Nosocomial Endocarditis Caused by Corynebacterium amycolatum and Other Nondiphtheriae Corynebacteria

Karen L. Knox* and Alison H. Holmes†

The nondiphtheriae corynebacteria are uncommon but increasingly recognized as agents of endocarditis in patients with underlying structural heart disease or prosthetic valves. We describe three cases of nosocomial endocarditis caused by nondiphtheriae corynebacteria, including the first reported case of Corynebacterium amycolatum endocarditis. These all occurred in association with indwelling intravascular devices.

The nondiphtheriae corynebacteria, major components of the normal flora of human skin and mucous membranes, are commonly isolated from clinical specimens. As such, they are frequently dismissed as contaminants. Recently, however, the microbiologic classification of this group of organisms and their role in clinical disease are being more clearly defined (1). In particular, Corynebacterium amycolatum, which was first described in 1988 (2), is becoming widely recognized as an important pathogen, although it has been underreported in part because of its misidentification as C. xerosis (3,4), an established human pathogen.

These organisms are causes of community-acquired endocarditis in patients who have underlying structural heart disease or are immunocompromised, as well as of prosthetic-valve endocarditis. Cases in previously healthy patients are rarely described (5,6). Hospital-acquired bacterial endocarditis accounts for 7.5% to 29% of cases of endocarditis in tertiary-care hospitals and is strongly associated with infection of indwelling intravascular lines (7). Common causative organisms include Staphylococcus aureus, enterococci, and coagulase-negative staphylococci. Corynebacterium species are rarely reported as agents of hospital-acquired endocarditis.

We describe three cases of endocarditis caused by three different species of corynebacteria in our hospital within an 18-month period, all associated with indwelling intravascular devices (IID). These accounted for 3 of 10 cases of hospital-acquired endocarditis identified during this period and include the first reported case of endocarditis caused by C. amycolatum. Two of the Corynebacterium species were resistant to multiple antibiotics. These cases highlight the importance of the nondiphtheriae corynebacteria as emerging multiresistant nosocomial pathogens in the growing population of patients with IID.

Case Reports

Case 1

A 74-year-old woman was hospitalized with a diagnosis of antineutrophil cytoplasmic antibody (ANCA)-positive vasculitis. Her treatment included daily plasma exchanges via a right internal jugular vascular catheter that was placed on the 5th hospital day. On day 21 of admission, she had a low-grade fever that resolved after removal of the catheter, the tip of which produced a culture of diphtheroids. Blood cultures were sterile.

The patient was discharged after 1 month but was readmitted 4 weeks later for control of vasculitis. On admission she was anemic (hemoglobin 7.0 g/dL) and had persistent microscopic hematuria and elevated C-reactive protein. She had no new vasculitic manifestations and, as previously, a grade II systolic murmur was noted at the left sternal edge. She had a fever of 38°C 48 hours after admission. Four sets of blood cultures were positive for C. amycolatum (Table 1). Transesophageal echocardiogram (TOE) showed a large mobile vegetation on the mitral valve, with prolapse and mild to moderate mitral regurgitation. A transthoracic echocardiogram on her previous admission had shown no mitral valve abnormality. A combination of intravenous vancomycin and oral rifampicin was begun, and the fever resolved within 48 hours. The patient refused surgical intervention, and dual antibiotic therapy was continued indefinitely. A follow-up TOE at 15 months showed improvement in mitral valve features and no vegetations. The antibiotics were stopped after 16 months, and 5 months later the patient had no clinical evidence of disease recurrence.

Case 2

A 69-year-old woman with hemodialysis-dependent chronic renal failure secondary to ANCA-positive vasculitis was admitted after acute thrombosis of a GoreTex dialysis fistula in her left arm. She had required temporary vascular access for hemodialysis for the preceding 12 days. On examination she was unwell and feverish. She had a grade I systolic murmur over the cardiac apex; no previous echocardiogram was available. Two sets of blood cultures grew C. striatum (Table 1), as did the tip of her vascular catheter on removal. A TOE revealed a large vegetation on her mitral valve. Because of penicillin allergy, intravenous vancomycin and oral rifampicin were begun. The prosthetic arteriovenous fistula could not be removed completely, although it was considered a potential reservoir of infection. The patient had a mitral valve replacement 22 days into medical therapy, on the 29th hospital day. Her valve was completely destroyed, but on culture was sterile. Antibiotics were continued for another 15 days; however, the patient had postoperative complications and died 4 weeks later.

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Table 1. Patient characteristics, heart valve affected, source of Corynebacterium infection and organism cultured, London

<table>
<thead>
<tr>
<th>Case</th>
<th>Age(yr)/sex</th>
<th>Underlying disease process</th>
<th>Site of endocarditis</th>
<th>Associated IID*</th>
<th>Therapy</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>74/F</td>
<td>ANCA + vasculitis</td>
<td>Native mitral valve</td>
<td>Vascular catheter for HD</td>
<td>Vancomycin i.v. + oral rifampicin, 16 months</td>
<td>Resolved at 21 months postdiagnosis</td>
</tr>
<tr>
<td>2</td>
<td>69/F</td>
<td>ANCA + vasculitis</td>
<td>Native mitral valve</td>
<td>Gortex AV fistula, vascular catheter for HD</td>
<td>Vancomycin i.v. + oral rifampicin, 37 days, mitral valve replacement</td>
<td>Died of unrelated causes 9 weeks after diagnosis</td>
</tr>
<tr>
<td>3</td>
<td>53/M</td>
<td>Postoperative acute renal failure</td>
<td>Prosthetic mitral valve (Starr Edwards)</td>
<td>Vascular catheter for HD CVC</td>
<td>Vancomycin i.v. + oral rifampicin, 42 days</td>
<td>Died of unrelated causes 8.5 months postdiagnosis</td>
</tr>
</tbody>
</table>

*IID = indwelling intravascular device; ANCA = anti-neutrophil cytoplasmic antibody; HD = hemodialysis; AV = arteriovenous; CVC = central venous catheter.

Case 3

A 53-year-old man underwent mitral valve replacement with a Starr Edwards metal valve. His postoperative course was complicated by acute renal failure, for which he required temporary hemodialysis by vascular catheter for 36 days. The maximum time a single line was in place was 13 days. Cultures of four vascular catheter tips after their removal grew both diphtheroids and coagulase-negative staphylococci, which were considered to be normal skin flora. During this time, a low-grade fever developed that resolved after a change of lines. Blood cultures remained sterile. Serial echocardiograms over the following 2 weeks, however, showed a worsening paraprosthetic leak. Subsequently, five sets of blood cultures grew Corynebacterium jeikeium (Table 1). The patient completed 6 weeks of intravenous vancomycin and oral rifampicin for prosthetic valve endocarditis and was well on discharge. At follow-up, TOE studies showed no further increase in the paraprosthetic leak, and the patient had no clinical evidence of relapse of infection. He died 7 months later from noninfective causes.

Discussion

Interest in the taxonomy of the nondiphtheriae corynebacteria has increased, with a resultant reclassification of earlier defined taxa and discovery of new pathogens in the group (1,8). Of the remaining seven cases, two were due to methicillin-resistant S. aureus and five to methicillin-sensitive S. aureus.

Since Lehmann and Neumann proposed in 1896 that bacteria morphologically resembling the diphtheria bacillus be incorporated into the genus Corynebacterium, the classification of the coryneform bacteria has been drastically altered (8). This change has, in turn, increased the difficulty of identifying these organisms, as methods that reliably differentiate related species, such as mycolic acid chromatography, gas liquid chromatography, and molecular amplification techniques, cannot be used in the routine laboratory setting. During 1987 to 1995, 11 new Corynebacterium species were described (1). Commercial identification systems need to be updated to include those species relevant in human disease. C. amycolatum has only recently been included in the updated API Coryne database 2.0 (4), which, as illustrated by our case, may still lead to misidentification if used alone. That there have been no previous reports of C. amycolatum causing endocarditis may be due to its misidentification as other nonlipophilic fermentative corynebacteria species such as C. xerosis and C. minutissimum (3,16), both of which are associated with human disease.

Published material provides useful schema for differentiating C. amycolatum, C. minutissimum, and C. striatum by using colonial morphology, carbohydrate assimilation tests, and sensitivity to amoxycillin and the vibriostatic compound O/129, in conjunction with the API Coryne and API 20NE systems (17). Antibiotic sensitivity patterns may support identification, with C. amycolatum and C. jeikeium generally resistant to multiple antibiotics (18). In contrast, C. striatum, C. minutissimum, and C. xerosis generally are sensitive to a wide range of antibiotics.

Funke et al. (1) have published guidelines for identifying the coryneform bacteria, including simple phenotypic characteristics but also recommending more complex chemotaxonomic investigations and molecular genetic analysis if...
phenotypic characteristics do not differentiate between species. These guidelines are intended to facilitate the establishment of true disease associations of these organisms. In our experience, morphologic features combined with antibiogram profiles and the Coryne API allowed identification of two out of three corynebacteria (Table 2). As the colonial morphology and the Coryne API profile did not match in the isolate from Case 1, the organism was sent to our reference laboratory, where identification as *C. amycolatum* was confirmed by gas liquid chromatography (Microbial Identification System, Microbial ID, Newark, DE).

Treatment regimens described include penicillin alone, beta-lactam antibiotics or erythromycin plus gentamicin, and vancomycin. Our three patients all received dual therapy with vancomycin and rifampicin. This combination has not previously been described but was effective in two of the three patients (including the case of prosthetic valve endocarditis), who were both free from infection at least 5 months after cessation of therapy. In a review of 19 patients with prosthetic valve endocarditis due to diphtheroids (19), 4 (57.1%) of 7 patients treated with antibiotics alone were reported as cured at least 1 year posttreatment. Of 12 patients treated both medically and surgically, 7 (58.3%) were reported as having a microbiologic cure (19).

This study highlights the importance of the nondiphtheriae corynebacteria in severe human disease. Specifically, clinicians and microbiologists must be aware of the potential risk factors for nondiphtheriae corynebacterial endocarditis in the hospital setting and the danger of overlooking positive long-line tip cultures as normal skin flora or contaminants. Stringent identification of clinical isolates will be required to define the role of the nondiphtheriae corynebacteria in human disease.

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**References**

3. Funke G, Lawson PA, Bernard KA, Collins MD. Most *Corynebacterium* *serosis* strains identified in the routine clinical laboratory correspond to *Corynebacterium amycolatum*. J Clin Microbiol 1996;34:1124-8.

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**Table 2. Identification and antibiograms of the Corynebacteria species in three cases of endocarditis, London**

<table>
<thead>
<tr>
<th>Organism: gram-positive rod, nonmotile, catalase positive</th>
<th>Colonial morphology</th>
<th>Further identification</th>
<th>Antibiotic sensitivity pattern (Stokes plate)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Corynebacterium amycolatum</em> Case 1</td>
<td>Dry, gray&lt;sup&gt;a&lt;/sup&gt;</td>
<td>Coryne API GLC&lt;sup&gt;b&lt;/sup&gt;</td>
<td>Rif, Teic, Vanc</td>
</tr>
<tr>
<td><em>C. striatum</em> Case 2</td>
<td>Moist, white, smooth&lt;sup&gt;a&lt;/sup&gt;</td>
<td>Coryne API</td>
<td>Cip, Ery, F, Gent, Pen, Trim</td>
</tr>
<tr>
<td><em>C. xerosis</em> Case 3</td>
<td>Gray, nonhemolytic&lt;sup&gt;a&lt;/sup&gt; (aerobic growth)</td>
<td>Coryne API</td>
<td>Rif, Teic, Vanc</td>
</tr>
</tbody>
</table>

<sup>a</sup>Horse blood agar at 37°C.

<sup>b</sup>API = analytical profile index; GLC = gas liquid chromatography; Cip = ciprofloxacin; Ery = erythromycin; F = fucidin; Gent = gentamicin; Pen = penicillin; Rif = rifampicin; Teic = teicoplanin; Trim = Trimethoprim; Vanc = vancomycin.