Rickettsia felis in Ctenocephalides spp. Fleas, Brazil

Riva P. Oliveira,* Márcio A.M. Galvão,* Claudio L. Mafra,* Chequer B. Chamone,† Simone B. Calic,† Sérgio U. Silva,‡ and David H. Walker§

In June 2000, suspected cases of Brazilian spotted fever (BSF) occurred in Coronel Fabriciano Municipality, Minas Gerais State, Brazil. Pooled fleas collected near two fatal cases contained rickettsial DNA. The nucleotide sequence alignment of the 391-bp segment of the 17-kDa protein gene showed that the products were identical to each other and to the R. felis 17-kDa gene, confirming circulation of R. felis in Brazil.

The pathogenic rickettsiae are a group of intracellular bacteria responsible for various human diseases. Rickettsia rickettsii and R. typhi and the diseases they cause—Brazilian spotted fever (BSF), transmitted by the Amblyomma cajennense tick, and murine typhus, transmitted by the Oriental rat flea—have been recognized in Brazil since the 1920s (1-3). Molecular methods, including detection by DNA amplification by polymerase chain reaction (PCR) and DNA sequence analysis, are useful in characterizing rickettsial agents in arthropods. This approach has allowed the identification of new species, such as R. felis in opossums, fleas (4,5), and blood and skin from ill humans from the United States, Mexico, France, and Brazil (6-9). We report the identification of R. felis in Ctenocephalides fleas collected during the investigation of an outbreak of spotted fever group rickettsiosis in Brazil.

Material and Methods

In June 2000, fleas and ticks were collected in a periurban area of the city of Coronel Fabriciano, Steel Valley, Minas Gerais State, Brazil (Figure 1). This survey was performed during an outbreak of suspected BSF in which two children died. They were brothers who lived in the same house. The first child who became ill was 12 years old; during the course of his disease he had fever, nausea, vomiting, diarrhea, abdominal pain, headache, myalgia, and edema. Later, renal failure and stupor occurred. The second patient had fever, rash, nausea, vomiting, diarrhea, abdominal pain, headache, myalgia, jaundice, and renal failure. Both patients reported a tick bite a day before the onset of disease. One death was later confirmed as a case of spotted fever group rickettsiosis by immunohistochemical technique in tissues collected at autopsy. PCR was performed on brain, stomach, liver, spleen, and kidney tissues collected at autopsy, preserved in formalin, and sent to the University of Texas Medical Branch at Galveston. Because the DNA was not preserved, the death could not be attributed specifically to R. rickettsii, R. felis, or other species of Rickettsia.

The ticks were collected from three dogs and five horses near the house where the deaths occurred and were stored in 70% ethanol at room temperature. Ticks were separated into 15 pools with three specimens per pool, undifferentiated by life stage or sex. Fleas were also removed from 10 dogs in the home of the child whose death was confirmed as being due to BSF (Galvão et al., unpub. data) and from 3 cats near this residence; fleas were stored at −70°C. The fleas were separated into six pools with five specimens per pool. The ticks and fleas were identified as A. cajennense and Ctenocephalides spp., respectively.

PCR amplifications were done as previously described (9) with the DNA extracted from pools of ticks and fleas (Figure 2). Each PCR product was cycle sequenced with the primers described above and fluorescein-labeled dideoxynucleotide bases in the Applied Biosystems model (11) DNA sequencing system (ABI, Foster City, CA). Sequences were edited and assembled using Chromas software (http://www.technelysium.com.au/chromas.html).

To arrive at the most accurate sequence for each PCR product, both forward and reverse sequences were determined. Where differences in nucleotide bases were observed, a predominant base was assigned if most of the sequences contained it. If one base did not predominate, the original chromatographs were consulted to resolve ambiguities.

*Universidade Federal de Ouro Preto, Minas Gerais State, Brazil; †Fundação Ezequiel Dias, Minas Gerais State, Brazil; ‡Secretaria Municipal de Saúde de Coronel Fabriciano, Minas Gerais State, Brazil; and §University of Texas Medical Branch, Galveston, Texas, USA

Figure 1. Map of Brazil and Minas Gerais State, showing Coronel Fabriciano municipality.
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confirmed that the three PCR products were identical to each sequence analysis of the aligned 391-bp segment of 17 kDa cycles for 1 minute at 94° from the rickettsial 17-kDa protein gene. PCR was performed at 30 °C. The PCR products were then separated by electrophoresis in 1% agarose gel and stained with ethidium bromide. Lanes 1-3: DNA from cat felis Rickettsia were crushed with a micropestle. The suspensions were lysed in 0.5% nolchloroform, followed by a single chloroform extraction. The lysed suspensions were extracted twice with an equal volume of phenolchloroform, and the DNA obtained from the GenBank database. These sequences were added another ecologic zone and geographic region of Latin Brazil. Our results show that, in addition to R. rickettsii and R. typhi, R. felis is also found in Brazil, as indicated by positive serology in human cases (8). Our data are the first indication by PCR of the presence of R. felis in fleas from Brazil. The Ctenocephalides spp. flea is proposed as a possible vector of this new rickettsial disease in Brazil.

Because of the complicated differential diagnosis of febrile exanthems, which includes dengue fever and other viral, rickettsial, and bacterial diseases, more attention should be paid to diagnostic laboratory investigation of rickettsial diseases. R. felis has been identified as the etiologic agent of a new rickettsiosis wherever it has been investigated: United States (Texas), Mexico, Brazil, and France (8). In Latin America, two reports have been published of human rickettssiases caused by R. felis in Mexico and Brazil (7,8). In both these reports, neurologic involvement was described, suggesting a severe clinical course associated with R. felis (15).

Although our investigation does not provide evidence for widespread flea infection by R. felis in Brazil, we demonstrate for the first time the presence of infection by this bacteria in Brazilian Ctenocephalides fleas. The descriptions of R. felis-positive human cases (8) in the same area where R. felis was identified in Ctenocephalides fleas indicate the possibility of this flea’s being the vector of human R. felis rickettsiosis in Brazil.

References


Address for correspondence: Márcio Galvão, Departamento de Nutrição Clínica e Social, Escola de Nutrição, Universidade Federal de Ouro Preto, Campus Morro do Cruzeiro, Ouro Preto–Minas Gerais, Brazil – CEP 34.500-000; fax: 55-31--35511689; e-mail: mgalvao@enut.ufop.br