Emerging Human Infectious Diseases: Anthroponoses, Zoonoses, and Sapronoses

To the Editor: The source of infection has always been regarded as an utmost factor in epidemiology. Human communicable diseases can be classified according to the source of infection as anthroponoses (when the source is an infectious human; interhuman transfer is typical), zoonoses (the source is an infectious animal; interhuman transfer is uncommon), and sapronoses (the source is an abiotic substrate, nonliving environment; interhuman transfer is exceptional). The source of infection is often the reservoir or, in ecologic terms, the habitat where the etiologic agent of the disease normally thrives, grows, and replicates. A characteristic feature of most zoonoses and sapronoses is that once transmitted to humans, the epidemic chain is usually aborted, but the clinical course might be sometimes quite severe, even fatal. An ecologic rule specifies that an obligatory parasite should not kill its host to benefit from the adapted long-term symbiosis, whereas an occasionally attacked alien host, such as a human, might be subjected to a severe disease or even killed rapidly by the parasite because no evolutionary adaptation to that host exists (1). In this letter, only microbial infections are discussed; metazoan invasion and infestations have been omitted.

Anthroponoses (Greek “anthrópos” = man, “nosos” = disease) are diseases transmissible from human to human. Examples include rubella, smallpox, diphtheria, gonorrhea, ringworm (Trichophyton rubrum), and trichomoniasis.

Zoonoses (Greek “zoon” = animal) are diseases transmissible from living animals to humans (2). These diseases were formerly called anthropozoonoses, and the diseases transmissible from humans to animals were called zooanthroponoses. Unfortunately, many scientists used these terms in the reverse sense or indiscriminately, and an expert committee decided to abandon these two terms and recommended “zoonoses” as “diseases and infections which are naturally transmitted between vertebrate animals and man” (3). A limited number of zoonotic agents can cause extensive outbreaks; many zoonoses, however, attract the public’s attention because of the high death rate associated with the infections. In addition, zoonoses are sometimes contagious for hospital personnel (e.g., hemorraghic fevers). Zoonotic diseases can be classified according to the ecosystem in which they circulate. The classification is either synanthropic zoonoses, with an urban (domestic) cycle in which the source of infection are domestic and synanthropic animals (e.g., urban rabies, cat scratch disease, and zoonotic ringworm) or exoanthropic zoonoses, with a sylvatic (feral and wild) cycle in natural foci of the agent (4) outside human habitats (e.g., arboviroses, wildlife rabies, Lyme disease, and tularemia). However, some zoonoses can circulate in both urban and natural cycles (e.g., yellow fever and Chagas disease). A number of zoonotic agents are arthropod-borne (5); others are transmitted by direct contact, alimentary (foodborne and waterborne), or aerogenic (airborne) routes; and some are rodent-borne.

Sapronoses (Greek “sapos” = decaying; “sapron” means in ecology a decaying organic substrate) are human diseases transmissible from abiotic environment (soil, water, decaying plants, or animal corpses, excreta, and other substrata). The ability of the agent to grow saprophytically and replicate in these substrata (i.e., not only to survive or contaminate them secondarily) are the most important characteristics of a sapronotic agent. Sapronotic agents thus carry on two diverse ways of life: saprophytic (in an abiotic substrate at ambient temperature) and parasitic (pathogenic, at the temperature of a homeotherm vertebrate host). Typical sapronoses are visceral mycoses caused by dimorphic fungi (e.g., coccidioidomycosis and histoplasmosis), “monomorphic” fungi (e.g., aspergillosis and cryptococcosis), certain superficial mycoses (Microsporum gypseum), some bacterial diseases (e.g., legionellosis), and protozoan (e.g., primary amebic meningoencephalitis). Intracellular parasites of animals (viruses, rickettsiae, and chlamydiae) cannot be sapronotic agents. The term “sapronosis” was introduced in epidemiology as a useful concept (6–8). For these diseases the expert committee applied the term “sapro-zoonoses,” defined as “having both a vertebrate host and a nonanimal developmental site or reservoir (organic matter, soil, and plants)” (3,9). However, the term sapronoses is more appropriate because animals are not the source of infection for humans. While anthroponoses and zoonoses are usually the domains for professional activities of human and veterinary microbiologists, respectively, sapronoses may be the domain for environmental microbiologists. The underdiagnosis rate for sapronoses is probably higher than that for anthroponoses and zoonoses, and an increase should be expected in both incidence and number of sapronoses. Legionellosis, Pontiac fever, nontuberculous mycobacterioses, and primary amebic meningoencephalitis are a few sapronoses that have emerged in the past decade. In addition, the number of opportunistic infections in immunosuppressed patients has grown markedly; many of these diseases and some nosocomial infections are, in fact, also sapronoses.

As with any classification, grouping human diseases in epidemiologic categories according to the source of infection has certain pitfalls. Some arthropod-borne diseases (urban yellow fever, dengue, epidemic typhus, tickborne relapsing fever, epidemic relapsing fever, and malaria) might be regarded as anthroponoses rather than zoonoses because the donor of the infectious blood for the vector is an infected human and not a vertebrate animal. However, the human infection is caused by an (invertebrate) animal in which the agent replicates, and the term zoonoses is preferred. HIV is of simian origin with a sylvatic cycling among wild primates and accidental infection of humans who hunted or ate them; the human disease (AIDS)
might thus have been regarded as a zoonosis in the very first phase but later has spread in the human population as a typical anthroponosis and caused the present pandemic. Similarly, pandemic strains of influenza developed through an antigenic shift from avian influenza A viruses. For some etiologic agents or their genotypes, both animals and humans are concurrent reservoirs (hepatitis virus E, Norwalk-like calicivirus, enteropathogenic Escherichia coli, Pseudomonas aeruginosa, Cryptosporidium, Giardia, and Cyclospora); these diseases might conditionally be called anthropozoonoses. Other difficulties can occur when classifying diseases caused by sporulating bacteria (Clostridium and Bacillus); their infective spores survive in the soil or in other substrata for very long periods, though they are usually produced after a vegetative growth in the abiotic environment, which can include animal carcasses. These diseases should therefore be called sapronoses. For some other etiologic agents, both animals and abiotic environment can be the reservoir (Listeria, Erysipelothrix, Yersinia pseudotuberculosis, Burkholderia pseudomallei, Rhodococcus equi), and the diseases might be, in fact, called saprozoones (not sensu 9) in that their source can be either an animal or an abiotic substrate.

For a concise list of anthropo-, zoo-, and sapronoses, see the online appendix available from: URL: http://www.cdc.gov/ncidod/EID/vol9no3/02-0208-app.htm.

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Multidrug-Resistant Shigella dysenteriae Type 1: Forerunners of a New Epidemic Strain in Eastern India?

To the Editor: Multidrug-resistant Shigella dysenteriae type 1 caused an extensive epidemic of shigellosis in eastern India in 1984 (1). These strains were, however, sensitive to nalidixic acid, and clinicians found excellent results by using it to treat bacillary dysentery cases. Subsequently, in 1988 in Tripura, an eastern Indian state, a similar outbreak of shigellosis occurred in which the isolated strains of S. dysenteriae type 1 were even resistant to nalidixic acid (2). Since then, few cases of shigellosis have occurred in this region, and S. dysenteriae type 1 strains are scarcely encountered (3). In other regions of the world, especially in Southeast Asia, low-level resistance to fluoroquinolones in Shigella spp. has been observed for some time (4,5).

After a lapse of almost 14 years, clusters of patients with acute bacillary dysentery were seen at the subdivisional hospital, Diamond Harbour, in eastern India. No cases of dysentery had been reported during the comparable period in previous years. A total of 1,124 case-patients were admitted from March through June 2002. The startling feature of these infections was their unresponsiveness to even the newer fluoroquinolones such as norfloxacin and ciprofloxacin, the drugs often used to treat shigellosis. Clinicians tried various antibiotics, mostly in combinations, without benefit. Clinicians also randomly used anti-amoebic drugs without success.

An investigating team collected nine fresh fecal samples from dysentery patients admitted to this hospital; 4 (44%) yielded S. dysenteriae type 1 on culture. For isolation of Shigella spp., stool samples were inoculated into MacConkey agar and Hektoen Enteric agar (Difco, Detroit, MI), and the characteristic colonies were identified by standard biochemical methods (6). Subsequently, serogroups and serotypes were determined by visual inspection of slide agglutination tests with commercial antisera (Denka Seiken, Tokyo). Antimicrobial susceptibility testing was performed by an agar diffusion disk method, as recommended by the National Committee for Clinical Laboratory Standards (7). Results showed that the organisms were resistant to all commonly used antibiotics, including the fluoroquinolones (norfloxacin and ciprofloxacin) but were sensitive to ofloxacin. On our advice, the clinicians used ofloxacin with good results.

A similar outbreak of S. dysenteriae type 1 occurred in the northern part of West Bengal in eastern India among tea garden laborers from April 2002 to May 2002; 1,728 persons were affected (attack rate of 25.6%). Sixteen persons died. The isolated S. dysenteriae type 1 strains were found intermediately sensi-
Appendix: Important Anthroponoses, Zoonoses, and Sapronoses

Anthroponoses

Measles*; rubella; mumps; influenza; common cold; viral hepatitis; poliomyelitis; AIDS*; infectious mononucleosis; herpes simplex; smallpox; trachoma; chlamydial pneumonia and cardiovascular disease*; mycoplasmal infections*; typhoid fever; cholera; peptic ulcer disease*; pneumococcal pneumonia; invasive group A streptococcal infections; vancomycin-resistant enterococcal disease*; meningococcal disease*; whooping cough*; diphtheria*; *Haemophilus* infections* (including Brazilian purpuric fever*); syphilis; gonorrhea; tuberculosis* (multidrug-resistant strains); candidiasis*; ringworm (*Trichophyton rubrum*); *Pneumocystis* pneumonia* (human genotype); microsporidial infections*; cryptosporidiosis* (human genotype); giardiasis* (human genotype); amebiasis; and trichomoniasis.

Zoonoses Transmitted by Direct Contact, Alimentary (Foodborne and Waterborne), or Aerogenic (Airborne) Routes

Rabies; hemorrhagic fever with renal syndrome*; hantavirus pulmonary syndrome*; Venezuelan*; Brazilian*; Argentinian and Bolivian hemorrhagic fevers; Lassa; Marburg; and Ebola hemorrhagic fevers*; Hendra and Nipah hemorrhagic bronchopneumonia*; hepatitis E*; herpesvirus simiae B infection; human monkeypox*; Q fever; sennetsu fever; cat scratch disease; psittacosis; mammalian chlamydiomiasis*; leptospirosis; zoonotic streptococcosis; listeriosis; erysipeloid; campylobacteriosis*; salmonellosis*; hemorhagic colitis*; hemolytic uremic syndrome*; yersiniosis; pseudotuberculosis; sodoku; Haverhill fever; brucellosis*; tularemia*; glands; bovine and avian tuberculosis*; zoonotic ringworm; toxoplasmosis; and cryptosporidiosis* (calf genotype 2).

Zoonoses Transmitted by Hematophagous Arthropods

**Hard ticks (Ixodidae)**

Russian spring-summer encephalitis; Central European encephalitis; louping ill; Kyasanur Forest disease; Powassan; Crimean-Congo hemorrhagic fever*; Colorado tick fever; Rocky Mountain spotted fever; boutonneuse fever; African tick typhus*; other rickettsial fevers*; human granulocytic ehrlichiosis*; Lyme disease*; tularemia; and babesiosis.

**Soft ticks (Argasidae)**

Tickborne relapsing fever

**Mites (Trombiculidae, Dermanyssidae)**

Scrub typhus; rickettsialpox

**Lice (Anoplura)**

Epidemic typhus; trench fever*; and epidemic relapsing fever
**Triatomine Bugs (Triatominae)**

Chagas disease

**Sandflies (Phlebotominae)**

Sandfly fever; vesicular stomatitis; Oroya fever; and leishmaniasis

**Mosquitoes (Culicidae)**

Eastern; Western; and Venezuelan equine encephalomyelitis; Sindbis fever; Chikungunya and O’nyong nyong fevers*; Ross River epidemic polyarthritis*; Japanese encephalitis*; West Nile fever*; St. Louis encephalitis; yellow fever; dengue/dengue hemorrhagic fever*; Murray Valley encephalitis; California encephalitis; Rift Valley fever*; and malaria*

**Biting Midges (Ceratopogonidae)**

Oropouche fever; vesicular stomatitis

**Tsetse-flies (Glossinidae)**

African trypanosomiasis

**Fleas (Siphonaptera)**

Murine typhus*; cat-scratch fever*; plague

**Sapronoses**

Chlamydia-like pneumonia* (amoebic endosymbionts Parachlamydia acanthamoebae and other Parachlamydiaceae); tetanus; gas gangrene (Clostridium perfringens; C. septicum; C. novyi); intestinal clostridiosis* (C. difficile; C. perfringens); botulism; food poisoning* (Bacillus cereus); anthrax; vibrio gastroenteritis* or dermatitis (Vibrio parahaemolyticus; V. vulnificus); nosocomial Klebsiella pneumoniae and Pseudomonas aeruginosa bacteremia* (including antibiotic-resistant strains); bacterial infections associated with cystic fibrosis* (Burkholderia cepacia; Ralstonia spp.); melioidosis* (B. pseudomallei); legionellosis* and Pontiac fever* (Legionella pneumophila; L. micdadei; and other spp.); atypical bacterial meningitis and sepsis* (Chryseobacterium meningosepticum); acinetobacter bacteremia* (Acinetobacter calcoaceticus; A. baumannii; A. radioresistens); corynebacterial endocarditis* (Corynebacterium serosum; C. amycolatum and other non-diphtheriae corynebacteria); rhodococcosis* (Rhodococcus equi); possibly leprosy (some strains of Mycobacterium leprae were detected as living saprophytically in wet moss habitats); Buruli ulcer disease* (M. ulcerans); mycobacterial diseases other than tuberculosis* (M. kansasii; M. xenopi; M. marinum; M. haemophilum; M. fortuitum; M. scrofulaceum; M. abscessus; and other spp.); nocardiosis (Nocardia asteroides; N. brasiliensis); actinomycetom (Actinomadura madurae; A. pelletieri; Streptomyces somaliensis); dermatophytosis (Microsporum gypseum); histoplasmosis*
(Histoplasma capsulatum; H. duboisii); blastomycosis (Blastomyces dermatitidis); emmonsiosis (Emmonsia crescens; E. parva); paracoccidioidomycosis (Paracoccidioides brasiliensis); coccidioidomycosis* (Coccidioides immitis); sporotrichosis (Sporothrix schenckii); cryptococciosis* (Cryptococcus neoformans); aspergillosis (Aspergillus fumigatus); mucormycosis (Absidia corymbifera and some other Mucorales); entomophthoromycosis (Basidiobolus; Conidiobolus; and Entomophthora spp.); maduromycetom (Madurella mycetomatis; M. grisea; Pseudoallescheria boydii; Leptosphaeria senegalensis; Neotestudina rosatii); chromoblastomycosis (Phialophora verrucosa; Exophiala jeanselmei; Fonsecaea compacta; F. pedrosoi; Cladosporium carionii; Rhinocladiella aquaspersa); phaeohyphomycosis (Wangiella dermatitidis; Dactylaria gallopava; Exophiala spinifera); fusariosis* (Fusarium oxysporum; F. solani); primary amebic meningoencephalitis* (Naegleria fowleri); and amoebic keratitis or chronic granulomatous amoebic meningoencephalitis* (Acanthamoeba castellanii; A. polyphaga).

1 Emerging and reemerging diseases are marked with an asterisk.