

Cefepime MIC as a Predictor of the Extended-Spectrum β -Lactamase Type in *Klebsiella pneumoniae*, Taiwan

Wen Liang Yu,*† Michael A. Pfaller,†
Patricia L. Winokur,† and Ronald N. Jones†‡§

To guide selection of carbapenems or fourth-generation cephalosporins as therapy, 110 *Klebsiella pneumoniae* isolates with extended-spectrum β -lactamases from Taiwan were characterized by phenotypic (MICs), molecular, and chemical methods. MIC patterns of ceftazidime and cefepime clearly differentiate strains treatable by cefepime and those capable of efficiently hydrolyzing available cephalosporins (CTX-M series and SHV-types). Continued use of cefepime appears to be a treatment option in cases for which MIC results are available and interpreted by the criteria presented.

In recent years, extended-spectrum β -lactamase-producing *Klebsiella pneumoniae* (ESBL-KP) strains of the TEM, SHV, and CTX-M types have been discovered worldwide. Reference broth microdilution susceptibility rates (MIC \leq 8 mg/L) for cefepime among ESBL-KP in various geographic regions show a wide range: Canada 94.4%, United States 87.6%, Western Pacific 76.1%, Europe 63.6%, and Latin America 49.6% (1). In Taiwan, the in vitro cefepime susceptibilities of ESBL-KP ranged from 37% to 100% (2,3). The gene encoding SHV-5 (pI 8.2) has been reported to be the most common ESBL in klebsiellae in Taiwan (2,4). The CTX-M-3 (pI 8.4) enzyme has also been discovered in *Escherichia coli* isolates in southern Taiwan (5). For our study, we focused on the mechanisms of cefepime resistance among ESBL-KP isolates in Taiwan and attempted to predict cephalosporin therapeutic potentials by simple phenotypic patterns.

The Study

We initially conducted reference broth microdilution tests (6,7) for 211 isolates of endemic and epidemic ESBL-KP from Taiwan; 53% of isolates had a cefepime MIC of \leq 8 mg/L (susceptible). Isoelectric focusing (IEF) was then performed by the method of Matthew et al. (8). Approximately 40% of isolates had an enzyme with a pI of 8.2 (SHV-5); 40% of isolates produced enzymes with a pI of 7.9, 8.4, or 8.8 (CTX-M-type); an

additional 20% of isolates contained both an SHV-5 plus a CTX-M enzyme.

The IEF results of 110 geographically representative isolates of ESBL-KP were categorized by cefepime MIC level (Table). The enzymes with pIs of 7.6 and 5.4 were SHV-1 and TEM-1, respectively, which have been reported previously in Taiwan hospitals (2,4). All the enzymes with pIs of 5.4, 7.6, and 8.2 were evenly distributed among the isolates regardless of cefepime MIC values, indicating no association with resistance to this fourth-generation cephalosporin. All 23 isolates with pI 8.2 enzymes and a nonsusceptible cefepime MIC (\geq 16 mg/L) contained enzymes with pIs of 7.9, 8.4, or 8.8. In the absence of these CTX-M enzymes, isolates with pI 8.2 enzymes remained susceptible to cefepime. Thus, the high MIC level for cefepime was attributed to enzymes with pIs of 7.9, 8.4, and 8.8. This finding is supported by the fact that those isolates with a single CTX-M enzyme (10 with pI 7.9 enzymes [CTX-M-14] and 8 with pI 8.4 enzymes [CTX-M-3]) had very elevated cefepime MIC results in the absence of a pI 8.2 enzyme (9). Two isolates with pI 8.4 enzymes remained susceptible to cefepime (MIC 2 μ g/mL) and probably produced low levels of CTX-M-3.

These data indicate that cefepime resistance in ESBL-KP isolates from Taiwan may result from either the cumulative effect of pI 7.9, 8.4, 8.8, or 8.2 enzymes or hyperproduction of any of the enzymes with the CTX-M phenotype (pI 7.9, 8.4, or 8.8). The enzyme with a pI of 8.8 is a novel CTX-M β -lactamase most similar to CTX-M-3 (9).

Several CTX-M enzymes have been shown to confer high MIC levels for cefepime (10-12). Bauernfeind et al. reported an isolate of *Salmonella Typhimurium* that had a CTX-M-2 enzyme (pI 7.9) and a cefepime MIC of 64 mg/L (10). Outbreaks have also been reported of isolates producing CTX-M enzymes (pI 8.4), including *K. pneumoniae* (cefepime MIC 4-8 mg/L), *E. coli* (cefepime MIC 8-32 mg/L), and *Serratia marcescens* (cefepime MIC 16-64 mg/L) (11).

Szabo et al. reported an outbreak of 14 ESBL-KP strains (pI 8.2, probably SHV-5) that had high-level resistance to cefepime (MIC₉₀ >256 mg/L) (12). Tzouveleki et al. also noticed seven isolates of ESBL-KP (SHV-5) with cefepime MICs ranging from 32 mg/L to 64 mg/L. These researchers described the elevated cefepime MIC as being due to the combined effect of SHV-5 hyperproduction and decreased outer membrane permeability (loss of 36-kDa outer membrane protein [OMP]) (13). The cefepime MIC for isolates hyperproducing SHV-5 without loss of the 36-kDa OMP remained <16 mg/L, a susceptible level (13). Loss of the 36-kDa OMP also conferred cefoxitin resistance, and introduction of a plasmid carrying the 36-kDa OMP gene markedly reduced the MIC of cefoxitin, from 128 mg/L to 16 mg/L (13). Whether the isolates reported by Szabo et al. also had concomitant outer membrane defects is unknown, but these authors later recommended that cefepime not be considered the treatment of choice against SHV-5-producing ESBL-KP (14). Whether our cefepime-resistant isolates had a concomitant OMP defect is

*China Medical College Hospital, Taichung, Taiwan; †University of Iowa College of Medicine, Iowa City, Iowa, USA; ‡JONES Group/JMI Laboratories, North Liberty, Iowa, USA; and §Tufts University School of Medicine, Boston, Massachusetts, USA

Table. Distribution of pI^a values in 110 isolates of extended-spectrum β -lactamase-producing *Klebsiella pneumoniae*, stratified by cefepime MIC level,^b Taiwan

Cefepime MIC (mg/L) (n=110)	pI 5.4 (n=89)	pI 6.3 (n=7)	pI 7.6 (n=92)	pI 7.8 (n=4)	pI 7.9 (n=40)	pI 8.2 (n=62)	pI 8.4 (n=43)	pI 8.8 (n=3)
≥32 (n=38)	38	0	31	0	29 ^c	20 ^d	18 ^e	2
16 (n=6)	6	0	5	0	5 ^c	3 ^d	5 ^e	0
8 (n=26)	20	2 ^f	22	0	6 ^g	8 ^h	12 ^g	0
4 (n=19)	9	4 ^f	17	1 ^f	0	12 ^h	6 ^g	1 ^g
2 (n=10)	7	1 ^f	8	3 ^f	0	8 ^h	2 ^g	0
≤1 (n=11)	9	0	9	0	0	11 ^h	0	0

^apI, Isoelectric point. Each strain may have multiple pIs.

^bMIC test, reference broth microdilution method according to National Committee for Clinical Laboratory Standards.

^cTen of 34 isolates having a cefepime MIC ≥16 mg/L did not have enzymes with pI values of 8.2 or 8.4.

^dAll 23 isolates were simultaneously coexistent with pI 7.9, 8.4, or 8.8 (12 with pI 7.9 plus 8.4; 9 with pI 7.9; and 2 with pI 8.8).

^eEight of 23 isolates (cefepime MIC ≥16 mg/L) were not coexistent with pI 8.2 or 7.9.

^fAll the 11 isolates (pI 7.8 or 6.3) were coexistent with pI 8.2.

^gCeftazidime MIC ≤8 mg/L; ceftriaxone MIC ≥32 mg/L.

^hCeftazidime MIC ≥16 mg/L.

similarly unknown. However, in 44 isolates with cefepime MICs ≥16 mg/L, only 7 were resistant to ceftazidime. Furthermore, for isolates with high cefepime MIC values resulting from single CTX-M enzymes (10 with pI 7.9 and 8 with pIs of 8.4), only two (one each with pIs of 7.9 and 8.4) were resistant to ceftazidime. The relatively low rates of ceftazidime coresistance provide indirect evidence that 36-kDa OMP loss may not play an important role in the expression of cefepime resistance in ESBL-KP strains in Taiwan.

Conclusions

Alternative therapy using cefepime against ESBL-KP strains in Taiwan could be reliable if appropriately guided by cefepime and ceftazidime MIC results. The cefepime MIC is useful for predicting the presence of CTX-M enzymes, which usually confer resistance to this fourth-generation cephalosporin. Cefepime cannot be used if the MIC exceeds 8 mg/L, which predicts the presence of CTX-M β -lactamases. Cefepime may reasonably be used clinically if the MIC is consistently ≤1 mg/L, which indicates the absence of a CTX-M enzyme. For isolates with cefepime MICs ≥2 to ≤8 mg/L, use of cefepime should be further guided by the ceftazidime MIC. If the ceftazidime MIC remains in the susceptible range (≤8 mg/L, predicting enzymes of pI 7.9, 8.4, or 8.8), cefepime should not be used. If the ceftazidime MIC is >8 mg/L (predicting enzymes of pI 8.2), cefepime at appropriate doses has a potential therapeutic role because most pI 8.2 enzymes rarely elevate the cefepime MIC to >8 mg/L.

In conclusion, outer membrane defects and the inoculum effects (13) that may adversely elevate MIC values must still be considered if cefepime is chosen as an alternative therapy against ESBL-KP strains. This strategy of focused utilization of a newer cephalosporin could reduce some selective pressures of carbapenem use among ESBL-KP and thus minimize the development of carbapenem-resistant strains. In addition, phenotypic characteristics appear to accurately differentiate two important endemic and epidemic groups of ESBL types (CTX-M series and SHV-like) in *K. pneumoniae* strains in Taiwan.

Acknowledgment

We thank Monto Ho, Microbial Infections Reference Laboratory, National Health Research Institutes, for providing subcultures of the strains from the Taiwan Surveillance of Antimicrobial Resistance collection.

Dr. Yu is the SENTRY Antimicrobial Surveillance Program Fellow for 2000-01 at the University of Iowa College of Medicine (Iowa City, Iowa). His research focuses on the detection (phenotypic and genotypic) and characterization of β -lactamases in gram-negative bacilli endemic and epidemic in Taiwan, where he is a member of the medical faculty at the China Medical College, Taichung.

References

1. Winokur PL, Canton R, Casellas JM, Legakis N. Variations in the prevalence of strains expressing an extended-spectrum β -lactamase phenotype and characterization of isolates from Europe, the Americas, and the Western Pacific Region. *Clin Infect Dis* 2001;32(Suppl 2):S94-103.
2. Jan IS, Hsueh PR, Teng LJ, Ho SW, Luh KT. Antimicrobial susceptibility testing for *Klebsiella pneumoniae* isolates resistant to extended-spectrum β -lactam antibiotics. *J Formos Med Assoc* 1998;97:661-6.
3. Siu LK, Lu PL, Hsueh PR, Lin FM, Chang S-C, Luh K-T, et al. Bacteremia due to extended-spectrum β -lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* in a pediatric oncology ward: clinical features and identification of different plasmids carrying both SHV-5 and TEM-1 genes. *J Clin Microbiol* 1999;37:4020-7.
4. Liu PY, Tung JC, Ke SC, Chen SL. Molecular epidemiology of extended-spectrum β -lactamase-producing *Klebsiella pneumoniae* isolates in a district hospital in Taiwan. *J Clin Microbiol* 1998;36:2759-62.
5. Yan JJ, Ko WC, Tsai SH, Wu HM, Jin YT, Wu JJ. Dissemination of CTX-M-3 and CMY-2 beta-lactamases among clinical isolates of *Escherichia coli* in southern Taiwan. *J Clin Microbiol* 2000;38:4320-5.
6. National Committee for Clinical Laboratory Standards. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Approved standard, 4th ed. M7-A5. Wayne (PA): The Committee, 2000.
7. National Committee for Clinical Laboratory Standards. Performance standards for antimicrobial susceptibility testing. Document M100-S11. Wayne (PA): The Committee, 2001.
8. Matthew M, Harris A, Marshall MG, Ross GW. The use of analytical isoelectric focusing for detection and identification of β -lactamases. *J Gen Microbiol* 1975;88:169-78.

9. Yu WL, Winokur PL, Von Stein DL, Pfaller MA, Wang JH, Jones RN. First description of *Klebsiella pneumoniae* harboring CTX-M β -lactamases (CTX-M-14 and CTX-M-3) in Taiwan. *Antimicrob Agents Chemother* 2002;46:1098-100.
10. Bauernfeind A, Casellas JM, Goldberg M, Holley M, Junwirth R, Mangold P, et al. A new plasmidic cefotaximase from patients infected with *Salmonella typhimurium*. *Infection* 1992;20:158-63.
11. Palucha A, Mikiewicz B, Hryniewicz W, Gniadkowski M. Concurrent outbreaks of extended-spectrum beta-lactamase-producing organisms of the family Enterobacteriaceae in a Warsaw hospital. *J Antimicrob Chemother* 1999;44:489-99.
12. Szabo D, Filetoth Z, Szentandrassy J, et al. Molecular epidemiology of a cluster of cases due to *Klebsiella pneumoniae* producing SHV-5 extended-spectrum β -lactamase in the premature intensive care unit of a Hungarian Hospital. *J Clin Microbiol* 1999;37:4167-9.
13. Tzouveleakis LS, Tzelepi E, Prinarakis E, Gazouli M, Katrahoura A, Giakkoupi P, et al. Sporadic emergence of *Klebsiella pneumoniae* strains resistant to cefepime and ceftiofime in Greek hospitals. *J Clin Microbiol* 1998;36:266-8.
14. Szabo D, Mathe A, Filetoth Z, Anderlik P, Rokusz L, Rozgonyi F. In vitro and in vivo activities of amikacin, cefepime, amikacin plus cefepime, and imipenem against an SHV-5 extended-spectrum β -lactamase-producing *Klebsiella pneumoniae* strain. *Antimicrob Agents Chemother* 2001;45:1287-91.

Address for correspondence: Ronald N. Jones, 345 Beaver Creek Centre, Suite A, North Liberty, IA 52317, USA; fax: 319-665-3371; e-mail: ronald-jones@jmlabs.com

OPPORTUNITIES FOR PEER REVIEWERS

The editors of Emerging Infectious Diseases seek to increase the roster of reviewers for manuscripts submitted by authors all over the world for publication in the journal. If you are interested in reviewing articles on emerging infectious disease topics, please e-mail your name, address, qualifications or curriculum vitae, and areas of expertise to eideditor@cdc.gov

At Emerging Infectious Diseases, we always request reviewers' consent before sending manuscripts, limit review requests to three or four per year, and allow 2-4 weeks for completion of reviews. We consider reviewers invaluable in the process of selecting and publishing high-quality scientific articles and acknowledge their contributions in the journal once a year.

Even though it brings no financial compensation, participation in the peer-review process is not without rewards. Manuscript review provides scientists at all stages of their career opportunities for professional growth by familiarizing them with research trends and the latest work in the field of infectious diseases and by improving their own skills for presenting scientific information through constructive criticism of those of their peers. To view the spectrum of articles we publish, information for authors, and our extensive style guide, visit the journal web site at www.cdc.gov/eid.

For more information on participating in the peer-review process of Emerging Infectious Diseases, e-mail eideditor@cdc.gov or call the journal office at 404-371-5329.