Evaluation in Nonhuman Primates of Vaccines against Ebola Virus

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Ebola virus (EBOV) causes acute hemorrhagic fever that is fatal in up to 90% of cases in both humans and nonhuman primates. No vaccines or treatments are available for human use. We evaluated the effects in nonhuman primates of vaccine strategies that had protected mice or guinea pigs from lethal EBOV infection. The following immunogens were used: RNA replicon particles derived from an attenuated strain of *Venezuelan equine encephalitis virus* (VEEV) expressing EBOV glycoprotein and nucleoprotein; recombinant *Vaccinia virus* expressing EBOV glycoprotein; liposomes containing lipid A and inactivated EBOV; and a concentrated, inactivated whole-virion preparation. None of these strategies successfully protected nonhuman primates from robust challenge with EBOV. The disease observed in primates differed from that in rodents, suggesting that rodent models of EBOV may not predict the efficacy of candidate vaccines in primates and that protection of primates may require different mechanisms.

In early attempts to develop a vaccine against EBOV, guinea pigs or nonhuman primates were vaccinated with formalin-fixed or heat-inactivated virion preparations. Results from these studies were inconsistent: Lupton et al. (5) partially protected guinea pigs against EBOV, while Mikhailov et al. (6) achieved complete protection of four of five hamadryad baboons by vaccinating them with an inactivated EBOV vaccine. However, other studies suggested that inactivated EBOV did not induce sufficient immunity to reliably protect hamadryl baboons against a lethal challenge (7). Conventional strategies of attenuating viruses for use as human vaccines have not been pursued for EBOV because of concerns about reversion to a wild-type form. However, the possibility of following this strategy by using newly developed infectious clones of EBOV may now be feasible (8).

Recent efforts have focused on the use of recombinant DNA techniques to stimulate cytotoxic T-lymphocyte responses. Vaccinating guinea pigs with plasmids against EBOV nucleoprotein (NP), soluble glycoprotein, or glycoprotein (GP) elicited humoral and cellular immune responses against these gene products but only partially protected them against lethal challenge (9). However, results of this study were difficult to interpret because all the guinea pigs were killed 10 days after EBOV challenge, which is within the expected survival time for untreated animals (8-14 days) (10). In 2000, Sullivan et al. (11) reported protection of cynomolgus monkeys from EBOV infection by injecting them with naked-DNA GP, followed by an adenovirus-expressing GP booster. Results of this study document the feasibility of vaccination against EBOV. However, these results require confirmation and further evaluation, as a low dose (6 PFU) was used for the challenge. Other studies reported a protective effect of EBOV vaccination with a low infective challenge dose (ten 50% lethal doses [LD50]) (7); however, all vaccinated animals in these dosing studies died after receiving higher infective doses (100 and 1,000 LD50), which may more accurately mimic natural or nosocomial exposures.

Our efforts to develop a vaccine against EBOV focused on several potential vaccine candidates. First, we used *Venezuelan equine encephalitis virus* (VEEV) replicon particles (VRP) expressing EBOV genes known to protect guinea pigs and mice from EBOV disease (10); VRP expressing MBGV genes also protected guinea pigs and cynomolgus monkeys against MBGV (12). Second, we used a recombinant *Vaccinia virus* (VACV) system expressing EBOV GP and demonstrated that this vector protected guinea pigs from EBOV hemorrhagic fever (13). A third strategy used encapsulated, gamma-irradiated EBOV particles in liposomes containing lipid A (14); and the fourth approach evaluated vaccination with a concentrated, gamma-irradiated whole-virion preparation. None of these approaches, which successfully protected rodents from lethal infection, were protective for cynomolgus or rhesus macaques challenged with EBOV.

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Materials and Methods

Cynomolgus macaques (Macaca fascicularis) or rhesus macaques (M. mulatta) weighing 4 to 6 kg were used. For vaccine studies with VEE replicons, EBOV GP or NP genes were introduced into the VEEV RNA as described (10). Groups of three cynomolgus macaques were vaccinated with VRP that expressed EBOV GP, EBOV NP, a mixture of EBOV GP and EBOV NP, or a control antigen (influenza hemagglutinin) that has no effect on EBOV immunity. Animals were vaccinated by subcutaneous injection of $10^7$ focus-forming units of VRP in a total of 0.5 mL at one site. Vaccinations were repeated 28 days after the first injection and 28 days after the second.

In conducting research with animals, the investigators followed the Guide for the Care and Use of Laboratory Animals prepared by the Committee on Care and Use of Laboratory Animals of the Institute of Laboratory Animal Resources, National Research Council (1996). The animal facilities and animal care and use program of the U.S. Army Medical Research Institute of Infectious Diseases are accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International.

For vaccine studies using primates, we adapted the optimal immunization regimens determined from the rodent studies. For the vaccine based on recombinant VACV, the EBOV GP gene was inserted into a VACV transfer vector plasmid, and recombinant VACV expressing EBOV GP were isolated as reported (13). Three cynomolgus macaques were injected subcutaneously with the EBOV GP-expressing VACV vector. Injections were repeated at 28 and 53 days after the first injection.

For vaccine studies with inactivated EBOV whole-virion preparation, viral particles were concentrated from Vero cell culture fluids by ultracentrifugation in a sucrose density gradient. Infectivity titer of the preparation was approximately 8.0 log_{10} PFU/mL. The preparation was inactivated by exposure to $^{60}$Co gamma rays ($6 \times 10^6$ rads). The absence of residual infectivity was proven by exhaustive testing for residual infectivity in assays in Vero cells (15,16). Two cynomolgus monkeys and two rhesus monkeys were injected subcutaneously with a 50-µg dose of the gamma-irradiated virion preparation in RIBI adjuvant (Corixa, Hamilton, MT). As a further check on complete viral inactivation, blood samples taken from the monkeys 3 and 5 days after they received the vaccine were free of infectious viremia. Injections were repeated at days 7 and 35 after the initial injection.

For vaccine studies using a liposome formulation, three cynomolgus monkeys were vaccinated with gamma-irradiated virus encapsulated in liposomes containing lipid A, as described for previous studies in mice (14). Animals received 1.0 mL of the liposome preparation by intravenous injections that were repeated at 28 and 55 days after the initial vaccination. Four macaques (two cynomolgus and two rhesus) served as unvaccinated controls for the VACV, gamma-inactivated virion, and liposome studies.

Anti-EBOV neutralizing antibody titers were monitored by measuring plaque reduction in a constant virus:serum dilution format (15). All macaques received intramuscular injections in the leg with 1,000 PFU of the Zaire subtype of EBOV, which was isolated from a human patient in 1995 (16). Blood was obtained from all monkeys under Telazol anesthesia (Fort Dodge Laboratories, Fort Dodge, IA) at 2- or 3-day intervals postinfection to determine infectious viremia, neutralizing antibody titers, and standard hematologic and clinical pathology parameters. All terminally ill monkeys were killed and necropsied for pathologic examination. Virus infectivity assays on plasma and tissue homogenates were done by forming plaques on Vero cell monolayers as described (15,16).

Tissues were immersion fixed in 10% neutral-buffered formalin and processed for histopathologic and immunohistochemical characteristics as described (17-19). Replicate sections of spleen were stained with phosphotungstic acid hematoxylin to demonstrate polymerized fibrin. Sections of spleen from five EBOV-infected guinea pigs and five mice from previous studies (20,21) were similarly stained for polymerized fibrin. Portions of selected tissues from 11 monkeys were also immersion fixed in 4% formaldehyde and 1% glutaraldehyde and processed for transmission electron microscopy according to conventional procedures (17-19).

Results

Serologic Response

Prechallenge EBOV neutralization titers were measured for the 26 nonhuman primates used in this study (Table 1). Although all vaccinated animals seroconverted by immunoglobulin G enzyme-linked immunosorbent assay, neutralizing antibody (PRNT_{50}) titers were very low. Only one macaque vaccinated with VRP-expressed EBOV GP had detectable antibody titers.

Table 1. Prechallenge neutralization titers of Ebola virus (EBOV)-vaccinated monkeys

<table>
<thead>
<tr>
<th>Nonhuman primate species</th>
<th>No. of animals</th>
<th>Vector Antigen</th>
<th>Neutralization titers&lt;sup&gt;a&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cynomolgus</td>
<td>3</td>
<td>Replicon GP</td>
<td>0, 0, 0</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>3</td>
<td>Replicon NP</td>
<td>0, 0, 0</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>3</td>
<td>Replicon GP + NP</td>
<td>0, 0, 10</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>3</td>
<td>Replicon Influenza HA</td>
<td>0, 0, 0</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>3</td>
<td>Vaccinia GP</td>
<td>10, 20, 20</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>3</td>
<td>Liposome Inactivated virion</td>
<td>20, 40, 80</td>
</tr>
<tr>
<td>Rhesus</td>
<td>2</td>
<td>Inactivated virion</td>
<td>10, 20</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>2</td>
<td>None GP</td>
<td>10, 20, 20</td>
</tr>
<tr>
<td>Rhesus</td>
<td>2</td>
<td>None NP</td>
<td>10, 20, 20</td>
</tr>
</tbody>
</table>

<sup>a</sup>Immunoglobulin G enzyme-linked immunosorbent assay, neutralizing antibody (PRNT_{50}).

<sup>b</sup>Animal survived challenge.
neutralizing antibody. The marginal PRNT did not preclude challenge of the monkeys; however, in previous studies, similar results were obtained when cynomolgus macaques were vaccinated with the VRP expressing MBGV genes, yet the animals were protected from lethal disease (12).

**Challenge of Vaccinated Monkeys with EBOV**

All animals, including the four untreated macaques, were challenged with 1,000 PFU of EBOV. Timing of challenge varied because of differences in the optimal immunization regimens determined by preliminary testing in rodents. VRP-vaccinated animals were challenged 49 days after the third vaccine dose. At postchallenge day 3, all animals became ill; two animals from each vaccination group (i.e., GP, NP, GP + NP, influenza HA) died on day 6, and the remaining animals died on day 7 (Table 2). VAC GP-inoculated macaques were challenged 45 days after the third vaccine dose, EBOV liposome-vaccinated animals 35 days after the third vaccine dose, and macaques vaccinated with the gamma-irradiated whole-virion preparation 43 days after the third vaccine dose. Again, all animals except one rhesus macaque, which received the gamma-irradiated virion preparation, became ill on the third day after challenge. Two cynomolgus macaques vaccinated with the gamma-irradiated virion preparation, one VAC-GP animal, and one untreated cynomolgus macaque died on postchallenge day 6 (Table 2). The two remaining VAC-GP animals died at day 7 after challenge, as did two of the animals vaccinated with the EBOV liposome preparation and the remaining untreated cynomolgus macaque. The untreated rhesus macaques died on days 8 and 9 postchallenge; one rhesus vaccinated with the gamma-irradiated virion preparation died on day 9, and the other survived challenge. The remaining animal vaccinated with the EBOV liposome preparation died 11 days after challenge. The rhesus macaque that survived challenge did not become ill during the study and had a PRNT\textsubscript{50} values >320 at day 26 postchallenge and 80 at days 26, 61, 99, and 902 postchallenge.

**Histopathologic Examination**

Conventional histopathologic and electron microscopic examination of lymphatic tissues, liver, and gastrointestinal tract showed no differences in lesions between the vaccinated animals and the unvaccinated EBOV-infected controls. Depletion and necrosis or apoptosis were noted in all lymphoid germinal centers in spleen, peripheral, and mesenteric lymph nodes, as described in other studies (17-19). The spleen had copious deposits of fibrin throughout the red pulp, as well as abundant karyorrhectic cellular debris. By electron microscopy, widespread bystander lymphocyte apoptosis was a prominent feature in all the lymphatic tissues examined. Fibrin and fibrinocellular thrombi were also prominent in the submucosa of the gastrointestinal tract and in hepatic sinusoids, again consistent with well-documented findings (17,18).

We also evaluated retrospectively EBOV-infected rodent tissues in parallel. Although sites of infection and morphologic changes between guinea pigs, mice, and nonhuman primates had many similarities, the lack of fibrin thrombi in spleen and visceral vasculature was particularly striking in the EBOV-infected mice (Figure). Fibrin deposition was seen in guinea pigs as reported (20), but fibrin deposits and thrombi were considerably less prevalent compared with deposits in nonhuman primates (Figure). Lymphocyte apoptosis was also less frequently observed by electron microscopy in rodent lymphatic tissues than in nonhuman primates. EBOV was demonstrated in liver, spleen, kidney, lung, adrenal gland, and lymph nodes of all necropsied monkeys by immunohistochemistry, electron microscopy, or virus infectivity titration.

**Discussion**

Our results indicate that rodent models of EBOV hemorrhagic fever do not consistently predict efficacy of candidate vaccines in nonhuman primates, perhaps because the disease course in rodents differs from that reported in human and nonhuman primates (17-19,22,23). Mice do not have the hallmark disseminated intravascular coagulation (DIC) found in endstage lesions of humans and nonhuman primates. Viremia and widespread tissue dissemination are much more apparent in nonhuman primates than in guinea pigs (20). In addition, guinea pigs have less DIC than do nonhuman primates. Lymphocyte apoptosis was not reported to be a prominent feature of EBOV infection in mice or guinea pigs (20,21) but was a consistent feature of disease in humans (24) and nonhuman primates (19). Clinical disease and related pathologic features in nonhuman primates infected with EBOV appear to more closely resemble those described in human EBOV hemorrhagic fever (22,23). Other studies have shown inconsistencies between rodent and nonhuman primate models of human hemorrhagic disease in the protective efficacy of candidate vaccines. For example, guinea pigs were protected from Lassa virus by VAC recombinants expressing the viral nucleopro-

<table>
<thead>
<tr>
<th>NHP Species</th>
<th>Vector</th>
<th>Antigen</th>
<th>Survival/total</th>
<th>Viremic/total</th>
<th>Day of death\textsuperscript{a}</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cynomolgus</td>
<td>Replicon</td>
<td>GP</td>
<td>0/3</td>
<td>3/3</td>
<td>6, 6, 7</td>
</tr>
<tr>
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<td>Replicon</td>
<td>NP</td>
<td>0/3</td>
<td>3/3</td>
<td>6, 6, 7</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>Replicon</td>
<td>GP + NP</td>
<td>0/3</td>
<td>3/3</td>
<td>6, 6, 7</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>Replicon</td>
<td>Influenza HA</td>
<td>0/3</td>
<td>3/3</td>
<td>6, 6, 7</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>Vaccinia</td>
<td>GP</td>
<td>0/3</td>
<td>3/3</td>
<td>6, 6, 7</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>Liposome</td>
<td>Inactivated virion</td>
<td>0/3</td>
<td>3/3</td>
<td>7, 7, 11</td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>Inactivated virion</td>
<td>0/2</td>
<td>2/2</td>
<td>6, 6</td>
<td></td>
</tr>
<tr>
<td>Rhesus</td>
<td>Inactivated virion</td>
<td>1/2</td>
<td>2/2</td>
<td>9</td>
<td></td>
</tr>
<tr>
<td>Cynomolgus</td>
<td>None</td>
<td></td>
<td>0/2</td>
<td>2/2</td>
<td>6, 7</td>
</tr>
<tr>
<td>Rhesus</td>
<td>None</td>
<td></td>
<td>0/2</td>
<td>2/2</td>
<td>8, 9</td>
</tr>
</tbody>
</table>

\textsuperscript{a}Number of days after challenge with 1,000 PFU of EBOV. NHP, nonhuman primate; GP, glycoprotein; NP, nucleoprotein.
monkey. Electron micrograph showing abundant fibrin deposits in pulpe
ized fibrin deposits in pulp (PTA hematoxylin, original magnification X400).

Lower left: cynomolgus monkey. Note deposition of polymerized fibrin in
red pulp (PTA hematoxylin, original magnification X5300). This field shows
infrquent fibrin deposits; most fields in five of five animals examined showed no evidence of polymerized fibrin. Lower right: cynomolgus
monkey. Electron micrograph showing abundant fibrin deposits in pulp
(original magnification X5300). Field representative of 11 of 11 monkeys examined.

Figure. Sections of spleen from Ebola virus (EBOV)-infected animals.

Top left, BALB/c mouse, note absence of polymerized fibrin (phospho-
tungstic acid [PTA] hematoxylin, original magnification X400). Field rep-
resentative of five of five mice tested. Top right: guinea pig. Note
discrete foci of polymerized fibrin (arrows) (PTA hematoxylin, original
magnification X400). This field shows infrequent fibrin deposits; most
fields in five of five animals examined showed no evidence of polymer-
ized fibrin. Lower left: cynomolgus monkey. Note deposition of polymer-
ized fibrin in red pulp (PTA hematoxylin, original magnification X4000). Field rep-
resentative of 25 of 25 monkeys. Lower right: cynomolgus
monkey. Electron micrograph showing abundant fibrin deposits in pulp
(original magnification X5300). Field representative of 11 of 11 mon-
keys examined.

tein (25, 26); however, this vaccination strategy failed to pro-
tect rhesus macaques (27).

The effort to develop an EBOV vaccine began after the ini-
tial identification of EBOV in 1976, but 25 years later the goal
remains elusive. Attempts to develop killed-virus vaccines
against EBOV hemorrhagic fever have had inconsistent results
(5-7). Recent progress in genetic vaccination strategies has
demonstrated that immunity can be achieved against a low
dose of EBOV. While protection against any lethal challenge
dose of EBOV is a remarkable achievement, we have set the
bar somewhat higher than 6 PFU, since a laboratory exposure
through a needlestick and infected blood would likely entail a
dose of at least 1,000 PFU. Therefore, our priority is to empir-
ically develop a vaccine that protects against at least 1,000
PFU rather than to initiate an exhaustive investigation of pro-
tective immune mechanisms. We were encouraged by the
demonstrated success of the VEEV replicon vector expressing
MBGV glycoprotein in protecting cynomolgus macaques from
challenge with homologous MBGV (12). No MBGV-neutral-
izing activity was observed at ≥1:20 dilutions in prechallenge
sera of any of the MBGV GP VRP-vaccinated macaques (12),
yet these animals did not become viremic, showed no signs of
disease, and survived challenge. Historically, Filovirus-
neutralizing antibodies have been difficult to demonstrate in vitro
(15); while the presence of neutralizing antibodies is desirable,
it is neither sufficient nor necessary to clear viral infection
(16). Unfortunately, the VEEV replicon strategy that was suc-
cessfully employed for MBGV in cynomolgus macaques and
for EBOV in mice and guinea pigs (10) did not protect cyno-
molgus macaques from EBOV disease. These differences
observed between EBOV and MBGV may result from differ-
ences in the course of infection. Specifically, the mean day of
death for untreated cynomolgus monkeys experimentally
infected intramuscularly with 1,000 PFU of EBOV (Zaire sub-
type) is 6.3 (n=15; data not shown), while the mean day of
death for cynomolgus monkeys infected intramuscularly with a
comparable dose of MBGV (Musoke isolate) is 9.1 (n=8;
data not shown). Thus, macaques infected with MGBV have
nearly three more days to mount an effective immune response
against the challenge virus than macaques infected with
EBOV (Zaire). Clearly, other variables, including differences
observed between EBOV (Zaire) and MBGV with respect to
GP gene expression (28), tropism, and host cell responses,
may contribute to differences in disease pathogenesis and out-
come of infections.

The induction of humoral and cytotoxic T-lymphocyte
responses to EBOV NP and GP has been demonstrated in
guinea pigs, although the relative contributions of these
responses to immune protection are unclear (9). Moreover,
transfer of EBOV immune serum in rodent and nonhuman pri-
mate models provided inconsistent results. Passive transfer
of immune serum from VRP-vaccinated animals did not protect
guinea pigs or mice against lethal challenge (10); however,
transfer of hyperimmune equine immune globulin (which had
high EBOV neutralization titers) to guinea pigs protected them
against disease (16, 29). Passive treatment of cynomolgus
monkeys with the equine immune globulin delayed death but
did not ultimately protect the monkeys against lethal EBOV
hemorrhagic fever (16, 29). In contrast, hamadryl baboons
were protected against lethal EBOV challenge by passive
treatment with the equine immune globulin and the use of a
lower challenge dose (30). These results suggest that cell-
mediated effector mechanisms may play a more important role
in protection than do humoral responses. Nonetheless, the role
of humoral immunity is in fact supported by studies showing
consistent delay in death or protection of primates therapeuti-
cally treated with EBOV-neutralizing antibodies (16, 29, 30).

We conclude that, although rodent models are useful as
preliminary screens for candidate vaccines and therapeutic
treatments, nonhuman primates likely provide a more useful
and definitive model for EBOV hemorrhagic fever in humans.
Furthermore, differences in disease pathology between rodent
and nonhuman primate models of EBOV suggest that protec-
tion of primates may require different protective mechanisms.

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References


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Perspectives. Articles should be approximately 3,500 words and should include references, not to exceed 40. Use of subheadings in the main body of the text is recommended. Photographs and illustrations are encouraged. Provide a short abstract (150 words) and a brief biographical sketch of first author—both authors if only two.

Articles in this section should provide insightful analysis and commentary about new and reemerging infectious diseases and related issues. Perspectives may also address factors known to influence the emergence of diseases, including microbial adaptation and change, human demographics and behavior, technology and industry, economic development and land use, international travel and commerce, and the breakdown of public health measures. If detailed methods are included, a separate section on experimental procedures should immediately follow the body of the text.