**Weissella confusa Infection in Primate (Cercopithecus mona)**

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We describe the first systemic infection by *Weissella confusa* in a mona monkey (*Cercopithecus mona*) on the basis of microbiologic, molecular genetic, and histologic data. The same strain of *W. confusa*, as determined by pulsed-field gel electrophoresis, was isolated in pure culture from the brain, liver, spleen, and intestine of this primate, illustrating the clinical importance of the isolations.

*Weissella* microorganisms are gram-positive, catalase-negative cocccobacilli, which have been isolated from a wide variety of habitats such as soil, fresh vegetables, fermented foods, or meat and meat products (1,2). The genus *Weissella* is peculiar since it currently includes 11 validated species, but only *Weissella confusa* (basonym *Lactobacillus confusus*) and *W. cibaria* have been isolated from human or animal clinical sources. *W. cibaria* has been isolated from human bile and feces, the liver of a canary, and material from an infected ear in a dog (1). *W. confusa* has been isolated from feces of children with bacteremic infections (3) and liver transplants (4), and from the peritoneal fluids and abdominal walls of two patients (5). In animals, *W. confusa* has been isolated from necropsy specimens from a dog and from the ear of a dog with otitis (1). However, with the exception of a thumb abscess caused by *W. confusa* in a healthy 49-year-old man (6), the clinical significance of all other clinical isolates was not clearly established. This article describes the first well-documented systemic infection caused by *W. confusa* in a primate.

**Case Report**

A juvenile female mona monkey (*Cercopithecus mona*) was found dead without clinical signs of disease in the previous 24 h. The animal had no previous relevant medical history. The monkey was housed in a cage with another monkey, which formed part of a primate bioacoustic research unit. None of the other monkeys housed in the same research unit died or exhibited any clinical sign. The dead monkey was sent to the hospital of the veterinary school at the Complutense University in Madrid for necropsy. Postmortem examination showed the existence of congestion, edema, and petechial hemorrhages in most internal organs, especially marked in the brain, liver, and spleen, which are typical lesions associated with systemic infections. Samples from intestine, lung, liver, and brain were collected under aseptic conditions for microbiologic studies. Tissue samples were surface-plated on Columbia blood agar (bio-Mérieux España, S.A. Madrid) and incubated aerobically and under anaerobic conditions for 48 h at 37°C. Gram-positive, catalase-negative, facultative anaerobic cocccobacilli were isolated in pure culture from lung, liver, brain, and intestine. Biochemical characterization was achieved by using the commercial Rapid ID32 Strep version 2.0 system (bio-Mérieux España, S.A. Madrid) according to the manufacturer’s instructions. The four isolates had identical biochemical profile (numerical code 72007000000), being identified as *Leuconostoc* spp. Acid production from ribose, L-arabinose, and galactose was also tested by using phenol red base medium (Difco Laboratories, Detroit, MI), supplemented with 1% (w/v) sugar, after 48 h of incubation at 37°C. Antimicrobial susceptibility was tested by the microdilution method and haemophilus test medium with lysed horse blood (7) with a commercially prepared dehydrated panel (Sensititre, Trek Diagnostic System, East Grinstead, UK) as previously described (8). MICs (in µg/mL) were as follows: tetracycline, ≤1; amoxicillin, ≤0.25; trimethoprim, 32; erythromycin, ≤0.5; penicillin, ≤0.5; chloramphenicol, 8; ciprofloxacin, ≤0.25; and vancomycin ≥128. Resistance of *W. confusa* to vancomycin has been reported previously (4,6,7).

For histopathologic studies, tissues were fixed in 10% neutral-buffered formalin, embedded in paraffin, cut in 4-µm sections, and stained with hematoxylin and eosin. Histologic examination of the lungs, spleen, and liver showed the existence of inflammatory infiltrates composed mainly of neutrophils, and in lower proportion, of lymphocytes and macrophages (Figure 1), suggesting the existence of an acute septicemic process. Gram-positive cocccobacilli emboli were observed in some hepatic vessels, suggesting a hematogenous dissemination.

Identifying *Weissella* species by classic phenotypic methods can be difficult (1,9). Comparing the 16S rRNA gene sequences of bacterial species is a useful approach for the identifying unusual clinical isolates or those which cannot be properly identified by conventional phenotypic methods (10,11). The 16S rRNA gene of each isolate was amplified by polymerase chain reaction (PCR) and further sequenced to determine genotypic identity (12). The determined sequences consisted of approximately 1,400
nucleotides and were compared with the sequences of other gram-positive, catalase-negative species available in the GenBank database, by using the BLAST program (available from: URL: http://www.ncbi.nlm.nih.gov/BLAST). The 16S rRNA gene analysis indicated that the four isolates were genotypically identical, displaying the closest sequence similarity (99.9%) with *W. confusa* (accession no. AB023241). Sequence similarity with *W. cibaria* was 99.2%, which agrees with the high sequence similarity reported for both species (1). Overall results of the phenotypic characterization of the clinical isolates were consistent with those described for this species (13). Like *W. confusa*, clinical isolates were able to produce acid from ribose and galactose but not from L-arabinose, one of the few biochemical tests that can differentiate this species and *W. cibaria* (1). These results support the identification of the clinical isolates as *W. confusa*. *Weissella* microorganisms can be isolated as normal flora of the intestinal tract (1,14,15). Thus, an extraintestinal origin of the systemic infection is most likely.

*W. confusa* isolates were molecularly characterized by pulsed-field gel electrophoresis (PFGE), according to the specifications of Vela et al. (16) with the CHEF-DR III system (Bio-Rad Laboratories, Hercules, CA). The restriction enzymes *ApaI* (Promega UK Ltd., Southampton, UK) and *SmaI* (MBI Fermentas Vilinius, Lithuania) were used according to the manufacturer’s recommendations. These enzymes have been successfully used for the molecular typing of microorganisms closely related to *Weisella* (17). Gels were interpreted by standard criteria (18). All *W. confusa* isolates displayed indistinguishable macrorestriction patterns by PFGE with *SmaI* (data not shown) and...
Apal (Figure 2) restriction enzymes, demonstrating that the systemic infection was caused by a single strain of *W. confusa*.

Conclusions

*Weissella* are considered nonpathogenic microorganisms because most of the strains isolated from clinical samples have been obtained as mixed cultures without evidence of their clinical significance (1, 7). In this study, the same strain of *W. confusa*, as determined by PFGE, was isolated in pure culture from the brain, liver, and spleen; the isolations from these organs, together with the histopathologic data, illustrate the clinical importance of the isolations. These results generate further speculation about the potential of *W. confusa* as an opportunistic pathogen. This is the first well-documented study in which, by combining microbiologic, molecular genetic, and histologic, data, the clinical importance of the isolation of *W. confusa* and its implication in an animal infection is clearly established.

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References


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