Yellow Fever in Pará State, Amazon Region of Brazil, 1998–1999: Entomologic and Epidemiologic Findings


*Instituto Evandro Chagas, FUNASA, Ministry of Health of Brazil, Belém, PA, Brazil; and †University of Texas Medical Branch, Galveston, Texas, USA

Yellow fever (YF) is frequently associated with high severity and death rates in the Amazon region of Brazil. During the rainy seasons of 1998 and 1999, 23 (eight deaths) and 34 (eight deaths) human cases of YF were reported, respectively, in different geographic areas of Pará State; most cases were on Marajó Island. Patients were 1 to 46 years of age. Epidemiologic and ecological studies were conducted in Afuá and Breves on Marajó Island; captured insects yielded isolates of 4 and 11 YF strains, respectively, from Haemagogus janthinomys pooled mosquitoes. The cases on Marajó Island in 1999 resulted from lack of vaccination near the focus of the disease and intense migration, which brought many nonimmune people to areas where infected vectors were present. We hypothesize that YF virus remains in an area after an outbreak by vertical transmission among Haemagogus mosquitoes.

Materials and Methods

Collection Sites

Mosquito and blood collections were made at three sites in Pará State (Figure 1). Afuá (0°06' S; 50°20' W; population approximately 30,000) and Breves (1°41' S, 50°19' W; population approximately 75,000) are municipalities of Marajó Island, in the northern region of Pará State, known for buffalo breeding and fish farming. The other site, Altamira (2° 51' S; 51° 57' W) (approximately 80,000 inhabitants), is in the central region of the state on the Xingu River delta near the Transamazon Highway; its chief products are wood, cattle, and sugar cane and cacao.

Samples

Blood samples were taken from persons who had clinical symptoms and signs compatible with YF for attempts at virus isolation, as well as from contacts (family and neighbors) for serologic examinations. Approximately 5 mL to 10 mL of blood was obtained by venipuncture. Serum samples were stored at -20°C until tested. From patients with fever and other clinical symptoms, specimens were also obtained for attempted virus isolation. Monkeys were euthanized, and blood samples and liver fragments obtained for virus isolation. All specimens were frozen in liquid nitrogen containers until processed in the laboratory.

Mosquitoes

Diurnal human-biting mosquitoes were collected in Afuá, Breves, and Altamira. The collections were made from 9:00 a.m. to 3:00 p.m. on the ground and at an elevation of approximately 15 m in the forest canopy, since these are the more active periods and places of potential YF virus vectors.
In Afuá, collections were made from May 17 to May 30, 1998, and from March 11 to March 25, 1999. Collections in Breves were from March 11 to March 25, 1999. In Altamira, collections were made on April 16 to April 28, May 20 to June 5, and July 10 to July 19, 1998, and from January 31 and February 13 (rainy season) and July 14 to July 27 (dry season) in 1999.

In the laboratory, mosquitoes were classified and pooled under refrigeration, by species, place, and dates of collection, and then preserved at -70°C until inoculation. Minimum infection rates (MIRs) of *Hg. janthinomys* mosquitoes were calculated by dividing the total of positive pools by the total number of specimens processed (12).

Serology

Serum samples were initially screened by hemagglutination inhibition (HI) against YF antigen (strain Be H 111). Tests were performed as described by Clarke and Casals, using a microtechnique in which serum samples were acetone extracted (13). All positive samples were later assayed by enzyme immunoassay for capture of immunoglobulin (Ig) M (MAC-ELISA) (14). All positive samples by both tests were later tested by plaque reduction neutralization test (PRNT) to confirm infection (15).

Serologic Criteria for Inclusion in Study

Positive diagnostic criteria for inclusion in our study were 1) serologic conversion by fourfold increase in antibody titers between acute- and convalescent-phase (7 to 14 days after first collection) serum samples and 2) presence of IgM without history of YF vaccination plus positive reaction (>1:10) by PRNT.

Pathology

From fatal cases, liver samples were obtained; histologic sections were stained by hematoxylin and eosin and examined by light microscopy. Specific YF antigens were detected in paraffin-embedded liver samples of fatal cases by means of an immunohistochemistry technique (16). All patients with these antigens were considered YF-positive cases and included in the study.

Virus Isolation and Identification

All samples (blood, viscera, and mosquitoes) were inoculated into suckling mice and C6/36 cells for virus isolation. Before processing, samples were thawed, triturated, and diluted in phosphate-buffered saline (pH 7.4) with 0.75% of bovine albumin and antibiotics (100 µg/mL of streptomycin and 100 IU/mL of penicillin) and centrifuged for 10 minutes at 2,100 x g (15). The supernatant of each specimen was then injected into suckling mice (0.02 mL intracerebrally) and into tubes of cells (0.1 mL), respectively. Isolated strains were identified by indirect immunofluorescence assay and complement fixation test (15). Isolation of YF virus from blood or tissues of human patients without history of vaccination was used as the positive criterion for inclusion in the study.

Results

Epidemiology

In 1998 and 1999 in Pará State, 23 and 34 YF cases, respectively, were diagnosed. In 1998, 17 of the 23 occurred in Afuá and 6 in municipalities not on Marajó Island (Bannach, Floresta, Gurupá, Itaituba, Óbidos, and Redenção). In 1999, 15 of the 34 diagnosed cases occurred in Afuá, 14 in Breves, and 5 in three other municipalities (Conceição do Araguaia, 2; Santa Maria das Barreiras, 2; Redenção, 1), all in the southeast region (Figure 1). The sex and age distributions of cases were determined (Figure 2). Comparison of total cases and deaths in both years shows a case-fatality rate of 34.8% in 1998 and 23.5% in 1999 (Figure 3). Most reported cases were diagnosed by serology or another technique combined with serology, with clinical and epidemiologic features taken into account (Figure 4).

Afuá

A scientific trip was undertaken to the municipality (May 17 to 25, 1998). From the 23 human samples, three isolations of the YF virus (H 603325, H 603327, and H 603797) were made. Of the monkey specimens, two samples of YF virus (AN 604552, AN 604555), were isolated from the blood and liver, respectively of monkey (PR 2968) of the species *Alouatta belzebul*. The YF cases occurred in several areas, among them...
the river Morego, Morceguinho, Tamanduá, Bom Jardim, and Furo da Cidade.

**Entomology**

**Afuá**

A total of 1,621 Culicidae were collected with human bait during the day in the canopy and at ground level, which after identification, provided 77 pools for inoculation. The most abundant species were *Wyeomyia* sp and *Hg. janthinomys*, with 1,119 (69%) and 296 (18.3%) specimens, respectively; after identification, these formed 23 and 14 pools, respectively, for virus isolation in tissues. Four samples of YF virus were isolated from pools of *Hg. janthinomys* (AR 605158, AR 605159, AR 605160, and AR 605161). The MIR for *Hg. janthinomys* was 1.35%.

**Altamira**

Deaths of monkeys in the forest on agricultural secondary roads of the Transamazon Highway in the stretch from Altamira to Marabá (km 20 and 27) motivated a scientific expedition from April 16 to 28, 1998. A total of 592 hematophagous insects were collected on human bait; 479 (80.9%) were the mosquito *Hg. janthinomys*, which after identification provided 38 and 24 lots, respectively, for virus research. The pools inoculated yielded 10 isolations of YF virus, all from *Hg. janthinomys* (MIR = 2.01%).

The high infection rate with YF virus observed in *Hg. janthinomys* motivated a second trip (May 20-June 5) to determine the spatial distribution and infection rate of *Hg. janthinomys*, as well as to evaluate the dynamics of the circulation of YF virus in the Altamira area. A total of 509 hematophagous diptera (66 lots) were captured; 312 (61.3%) belonged to the species *Hg. janthinomys*. Three samples were identified as YF virus, as occurred in the first trip; all positive samples were from pools of *Hg. janthinomys*, for an MIR of 0.96%.

From July 10 to July 19 (dry season), a third trip was made to the same area of Altamira to monitor circulation of YF virus. In this trip, 120 hematophagous insects were captured in 14 lots; 28 (23.3%) of them were *Hg. janthinomys* from a single lot. Injection of these lots into newborn mice did not produce virus.

In the rainy season (January 31 to February 13) of 1999, 1,105 (93 pools) mosquitoes were captured; 84 (5 pools) were *Hg. janthinomys*. During the dry season (July 14 to 27), 133 mosquitoes (14 pools) were collected; 44 (3 pools) were *Hg. janthinomys*. No virus was isolated.

**Afuá/Breves**

The occurrence of human cases motivated the expedition to Afuá and Breves to carry out entomologic studies of potential vectors of YF virus. From March 11 to March 25, 1999, captures of hematophagous insects were performed at ground level and in the forest canopy. A total of 2,164 insects were collected on human bait in 126 pools; 546 of these were *Hg. janthinomys*, which furnished 23 pools for inoculation in attempts at virus isolation. Eleven strains of YF virus were obtained from pools of *Hg. janthinomys*, for an MIR of 2.01%. No virus was isolated from the other mosquito pools. During this trip, three howler monkeys (*Alouatta belzebul*) were euthanized. The specimens from the monkeys produced three
YF virus isolates, two from the blood and liver of the same monkey and another from the liver of a second monkey. These monkeys were found within 200 m to 500 m of human dwellings. They had been showing abnormal behavior, i.e., moving slowly and not trying to escape from people.

Discussion

In 1998, in Afuá municipality, the first YF human case (based on epidemiologic information) had onset of symptoms on February 3. As the YF medium incubation period ranges from 3 to 6 days, infection probably occurred at the end of January. Our trip to the municipality was in May, >3 months after the index case. Despite the long interval between the index case and our expedition, we recovered YF virus from pools of Haemagogus janthinomys mosquitoes and from howler monkeys. These findings strongly suggest an elevated natural average infection rate of the vector mosquitoes in the area. This was the first detection of YF virus in the municipality of Afuá. YF virus has not been reported on Marajó Island since 1988, when a sporadic case occurred near Breves in a man who cut down a tree.

The study in Altamira in 1998 shows clearly how YF virus outbreaks happen. The rainy season, in the first months of the year, has the highest rainfall indexes in the Amazon forest region. This facilitates breeding of mosquitoes, including the potential vector Haemagogus janthinomys, in the forest. When the rains decrease, the amount of mosquitoes in the forest gradually decreases (Table). As the population of mosquito vectors decreases, YF virus disappears.

In Breves and Afuá in 1999, reported cases clearly resulted from a failure of the vaccination campaign, since after the outbreak in 1998, the inhabitants of Afuá and Breves were vaccinated. However, some people from Afuá were not immunized, and they migrated to areas near places where they had been vaccinated. The occurrence of these immunized, and they migrated to areas near places where the outbreaks happened. Therefore, appearance of cases is a direct function of migration of nonimmune persons. In places such as Altamira, where people show high YF vaccination rates, it is quite difficult to find a human case, despite the municipality’s situation in the endemic area with virus circulation.

The occurrence of YF cases in other areas at the same time or within a short time period also supports our hypothesis. Since some municipalities are located >1,500 km from Afuá and Breves municipalities (Figure 1), YF virus must be present there; when nonimmune people enter the forest, they become infected. Therefore, appearance of cases is the result of a silent restricted circulation of the virus in an area’s forest.

On the other hand, YF cases in South America have thus far only been transmitted by sylvatic vectors, especially Haemagogus janthinomys (3,11,21). The susceptibility of the Aedes aegypti population in South America to YF virus must be established, in the face of the increased risk of reemergence of urban transmission (22-24). The annual occurrence of several cases in Brazil and hundreds of them in Peru and Bolivia may have permitted contact of YF virus with human mammals (probably Internet based) to keep all countries in the continent quickly informed of YF cases and outbreaks, especially in major risk areas.

Table. Comparison of number of mosquitoes collected on human bait, number of YF strains isolated by place of capture, and minimum infection rate (MIR) for Haemagogus janthinomys, Pará State, Brazil, 1998–1999

<table>
<thead>
<tr>
<th>Place</th>
<th>Year</th>
<th>Strains isolated</th>
<th>Hg. janthinomys baited</th>
<th>MIR</th>
<th>Season</th>
</tr>
</thead>
<tbody>
<tr>
<td>Afuá</td>
<td>1998</td>
<td>4</td>
<td>296 (14)</td>
<td>1.35%</td>
<td>Rainfall</td>
</tr>
<tr>
<td>Altamira</td>
<td>1998</td>
<td>13</td>
<td>819 (54)</td>
<td>0.96%–2.01%</td>
<td>Rainfall</td>
</tr>
<tr>
<td>Altamira</td>
<td>1999</td>
<td>-</td>
<td>129 (8)</td>
<td>-</td>
<td>Dry</td>
</tr>
<tr>
<td>Breves</td>
<td>1999</td>
<td>11</td>
<td>546 (23)</td>
<td>2.01%</td>
<td>Rainfall</td>
</tr>
</tbody>
</table>

(≥) No. of lots of pooled mosquitoes in which virus isolation was attempted.
To date, financing YF vaccine has been a major problem for disease-endemic countries, but studies developed in Africa have suggested that low costs are possible (26). Moreover, Brazil has been producing YF vaccine with the 17D strain on a large scale for a long time. We believe that combined efforts under PAHO/WHO support to supply other countries in the region with vaccine for a massive vaccination campaign would save thousands of lives.

Acknowledgments

We thank the National Health Foundation (FUNASA) in Pará and Amapá States for logistic support in Afiú and Breves municipalities and NUPE (Pará State Epidemiology Nucleus) of SESPA (Department of Public Health of Pará State).

This work was financed by FUNASA (National Health Foundation) and Instituto Evandro Chagas, Ministry of Health of Brazil.

Dr. Vasconcelos works in virology, especially arboviruses, hantaviruses, and incoming and reemerging viruses. He is chief of the arbovirus section of the Instituto Evandro Chagas, FUNASA, Ministry of Health, which is a World Health Organization/Pan American Health Organization Collaborating Center for Arbovirus Reference and Research.

References