RESEARCH LETTERS

D.A.S. was supported by the Boehringer Ingelheim Veterinary Scholars Program through the Veterinary Medical Scientist Research Training Program at the Texas A&M University College of Veterinary Medicine and Biomedical Sciences.

About the Author

Ms. Sierra is a DVM student at Texas A&M University College of Veterinary Medicine and Biomedical Sciences and has a strong interest in zoologic medicine.

References

- Cowie RH, Ansdell V, Panosian Dunavan C, Rollins RL. Neuroangiostrongyliasis: global spread of an emerging tropical disease. Am J Trop Med Hyg. 2022;107:1166–72. https://doi.org/10.4269/ajtmh.22-0360
- da Silva AJ, Morassutti AL. Angiostrongylus spp. (Nematoda; Metastrongyloidea) of global public health importance. Res Vet Sci. 2021;135:397–403. https://doi.org/10.1016/ j.rvsc.2020.10.023
- 3. Ansdell V, Kramer KJ, McMillan JK, Gosnell WL, Murphy GS, Meyer BC, et al. Guidelines for the diagnosis and treatment of neuroangiostrongyliasis: updated recommendations. Parasitology. 2021;148:227–33. https://doi.org/10.1017/S0031182020001262
- Flerlage T, Qvarnstrom Y, Noh J, Devincenzo JP, Madni A, Bagga B, et al. *Angiostrongylus cantonensis* eosinophilic meningitis in an infant, Tennessee, USA. Emerg Infect Dis. 2017;23:1756–8. https://doi.org/10.3201/eid2310.170978
- Gottdenker NL, Nascimento Ramos RA, Hakimi H, McHale B, Rivera S, Miller BM, et al. Angiostrongylus cantonensis infection in brown rats (Rattus norvegicus), Atlanta, Georgia, USA, 2019–2022. Emerg Infect Dis. 2023;29:2167–70. https://doi.org/10.3201/eid2910.230706
- Edwards EE, Borst MM, Lewis BC, Gomez G, Flanagan JP. *Angiostrongylus cantonensis* central nervous system infection in captive callitrichids in Texas. Vet Parasitol Reg Stud Reports. 2020;19:100363. https://doi.org/10.1016/j.vprsr.2019.100363
- Stockdale Walden HD, Slapcinsky JD, Roff S, Mendieta Calle J, Diaz Goodwin Z, Stern J, et al. Geographic distribution of Angiostrongylus cantonensis in wild rats (Rattus rattus) and terrestrial snails in Florida, USA. PLoS One. 2017;12:e0177910. https://doi.org/10.1371/journal.pone.0177910
- Rael RC, Peterson AC, Ghersi-Chavez B, Riegel C, Lesen AE, Blum MJ. Rat lungworm infection in rodents across post-Katrina New Orleans, Louisiana, USA. Emerg Infect Dis. 2018;24:2176–83. https://doi.org/10.3201/eid2412.180056
- Foster CE, Nicholson EG, Chun AC, Gharfeh M, Anvari S, Seeborg FO, et al. *Angiostrongylus cantonensis* infection: a cause of fever of unknown origin in pediatric patients. Clin Infect Dis. 2016;63:1475–8. https://doi.org/10.1093/cid/ciw606
- Jarvi SI, Quarta S, Jacquier S, Howe K, Bicakci D, Dasalla C, et al. High prevalence of Angiostrongylus cantonensis (rat lungworm) on eastern Hawai'i Island: a closer look at life cycle traits and patterns of infection in wild rats (*Rattus* spp.). PLoS One. 2017;12:e0189458. https://doi.org/10.1371/ journal.pone.0189458

Address for correspondence: Guilherme G. Verocai, Department of Veterinary Pathobiology, Texas A&M Veterinary Medicine & Biomedical Sciences, 4467 TAMU, College Station, TX 77843-4467, USA; email: gverocai@cvm.tamu.edu

Human Babesiosis Caused by *Babesia venatorum*, Russia, 2024

Olga P. Zelya, Irina V. Kukina, Ludmila S. Karan, Elena A. Krasilovskaya, Vadim V. Garin

Author affiliations: First Moscow State Medical University (Sechenov University), Moscow, Russia (O.P. Zelya, I.V. Kukina, V.V. Garin); Research Center of Neurology, Moscow (L.S. Karan); Centre for Strategic Planning of the Federal Medical and Biological Agency, Moscow (L.S. Karan); Sayanogorsk Interdistrict Hospital, Sayanogorsk, Russia (E.A. Krasilovskaya)

DOI: http://doi.org/10.3201/eid3109.250319

We report a case of acute babesiosis in a splenectomized 63-year-old man in Siberia, Russia. We confirmed the causative agent, *Babesia venatorum*, by PCR. Our study demonstrated a change in the structure of the parasite population, from single parasite invasion of erythrocytes to multioccupancy, without an increase in parasitemia level.

abesiosis is an emerging tickborne infection Dcaused by intraerythrocytic protozoa. To date, researchers have described more than 50 cases of babesiosis in humans in Europe, almost always fulminant in splenectomized patients and typically attributed to Babesia divergens. Some recent reports also describe several cases of human infection with B. venatorum, associated with milder infections than those caused by B. divergens (1). Researchers have also described sporadic cases of babesiosis caused by infection with Babesia microti and B. divergens in Asia-Pacific regions (2,3), but the practically asymptomatic course of the human infection with *B. venatorum* is more common (4). Although reports have noted detection of *Babesia* spp. DNA in *Ixodes persulcatus* ticks in Siberia (5), cases of human infections have yet to be reported in that region of Russia.

We report the case of a 63-year-old man who resided in a forested, mountainous area of Khakassia, East Siberia, Russia, and had undergone splenectomy. On September 30, 2024, the man sought treatment for an influenza-like syndrome with signs and symptoms that included a fever of 38°C, severe general weakness, darkening of urine, a decrease in diuresis, jaundice, dyspnea, and stomachache. Attending physicians admitted the patient to the hospital on October 2, 2024 (Table). The patient reported no awareness of a tick bite and had received no blood products in the previous 3 months.

Table. Characteristics of case-patient in study of human babesiosis caused by Babesia venatorum, Russia, 2024*

	Results by date						
Tests performed	Oct 3	Oct 7	Oct 10	Oct 14	Oct 22	Nov 8	Reference range
Hematologic parameters							_
Hemoglobin, g/dL	11.2	9.3	11.6	11.6	12.0	15.5	11.7–18.0
Hematocrit, %	32.0	27.0	34.8	32.7	36.1	46.8	35–52
Erythrocytes, ×10 ¹² cells/L	3.48	3.01	3.96	3.51	3.85	5.1	3.8-6.1
RBCN, L	10:100	4:100	7:100	NA	NA	NA	0:100
Platelets, ×10 ⁹ /L	21	127	154	346	230	217	150-450
Leukocytes, ×109 cells/L	4.8	8.3	7.8	5.7	9.1	10.4	4–11
Parasitemia level, %†	2.43	2.91	1.56	0	0	0	0
Biochemistry							
Alanine aminotransferase, U/L	7.4	22.0	NA	21.0	16.0	NA	≤40
Aspartate aminotransferase, U/L	27.0	67.0	NA	25.0	30.0	NA	≤40
Total bilirubin, µmol/L	84.9	38.1	NA	11.8	9.0	NA	3.1-16.9
C-reactive protein, mg/L	1.3	0.7	NA	1.6	4.0	NA	0-0.5
Urea, μmol/L	na	7.5	NA	3.3	6.0	NA	2.5-8.3
Creatinine, mcM/L	97.0	91.0	NA	111.0	107.0	NA	53-106
Glucose, µmol/L	25.6	14.6	NA	8.4	7.5	NA	3.3-5.5
Total protein, g/L	56.0	55.0	NA	68.5	93.0	NA	62–81

*NA, not available; RBCN, red blood cell normoblasts.

†Percentage of erythrocytes that were infected.

Blood smears obtained on hospital admission tested positive for *Plasmodium* spp. However, because the patient reported no recent travel to malaria-endemic areas, we sent the blood samples for retesting at Sechenov University (Moscow, Russia), where results confirmed babesiosis. We then examined the blood smears after Romanovsky-Giemsa staining, noting the paired forms diverging at a wide angle (≤180°), which is characteristic of both *B. divergens* and *B. venatorum* (Appendix Figure 1, https://wwwnc.cdc.gov/EID/article/31/9/25-0319-App1.pdf).

We screened DNA samples extracted from the blood smears for *B. divergens*, *B. microti*, and *B. venatorum* by PCR, using methods described previously (6). We partially sequenced the 18S rRNA gene (1,112 bp; GenBank accession no. PV086113) (5). We aligned, compared, and analyzed the resulting nucleotide sequences and reference sequences downloaded from GenBank by using MEGA X (7). We also reconstructed a phylogenetic tree (Appendix Figure 2). Using forward and reverse primers form 18S RNA of *Babesia*

spp. from Europe, we were able to detect only *B. venatorum* DNA.

We started etiotropic treatment for the patient 3 days from the time we initially detected intraerythrocytic parasites and identified the causative agent as Babesia spp. The patient responded to therapy (clindamycin and quinine sulfate), and by day 9 of treatment, parasites were no longer detectable with microscopy (Table). We noted that the ratio of erythrocytes invaded by >1 trophozoite changed as the infection progressed. Four days after symptom onset, single trophozoites and pairs (figure 8 pattern) predominated. On the 8th day of infection, with the same level of parasitemia, almost half of the infected erythrocytes contained ≥4 trophozoites (Figure). Parasites continued to divide intensively but did not leave the host cell. Prior research has noted multiple parasites present in individual erythrocytes during fulminant infections in humans (8) and in heavily infected in vitro cultures (9). Results of such studies suggests that multioccupancy of trophozoites in the erythrocyte

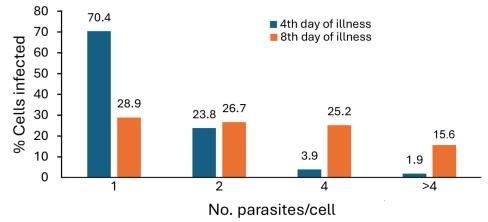


Figure. Dynamics of the parasite population in erythrocytes in case-patient in study of human babesiosis caused by *Babesia venatorum*, Russia, 2024. As the infection progressed with constant parasitemia (<3%), 4 main populations of infected red blood cells were seen: 1 parasite, 2 parasites, 4 parasites, and >4 multioccupancy (>4 parasites).

RESEARCH LETTERS

prevents a sharp increase in parasitemia and helps to preserve the parasite population in the host (9). However, the phenomenon we observed occurred at both high (>23%) and low (<3%) parasitemia.

Our study of this unique case of human babesiosis in Siberia, Russia, provides molecular evidence that the etiologic agent was *B. venatorum*. Prior research noted the Asia variant of *B. venatorum* in asymptomatic persons from China (4). *B. divergens* infection in splenectomized humans could lead to death, even with timely treatment (1). However, the similar course and positive outcome of babesiosis in our patient resembled those in cases previously reported in Europe (1), suggesting that the causative agent of the disease was *B. venatorum*, which we confirmed by PCR.

Our patient resided in a village located in a forested area, which is a natural habitat for ticks. The man's work involved staying in the forest, again increasing his risk for tick bites. The fact that the patient did not notice a tick bite is not unusual. Only 50% to 70% of patients with tickborne diseases recall being bitten by a tick (2). Considering the ability of *B. venatorum* to be transmitted transovarially and transstadially within *I. persulcatus* ticks, it is possible for a person to become infected through the bite of a tick nymph, which is smaller and less noticeable than the adult (10).

In conclusion, our case study revealed the potential risk of *B. venatorum* infection for persons living in Siberia, Russia. Clinicians should be aware that infection can occur as an influenza-like illness and may go unnoticed in immunocompetent persons.

Acknowledgment

We are deeply grateful to Svetlana L. Lukovenko for her collaboration in preparing and sending us blood products (thick and thin blood films) with intraerythrocytic protozoa.

About the Author

Dr. Zelya is a parasitologist working as an assistant professor at Martsinovsky Institute of Medical

Parasitology, Tropical and Vector-Borne Diseases, Sechenov University, Moscow, Russia. Her primary research interests are centered on laboratory diagnostics and epidemiology of parasitic diseases.

References

- Hildebrandt A, Zintl A, Montero E, Hunfeld K-P, Gray J. Human babesiosis in Europe. Pathogens. 2021;10:1165. https://doi.org/10.3390/pathogens10091165
- Zhou X, Xia S, Huang JL, Tambo E, Zhuge HX, Zhou XN. Human babesiosis, an emerging tick-borne disease in the People's Republic of China. Parasit Vectors. 2014;7:509. https://doi.org/10.1186/s13071-014-0509-3
- 3. Kumar A, O'Bryan J, Krause PJ. The global emergence of human babesiosis. Pathogens. 2021;10:1447. https://doi.org/10.3390/pathogens10111447
- Jiang JF, Zheng YC, Jiang RR, Li H, Huo QB, Jiang BG, et al. Epidemiological, clinical, and laboratory characteristics of 48 cases of "Babesia venatorum" infection in China: a descriptive study. Lancet Infect Dis. 2015;15:196–203. https://doi.org/ 10.1016/S1473-3099(14)71046-1
- Rar VA, Epikhina TI, Livanova NN, Panov VV. Genetic diversity of Babesia in *Ixodes persulcatus* and small mammals from North Ural and West Siberia, Russia. Parasitology. 2011;138:175–82. https://doi.org/10.1017/S0031182010001162
- Michelet L, Delannoy S, Devillers E, Umhang G, Aspan A, Juremalm M, et al. High-throughput screening of tick-borne pathogens in Europe. Front Cell Infect Microbiol. 2014;4:103. https://doi.org/10.3389/fcimb.2014.00103
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K. MEGA X: Molecular evolutionary genetics analysis across computing platforms. Mol Biol Evol. 2018;35:1547–9. https://doi.org/ 10.1093/molbev/msy096
- Kukina IV, Zelya OP. Extraordinary high level of propagation of *Babesia divergens* in severe human babesiosis. Parasitology. 2022;149:1160–3. https://doi.org/10.1017/ S0031182022000439
- 9. Lobo CA, Cursino-Santos JR, Singh M, Rodriguez M. *Babesia divergens*: a drive to survive. Pathogens. 2019;8:95. https://doi.org/10.3390/pathogens8030095
- Bonnet S, Brisseau N, Hermouet A, Jouglin M, Chauvin A. Experimental in vitro transmission of *Babesia* sp. (EU1) by *Ixodes ricinus*. Vet Res. 2009;40:21. https://doi.org/10.1051/ vetres/2009004

Address for correspondence: Olga P. Zelya, Martsinovsky Institute of Medical Parasitology, Tropical and Vector-Borne Diseases, Sechenov University, Malaya Pirogovskaya st., 20-1, Moscow 119435, Russia; email: zelya_o@mail.ru