Three Drinking-Water–Associated Cryptosporidiosis Outbreaks, Northern Ireland

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Three recent drinking-water–associated cryptosporidiosis outbreaks in Northern Ireland were investigated by using genotyping and subgenotyping tools. One Cryptosporidium parvum outbreak was caused by the bovine genotype, and two were caused by the human genotype. Subgenotyping analyses indicate that two predominant subgenotypes were associated with these outbreaks and had been circulating in the community.

Human cryptosporidiosis is predominantly caused by the human and bovine Cryptosporidium parvum genotypes, which differ in host range; the former infects mostly humans under natural conditions, and the latter infects both humans and some farm animals such as cattle, sheep, and goats (1). In many geographic areas, both C. parvum transmission cycles can occur in humans, but the importance of each genotype as a source of human infection probably varies (2–4). Both genotypes have been involved in waterborne outbreaks of human cryptosporidiosis in the United States, Canada, and the United Kingdom (2,5,6).

From April 2000 to April 2001, three drinking-water–associated outbreaks of cryptosporidiosis occurred in Northern Ireland. These outbreaks were epidemiologically unrelated and originated from geographically separate areas. Concerns have been raised about a possible relationship between human and bovine C. parvum genotypes, and some farm animals such as cattle, sheep, and goats (1). In many geographic areas, both C. parvum transmission cycles can occur in humans, but the importance of each genotype as a source of human infection probably varies (2–4). Both genotypes have been involved in waterborne outbreaks of human cryptosporidiosis in the United States, Canada, and the United Kingdom (2,5,6).

The Study

The three drinking-water–associated outbreaks occurred in the greater Belfast area. Outbreak A occurred during April and May 2000; at least 129 cases were laboratory confirmed. Outbreak B occurred in August 2000, involving at least 117 cases. Outbreak C occurred in April 2001; at least 230 people were infected (7–9; unpub. data). An outbreak patient was defined as a person with microscopically confirmed Cryptosporidium infection who became ill during the outbreak period and who was a resident in the water supply areas. The attack rates for outbreaks A, B, and C were 34, 180, and 58 cases/100,000 persons, respectively. Outbreak B was thought to be caused by the ingestion of human sewage from a septic tank into the drinking-water distribution system and C from the ingestion of wastewater from a blocked drain.

For molecular analysis, 34, 42, and 44 microscopically positive stool samples from outbreaks A, B, and C, respectively, were used. One wastewater sample from a blocked drain implicated in outbreak C was also analyzed. Control isolates of the C. parvum genotypes were also included in the subgenotyping analysis. Fourteen control isolates were from sporadic C. parvum infections of the bovine genotype in a rural area in the west of Ireland about 100 miles from Belfast, where the water supply was entirely different. Ten control isolates were from sporadic C. parvum infections of the human genotype in northwest England during the same time as outbreak C.

C. parvum genotype in human fecal samples was first determined by a COWP gene-based PCR-RFLP tool (10). Oocyst suspensions were prepared from feces by using salt flotation (11). The oocysts were washed and resuspended in deionized water and stored at 4°C before use. To extract DNA, oocyst suspensions were incubated at 100°C for 60 minutes, digested with proteinase K (3 mg/mL) in lysis buffer at 56°C for 30 minutes, and extracted by spin-column filtration (QiAMP DNA kit, Qiagen, Crawley, UK). Extracted DNA was stored at -20°C before use. Genotypes were investigated by using the COWP gene primers cry15 and cry9 to amplify a 553-bp region, which was then subjected to endonuclease digestion by Rsal (10).

Genotypes were confirmed by using an SSU rRNA-based PCR-RFLP tool (12). Subgenotyping was done by sequence analysis of the GP60 gene (13). Before molecular analysis, the wastewater sample was processed by both salt flotation (11) and immunomagnetic separation (Dynal, Lake Success, NY), following the manufacturer-recommended procedures (14). Both genotyping and subgenotyping tools used nested PCR amplification of targeted genes. The primers used for GP60 were 5′-ATA GTC TCC GCT GTA TTC-3′ and 5′-TCC GCT GTA TTC TCA GCC-3′ for primary PCR and 5′-GGA AGG AAC GAT GTA TTC-3′ and 5′-GCA GAG GAA CCA GCA TC-3′ for secondary PCR. The PCR reaction contained 1X Perkin-Elmer (Norwalk, CN) PCR buffer, 3 mM MgCl2, 200 µM (each) deoxynucleoside triphosphate, 200 nM of the forward and reverse primers, 5 units of Taq polymerase, and 0.5–2 µL of DNA template (for primary PCR) or 2 µL of primary PCR product (for secondary PCR) in a total 100-µL reaction mixture. Each PCR reaction was then subjected to 35 cycles of denaturation at 94°C for 45 seconds, annealing at 50°C for 45
seconds, and extension at 72°C for 60 seconds, with an initial
denaturation at 95°C for 3 minutes and a final extension at
72°C for 10 minutes. PCR products were sequenced in both
directions on an AB3100 (Applied Biosystems, Foster City,
CA) with forward and reverse primers. An additional sequenc-
ing primer (5’-GAG ATA TAT CTT GGT GCG-3’) was used
in the sequencing of GP60 PCR products. We aligned the
study’s GP60 nucleotide sequences with each other and with
sequences from the GenBank database with GCG software
(Genetics Computing Group, Madison, WI). A neighbor-join-
ing tree was constructed from the aligned sequences as
described (15).

Thirty-three of the 34 stool samples from outbreak A were
amplified by both the COWP and SSU rRNA-based nested
PCRs. RFLP analysis of the PCR products showed that all 33
PCR-positive samples had the *C. parvum* bovine genotype.
Thirty-two of the 42 stool samples from outbreak B were also
positive by PCR, and all belonged to the *C. parvum* human
 genotype. Furthermore, in outbreak C, 36 of 44 samples had
the *C. parvum* human genotype, and 8 had the bovine geno-
type. After further epidemiological investigations, these eight
bovine genotypes, although submitted to the primary diagno-
sis laboratory at the same time as the human genotypes, were
considered contemporary sporadic cases and not part of out-
brea C. These patients did not live in the distribution area of
the water supply implicated in the outbreak. The patients lived
in County Down (South Down), whereas the outbreaks
occurred in south Antrimy and north Down. Results of the two
genotyping methods were in complete agreement in both
detection rates and genotyping result.

Subgenotype analyses of the GP60 gene showed that of the
30 stool isolates of the *C. parvum* bovine genotype examined
for outbreak A, 25 isolates belonged to a single GP60 sub-
genotype and 5 isolates belonged to another subgenotype. In
contrast, 14 samples of the *C. parvum* bovine genotype iso-
lated from sporadic cases of human cryptosporidiosis from the
west of Ireland, which were unrelated to any of the Northern
Ireland outbreaks, belonged to nine subgenotypes. Subgeno-
type analysis of 31 stool samples from outbreak B showed the
presence of only one subgenotype of the *C. parvum* human
 genotype. For outbreak C, all 36 *C. parvum* human genotype
stool isolates were identical to the subgenotype involved in
outbreak B. In addition, all eight *C. parvum* bovine genotype
stool isolates, which were contemporary with, but not from,
the area affected by the outbreak, were identical to the pre-
dominant subgenotype in outbreak A. The wastewater sample
from the blocked drain implicated as the cause of outbreak C
contained oocysts of the same subgenotype as the *C. parvum*
human genotype. Of the nine sporadic isolates of the *C. par-
vum* human genotype from northwest England, eight belonged
to the same subgenotype as the *C. parvum* human genotype
involved in outbreaks B and C (Figure). Most infected persons
each had only one genotype/subgenotype of *C. parvum*,
judged by the RFLP profile, the absence of underlying signal
in the chromatogram of the sequencing result, and at least five
independent PCR analyses of each sample. The SSU rRNA
technique can detect multiple *Cryptosporidium* parasites in
individual samples (16).

**Discussion**

Results of genotyping analysis support epidemiologic obser-
vations that these three drinking-water–associated outbreaks
of cryptosporidiosis in Northern Ireland were unrelated, although
they all occurred in the greater Belfast area over a 1-year period.
Outbreak A was caused by the *C. parvum* bovine genotype, and
outbreaks B and C were caused by the *C. parvum* human geno-
type. The occurrence of the *C. parvum* human genotype in
outbreaks B and C suggests that these two outbreaks were, at least
in part, caused by contamination of the drinking-water supply
by seepage of raw sewage and through wastewater into the
drinking water distribution systems, respectively. This finding
illustrates the value of timely genotyping analysis during out-
break investigations. The source of contamination is further
supported by subgenotyping analysis of the wastewater sample from the blocked drain that was epidemiologically implicated in outbreak C. This sample contained one subgenotype of the C. parvum human genotype indistinguishable from the subgenotype found in most infected persons.

The failure to detect Cryptosporidium in 10 of the microscopically positive samples in outbreak B was most likely not because of rare Cryptosporidium genotypes; the SSU rRNA technique is Cryptosporidium genus specific and detects all known Cryptosporidium spp. (12,14–16). The presence of PCR inhibitors in the extracted DNA may have prevented the detection of Cryptosporidium by PCR.

Results of subgenotyping analysis nevertheless indicate that the three recent cryptosporidiosis outbreaks in Northern Ireland were caused by two predominant subgenotypes of C. parvum that probably had been circulating in the community before the outbreaks. These two subgenotypes of C. parvum are also the most common subgenotypes found in Northern Ireland and northwest England. The human subgenotype was found in 8 of 9 sporadic isolates from northwest England and the bovine subgenotype in 4 of 14 isolates in another part of Ireland.

The two subgenotypes of the C. parvum bovine genotype found in outbreak A and concurrent with outbreak C have not been found in most other areas (3,4). The only C. parvum isolate identical to one of the subgenotypes is an unpublished sequence (AF2030016) deposited in GenBank (Figure). The source of the other genotype, however, is unknown. In contrast, the subgenotype of the C. parvum human genotype involved in outbreaks B and C has a wide geographic distribution, with isolates from United States, Canada, United Kingdom, Portugal, and Peru (3,4). This subgenotype, the most common subgenotype of the C. parvum human genotype found in the United States, was responsible for several waterborne and foodborne outbreaks of human cryptosporidiosis (3). This subgenotype has a worldwide distribution and is the cause of many outbreaks. Whether the wide distribution of this subgenotype of the C. parvum human genotype and apparent association with multiple outbreaks in geographically distinct areas result from unusual biologic fitness of this parasite is unknown.

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