Infections caused by community-acquired (CA)-methicillin-resistant \textit{Staphylococcus aureus} (MRSA) have been reported worldwide. We assessed whether any common genetic markers existed among 117 CA-MRSA isolates from the United States, France, Switzerland, Australia, New Zealand, and Western Samoa by performing polymerase chain reaction for 24 virulence factors and the methicillin-resistance determinant. The genetic background of the strain was analyzed by pulsed-field gel electrophoresis (PFGE) and multi-locus sequence typing (MLST). The CA-MRSA strains shared a type IV SCC\textit{mec} cassette and the Panton-Valentine leukocidin locus, whereas the distribution of the other toxin genes was quite specific to the strains from each continent. PFGE and MLST analysis indicated distinct genetic backgrounds associated with each geographic origin, although predominantly restricted to the \textit{agr}3 background. Within each continent, the genetic background of CA-MRSA strains did not correspond to that of the hospital-acquired MRSA.

\textbf{Materials and Methods}

\textbf{Bacterial Strains}

A total of 117 different isolates of CA-MRSA were examined. Community acquisition was defined as growth of the isolates within 48 hours after hospital admission in patients who had no risk factors for nosocomial acquisi-
tion, including no hospitalizations or nursing home residence in the year before admission. Thirty-three isolates were from the United States. The U.S. isolates all belonged to the previously identified major CA-MRSA clonal group and included strain MW2 (2). They originated from 12 different facilities in Minnesota; two isolates were from North Dakota. Sixty-seven isolates originated from Europe (61 isolates from France and 6 from Switzerland). The French CA-MRSA isolates originated from 10 different hospitals located throughout France, and the Swiss isolates were from the Geneva area. Seventeen isolates were from Oceania and included two different clones: 1) 13 isolates corresponded to the Western Samoan phage pattern strains, also designated the Southwest Pacific clone (9) and were isolated from Australia (8 isolates), New Zealand (4 isolates), and Western Samoa (1 isolate) (5); 2) 4 other isolates from Australia belonged to a recently described clone designated as the Queensland clone (10). The Southwest Pacific clone has two distinct phage-typing patterns known as Western Samoan Phage Pattern (WSPP) 1 and 2 (5). The Queensland clone, which cannot be phage typed, was first detected in the southeast Queensland city of Ipswich in 2000 (10). Most isolates were from primary skin and soft tissue infections, with some cases of bacteremia and at least five cases of necrotizing pneumonia (2,3,9,11,12).

A set of representative hospital-acquired (HA)MRSA isolates was included in the study: 24 from France and 33 from the United States. French HA-MRSA isolates corresponded to those of the major clones isolated throughout France as described by Lelievre et al. (13). HA-MRSA isolates were recovered from the same 12 different facilities that CA-MRSA. HA-MRSA from the United States originated from the same geographic area as the CA-MRSA.

Antimicrobial Susceptibility Testing

The MICs of benzyl-penicillin, oxacillin, gentamicin, tobramycin, kanamycin, chloramphenicol, tetracycline, minocycline, erythromycin, lincomycin, pristinamycin, fusidic acid, rifampicin, ofloxacin, co-trimoxazole, linezolid, mupirocin, vancomycin, and teicoplanin were determined for selected isolates (46 European, 22 U.S., and 13 Oceanian isolates) by using the standardized agar dilution technique as recommended by the French Society for Microbiology (14).

Detection of Accessory Genes by PCR

Using polymerase chain reaction (PCR), we determined the presence of accessory gene regulator (agr) allele group (1–4), SCCmec element (I–IV, according to the designation of Oliveira [15]), and 22 specific staphylococcal virulence genes (including 16 super-antigenic toxins, 3 hemolysins, and 3 leukocidins), as described previously (16).

Amplification of gyrA was used as a quality control of each DNA extract and the absence of PCR inhibitors. S. aureus strains Fri 913 (sea, see, sec, tst, lukE lukD, sek, sel, sep, and hlg), Fri 1151m (sed, sej, lukE lukD, hlgv, and hlb), ATCC 14458 (SCCmec II), NCTC 7428 (tst, lukM, seg, sei, sem, sen, sef, lukE lukD, hlgv, and hlb), A92 0211 (seg, sei, sem, sen, eta, etb, lukE lukD, and hlgv), RN6390 (lukE lukD, hlgv, hlb, and agrI), RN6607 (sed, seg, sei, sem, sen, sef, lukE lukD, hlgv, and agr2), RN8465 (seg, sei, sem, sen, see, tst, hlg, and agr3), RN4850 (seg, sei, sem, sen, see, eta, etb, lukE lukD, hlgv, and agr3), RN 6911(lukE lukD, hlgv, hlb, agr null), E-1 (seg, sei, sem, sen, see, lukE lukD, eta, hlgv, edinB and C), ATCC 49775 (seg, sei, sem, sen, see, lukS lukF, and hlg), and ATCC 51811 (FRI 569) (seg, lukE lukD, hlb, and hlgv) were used as positive controls for PCR (17,18). S. aureus COL (SCCmec I), PER34 (SCCmec IA), BK2464 (SCCmec II), ANS46 (SCCmec III), HU25 (SCCmec IIIA), and IDE288 (SCCmec IV) were used as controls for characterization of the mec element according to Oliveira and de Lencastre (15).

The overall genetic background of the isolates was evaluated: 1) by digesting whole cell DNA with Smal macrorestriction enzyme and determining the fragment-size patterns obtained on pulsed-field gel electrophoresis (PFGE) using a contour-clamped homogeneous electric field system on a CHEF DR-II apparatus (Bio-Rad Laboratories, Marnes-la-Coquette, France) as previously described (19). Resolved macrorestriction patterns were compared as recommended by Tenover et al. (20). Isolates differing by up to three fragments were considered as subtypes of a given clonal type. MLST was performed as described by Enright et al. (21). Briefly, seven housekeeping genes were used in the scheme; for each isolate, the alleles at each of the seven loci defined the allelic profile, which corresponded to a sequence type (ST). ST designations were those assigned by the MLST database (available from: URL: http://www.mlst.net).

Results

Distribution of Accessory Genes from MRSA in Three Continents

Overall, we detected 12 different virulence genes or gene clusters among the 117 isolates (Table 1). Two gene loci were common to CA-MRSA isolates from all locations. Meticillin resistance was conferred in all 117 CA-MRSA isolates by the truncated SCCmec type IV element, and all the isolates contained the PVL locus. In addition, 112 isolates harbored the related lukE-lukD genes of another leukocidin frequently recovered from patients with all types of staphylococcal infections (3). Most (113 [97%] of 117) isolates were of agr type 3.
The distribution of other genes varied by the continent of origin. The European and Southwest Pacific isolates had consistent, relatively simple, patterns of virulence-associated genes. In addition to the SCCmec type IV element, PVL genes, and lukE-lukD, all the European isolates were positive for hlg-v (the γ-hemolysin variant gene) and the Southwest Pacific isolates were positive for hlg (the γ-hemolysin gene) and egc (the enterotoxin gene cluster coding for the enterotoxins seg, sei, sem, sen, and seo) (Table 1). The Queensland isolates had not been tested positive for the toxin genes, except the PVL locus. In contrast, considerable variability existed among the 33 U.S. isolates. As for the European isolates, the U.S. isolates showed both the presence of hlg-v and the absence of hlg. However, seven other toxins (the enterotoxins sea–sed, sej, she, and sek), which were absent in non–U.S. isolates, were variably found in up to 29% (3% to 87%) of these 33 isolates (Table 1).

**CA-MRSA Antibiotic Susceptibility Profiles**

MICs were determined on a selected number of isolates (81) that corresponded to the different PFGE patterns. Overall, CA-MRSA isolates from all locations were susceptible to numerous antimicrobial drugs, including tobramycin, gentamicin, lincomycin, pristinamycin, minocycline, chloramphenicol, ofloxacin, vancomycin, teicoplanin, fosfomycin, rifampin, co-trimoxazole, and linezolid (Table 2). For the purposes of assessing differences between locations, isolates from the United States and Oceania were combined since their susceptibility profiles were similar (Table 2). Minor heterogeneity in erythromycin and mupirocin susceptibility was noted among European isolates. In contrast, U.S. and Oceanian isolates were uniformly susceptible to these antibiotics (Table 2).

---

**Table 1. Distribution of virulence and resistance determinants in 117 CA-MRSA isolates from three continents**

<table>
<thead>
<tr>
<th>Genesb</th>
<th>France-Switzerlandc</th>
<th>USA</th>
<th>Oceania</th>
<th>Australia</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sequence type</td>
<td>n=67 (%)</td>
<td>n=29 (%)</td>
<td>n=4 (%)</td>
<td>Southwest Pacific clone n=13 (%)</td>
<td>Queensland clone n=4 (%)</td>
</tr>
<tr>
<td>PFGE pattern</td>
<td>A1-7</td>
<td>B1-5</td>
<td>D1 &amp; F1</td>
<td>C1-3</td>
<td>E1</td>
</tr>
<tr>
<td>agr type</td>
<td>3</td>
<td>3</td>
<td>1</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>SCC IV</td>
<td>67 (100)</td>
<td>29 (100)</td>
<td>4 (100)</td>
<td>13 (100)</td>
<td>13 (100)</td>
</tr>
<tr>
<td>Leukocidins PVL genes</td>
<td>67 (100)</td>
<td>29 (100)</td>
<td>4 (100)</td>
<td>13 (100)</td>
<td>13 (100)</td>
</tr>
<tr>
<td>lukE-lukD</td>
<td>67 (100)</td>
<td>29 (100)</td>
<td>3 (75)</td>
<td>13 (100)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Hemolysinsc</td>
<td>hlg</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>13 (100)</td>
</tr>
<tr>
<td>hlg-v</td>
<td>67 (100)</td>
<td>29 (100)</td>
<td>4 (100)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>hlb</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>1 (25)</td>
<td>0 (0)</td>
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<tr>
<td>Enterotoxins sea</td>
<td>0 (0)</td>
<td>23 (79)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>seb</td>
<td>0 (0)</td>
<td>8 (28)</td>
<td>1 (25)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>sec</td>
<td>0 (0)</td>
<td>20 (69)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>sed-sej</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>3 (75)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>seh</td>
<td>0 (0)</td>
<td>29 (100)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>sek</td>
<td>0 (0)</td>
<td>24 (83)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>egc</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>13 (100)</td>
<td>0 (0)</td>
</tr>
</tbody>
</table>

The distribution of CA-MRSA isolates from different continents is presented.

CA-MRSA isolates from different continents varied in their genetic background. The Queensland clone had the same E1 pattern. The phylogenetic tree shown on the left side of the figure confirms the diversity in PFGE patterns between the CA-MRSA from different continents.
al interpretation of PFGE results.\(^{a}\) Capital letters indicate macrorestriction types based on UPGMA clustering. The scale indicates the level of pattern similarity. Dice coefficients of correlation and to generate a dendrogram by the unweighted pair group method using arithmetic averages (UPGMA) clustering. The scale indicates the level of pattern similarity. Capital letters indicate macrorestriction types based on visual interpretation of PFGE results.

The main difference between the two groups was their susceptibility to kanamycin, tetracycline, and fusidic acid, with the European group of isolates being more resistant than the U.S. and Oceanian group (Table 2).

Comparison of CA-MRSA with HA-MRSA

Using PFGE to compare CA-MRSA isolates with representative HA-MRSA isolates from both France (24 isolates) and the United States (33 isolates), we found that the last isolates grouped into lineages that clearly differed from any of the CA-MRSA isolates of the same continent (not shown). None of the HA-MRSA harbored PVL genes or the SCCmec IV element. Moreover, all U.S. HA-MRSA had SCCmec type II element; most (31 of 33) of these isolates were agr type 2, and the remaining 2 were type 1. Conversely, all French HA-MRSA had an unknown SCCmec element according to the method of Oliveira (15). These strains were further designated as SCCmec IVc by Hiramatsu (22). Twenty-three of the 24 French HA-MRSA isolates were agr type 1, and only 1 was a type 2. None of the HA-MRSA tested was agr type 3, the predominant type for CA-MRSA. In addition, unlike the PVL genes, luke-leukD leukocidin genes were found in most HA-MRSA (95%) as well as in CA-MRSA.

Analysis of Genetic Background by MLST

Twenty-one representative isolates of each PFGE pattern of CA-MRSA were further characterized by MLST. Overall, the results perfectly matched those of the PFGE with a unique ST corresponding to each group of related PFGE patterns (Table 1). The STs of CA-MRSA were compared with those in the MLST database (Table 3). Within each continent, the most frequent STs of CA-MRSA (i.e., ST1 for the U.S. clone, ST30 for the European clone, and ST80 for the European clone)
Discussion

The characterization of 117 CA-MRSA isolates from three continents indicated four major findings. First, only two genes were unique to CA-MRSA isolates and shared by isolates from all three continents: a type IV SCC\(\text{mec}\) cassette (further designated IVa by Okuma et al. [4]) and the PVL locus. Otherwise, the distribution of the other toxin genes was continent-specific. This finding suggests that PVL and SCC\(\text{mec}\) type IV may confer a selective advantage for community-based MRSA pathogens. Second, CA-MRSA isolates were generally susceptible to most of antibiotics tested apart from \(\beta\)-lactams, although European isolates appeared more resistant (i.e., to kanamycin, tetracycline, and fusidic acid) than U.S. and Oceanian isolates. Third, the genetic background of CA-MRSA organisms was different in each of the three continents (Table 3). Fourth, MLST and PFGE analysis showed that with- in a continent, the genetic background of CA-MRSA strains did not correspond to that of the HA-MRSA in the same continent, suggesting that CA-MRSA did not emerge from local HA-MRSA.

The STs of CA-MRSA clones were not related to the STs of any described pandemic clones of MRSA, such as the Archaic clone (ST250 with a SCC\(\text{mec}\) I element), the Iberian clone (ST247 with a SCC\(\text{mec}\) IA element), the New York/Japan clone (ST5 with a SCC\(\text{mec}\) II element), the Hungarian clone (ST239 with a SCC\(\text{mec}\) III element), the Brazilian clone (ST239 with a SCC\(\text{mec}\) IIIA element), or the pediatric clone (ST5 with a SCC\(\text{mec}\) IV element) (23). However, analysis of the MLST database indicated that the CA-MRSA of each continent shared a common genetic background with HA-MRSA or MSSA of other continents (Table 3). This suggests that intercontinental exchange of MRSA or MSSA had occurred, possibly followed by the introduction of the SCC\(\text{mec}\) in MSSA and the PVL locus in MSSA or MRSA. However, we cannot rule out the converse hypothesis that, for instance, MRSA of ST8 from France, which do not harbor the PVL locus and are of \(agr1\) allele, derive from an ancestor of \(agr1\) allele found in the United States, carrying the PVL locus (Table 3 and data not shown). In any case, the association of SCC\(\text{mec}\) IV (SCC\(\text{mec}\)IVa according to Baba et al. deno- mination [6]) with PVL in the CA-MRSA strains most likely did not result from co-acquisition of the two determinants on a single mobile genetic element because the two loci are widely separated on the \(S.\ aureus\) chromosome (6).

The CA-MRSA isolates contained the SCC\(\text{mec}\) type IV element, according to the designation of Oliveira and de Lencastre (15). If MRSA isolates with the SCC\(\text{mec}\) type IV element contained an additional 381–base pair band because of the integration of pUB110, the isolates are said to be SCC\(\text{mec}\) IVA. However, other groups in Japan and the United States have used region-specific primers to define the L-C region of the SCC\(\text{mec}\) element (4). On the basis of L-C polymorphism, researchers have identified three types to date, designated IVa, IVb, and IVc. Thus, the IVa of these latter groups (4) is not the same as the IVA of Oliveira and de Lencastre (15).

The exact nature of the selective advantage conferred by the observed combination of genetic traits remains to be elucidated, but simple antibiotic selection seems unlikely in a community context of widely different populations with various degrees of methicillin exposure. Okuma et al.

<table>
<thead>
<tr>
<th>Sequence type</th>
<th>Present study</th>
<th>Data from the MLST Web site</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Country of origin of CA-MRSA</td>
<td>n</td>
</tr>
<tr>
<td>1</td>
<td>USA</td>
<td>29</td>
</tr>
<tr>
<td>8</td>
<td>USA</td>
<td>3</td>
</tr>
<tr>
<td>30</td>
<td>Oceania (Southwest Pacific clone)</td>
<td>13</td>
</tr>
<tr>
<td>59</td>
<td>USA</td>
<td>1</td>
</tr>
<tr>
<td>80</td>
<td>France, Switzerland</td>
<td>67</td>
</tr>
<tr>
<td>93</td>
<td>Australia (Queensland clone)</td>
<td>4</td>
</tr>
</tbody>
</table>

MLST, multi-locus sequence typing; MRSA, methicillin-resistant \(S.\ aureus\); MSSA, methicillin-susceptible \(S.\ aureus\); CA, community acquired.
(4) suggested that CA-MRSA should display enhanced ecologic fitness, as they had a shorter doubling time than HA-MRSA. The real impact of this in vitro observation needs to be evaluated. Unlike other SCCmec elements, SCCmec IV and SCCmec I do not code for additional resistance determinants; however, SCCmec IV does code for mecA, a peptidoglycan transpeptidase. This protein is expressed at the external surface of the cytoplasmic membrane, where it could interact with the extracellular protein PVL. We are investigating the possibility of PVL activity or of peptidoglycan formation as a result of such an association.

CA-MRSA infections appear to be an emerging phenomenon worldwide. The PVL locus represents a stable genetic marker of these CA-MRSA strains, which explains the frequency of primary skin infections and occasionally necrotizing pneumonia associated with these strains (2,8,24,25). Although the selective advantage conferred by the combination of genetic traits (i.e., PVL locus and SCCmec IV in an agr3 background) is not clear, the spread of a limited number of clones in each continent suggests that these CA-MRSA strains are particularly suited to be successful community-based pathogens.

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References


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