tick data dashboard. [cited 2023 Oct 10]. https://doh.wa.gov/data-and-statistical-reports/washington-tracking-network-wtn/tick-data/tick-dashboard

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## etymologia revisited

### Schizophyllum commune

[skiz-of'-i'-ləm kom'-yoon]

Schizophyllum commune, or split-gill mushroom, is an environmental, wood-rotting basidiomycetous fungus. Schizophyllum is derived from "Schiza" meaning split because of the appearance of radial, centrally split, gill-like folds; "commune" means common or shared ownership or ubiquitous. Swedish mycologist Elias Magnus Fries (1794–1878) the Linnaeus of Mycology assigned the scientific name in 1815. German mycologist Hans Kniep in 1930 discovered its sexual reproduction by consorting and recombining genomes with any one of numerous compatible mates (currently >2,800).

#### References

- 1. Chowdhary A, Kathuria S, Agarwal K, Meis JF. Recognizing filamentous basidiomycetes as agents of human disease: a review. Med Mycol. 2014;52: 782–97. https://doi.org/10.1093/mmy/myu047
- Cooke WB. The genus Schizophyllum. Mycologia. 1961;53:575–99. https://doi.org/10.1080/00275514.1961.12017987
- Greer DL. Basidiomycetes as agents of human infections: a review. Mycopathologia. 1978;65:133–9. https://doi.org/10.1007/BF00447184
- 4. O'Reilly P. *Schizophyllum commune*, split gill fungus, 2016 [cited 2021 Aug 23]. https://www.first-nature.com/fungi/schizophyllum-commune.php
- Raper CA, Fowler TJ. Why study *Schizophyllum?* Fungal Genet Rep. 2004;51:30–6. https://doi.org/10.4148/1941-4765.1142

https://wwwnc.cdc.gov/eid/article/28/3/et-2803\_article