

remains nonpathogenic for humans, although a case of human infection without clinical signs has been described in the Czech Republic (9).

Blood serum samples from 710 randomly selected persons >20 years of age from the Czech Republic were screened for antibodies against Puumala and Hantaan antigens with commercial enzyme-linked immunosorbent assay (ELISA) sets manufactured by PROGEN (Biotechnik, Heidelberg, Germany). The Hantaan antigen was used because of its antigenic relatedness with Dobrava virus, which is not included in available commercial ELISA sets.

Five participants showed immunoglobulin (Ig) G reactivity to Hantaan virus (cross-reactive with Dobrava antigen), and two participants tested positive for both IgG and IgM antibodies. Two other persons showed IgM reactivity alone. These findings indicate that as many as seven (1.0%) study participants showed reactivity to Hantaan antigen. Eight persons showed IgG reactivity to Puumala antigen, none of them IgM positive. Altogether, 10 (1.4%) study participants were reactive to Puumala antigen. Three persons showed reactivity to both antigens tested.

A total of 1,494 small mammals of different Czech regions were screened with ELISA for hantavirus antigen in the lungs. The antigen was detected in the lungs of 101 animals; the highest positivity rate was in Common Voles (*Microtus arvalis*). The difference in positivity between male and female voles was not statistically significant. The positivity rate was markedly associated with rodent size. With the use of molecular genetic methods (polymerase chain reaction), genotype Tula was identified as the causative agent of infection in rodents. Genotype Puumala was identified in bank voles (*Clethrionomys glareolus*) in Moravia. Nucleotide sequences of Dobrava genotype were identified in southern Bohemian rodents

(K. Krivanec, pers. comm.).

In the Czech Republic, Tula virus is the most frequent hantavirus circulating in Common Voles. This agent is not pathogenic for humans. The hantavirus seroprevalence rate in the adult population of the Czech Republic is close to 1%. Dobrava and Puumala viruses are causative agents of these infections in humans.

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References

1. Lee HW, Lee PW, Johnson KM. Isolation of the etiologic agent of Korean hemorrhagic fever. *J Infect Dis* 1978;137:298-308.
2. Grešikova M, Rajcáni J, Sekeyova M, Brummer-Korvenkontio M, Kozuch O, Labuda M, et al. Haemorrhagic fever virus with renal syndrome in small rodents in Czechoslovakia. *Acta Virol* 1984;28:416-21.
3. Danes L. [Hemorrhagic fever with renal syndrome (HFRS)] (in Czech). *Acta Hygienica, Epidemiologica et Microbiologica* 1985;15:52-7.
4. Pilaski J, Zöller L, Blenk H. Hämorrhagisches Fieber mit renalem Syndrom (HFRS): eine durch Nagetiere übertragene Nephropathie des Menschen. *Wehrmedizinische Monatschrift* 1986;10:435-44.
5. Kobzik J, Danes L. [Laboratory-confirmed cases of hemorrhagic fever with renal syndrome which occurred in Breclav 1989-1990] (in Czech). *Ceskoslovenska Epidemiologie, Mikrobiologie a Imunologie* 1992;41:65-8.
6. Petru K, Pejcoch M, Monhart V, Matyasov al. [Hemorrhagic fever with renal syndrome] (in Czech) *Cas Lek Cesk* 1997;136:739-40.
7. Plyusnin A, Cheng Y, Vapalahti O, Pejcoch M, Unar J, Jelinkova Z, et al. Genetic variation in Tula hantaviruses: sequence analysis of the S and M segments of strains from central Europe. *Virus Res* 1995;39:237-50.
8. Sibold C, Meisel H, Lundkvist, Schulz A, Cifire F, Ulrich R, et al. Short report: simultaneous occurrence of Dobrava, Puumala, and Tula hantaviruses in Slovakia. *Am J Trop Med Hyg* 1999;61:409-11.
9. Vapalahti O, Lundkvist A, Kukkonen SK, Cheng Y, Gilljam M, Kanerva M, et al. Isolation and characterization of Tula virus, a distinct serotype in the genus Hantavirus, family *Bunyaviridae*. *J Gen Virol* 1996;77:3063-7.

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Antibiotics and Airline Emergency Medical Kits

To the Editor: Medical events during airline flights have drawn some attention in recently published articles and letters (1-4). We would like to share our experience of meningococemia/ and meningococcal meningitis during a transatlantic flight.

In June 2000, a 20-year-old student with a mild viral illness (diagnosed before the flight) boarded a flight from Tel-Aviv, Israel, to Newark, New-Jersey, USA (approximate flight time, 11-12 hours), with a tour group of college-age students and their chaperones. We, a neonatologist and a neonatal intensive care nurse, were on the same flight to later transport a prematurely born infant from the United States back to Israel.

About 90 minutes before landing in New Jersey, the chief flight attendant asked me (B.B-O.) to check the passenger, who said he did not feel well. His medical history indicated no past illness, which was corroborated by the director of the tour group. The patient reported general malaise and numbness in his feet. In the 2 weeks before the flight, he had traveled in Israel, visiting cities, caves, and mountains. He and his group had slept in different hostels in those areas.

On examination, he was fully conscious, and his blood pressure and

pulse rate were normal. He had a blue-purple skin rash, particularly on the upper extremities. The rash worsened in the course of 20 minutes and resembled the “blueberry muffin-like” rash described in other pathologic conditions. Considering a diagnosis of either tick-borne or meningococcal disease, I decided to give the patient the first dose of antibiotics after obtaining a verbal consent from him and from the head of the group. I also asked the crew to have an ambulance and a physician waiting for us at the destination airport.

When we checked the emergency medical kit, we found that it did not contain any antibiotics. For our transport mission, we had two ampules of cefotaxime, 2 g each, one of which we gave the patient. After we landed, an ambulance crew (which did not include a physician) took the patient to the nearest hospital. The patient died 2 hours later in the hospital emergency department. His laboratory tests showed meningococcal meningitis and meningococemia. The Centers for Disease Control (CDC),

the airline company, and Israel’s Ministry of Health notified all close contacts of the patient in Israel and during the transatlantic flight, including everyone in the tour group, and recommended that they be given chemoprophylaxis.

CDC has received 21 reports about air travel-associated meningococcal disease in 2 years; in 5 reports, the symptoms began before the plane arrived at its destination (5). However, advance notice of the symptoms was given only in our case. Although one case is not enough to substantiate recommendations, we believe that the appropriate authorities should require airline companies to add a broad-spectrum antibiotic preparation to the emergency kit. This drug should be used only when aircraft diversion is not possible and when the diagnosis is clinically identified or highly suspected.

We still wonder whether an earlier intervention and treatment with a more appropriate on-board antibiotic treatment would have saved this young man.

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References

1. Gendreau MA., DeJohn C. Responding to medical events during commercial airline flights. *N Engl J Med* 2002;346:1067-73.
2. Baevsky R. Medical events during airline flights. *N Engl J Med* 2002;347:535.
3. Ross SC. Medical events during airline flights. *N Engl J Med* 2002;347:536-535.
4. Roth WT. Medical events during airline flights. *N Engl J Med* 2002;347:535.
5. Centers for Disease Control and Prevention. Exposure to patients with meningococcal disease on aircrafts—United States, 1999–2001. *MMWR Morb Mortal Wkly Rep* 2001;50:485-9.

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The image shows a screenshot of the CDC Emerging Infectious Diseases Journal homepage. The page features a search bar, a navigation menu, and several article highlights. A large, stylized graphic with the text 'SEARCH EID ONLINE' is overlaid on the right side of the screenshot. Below the graphic, the URL 'www.cdc.gov/eid' is displayed in a large, bold font.